

15 - 60 mg/kg) reduces psychotic-like behavioural effects in a manner comparable to that observed with atypical anti-psychotic drugs (696,697). Furthermore, one clinical study showed that pre-treatment of a small number of human subjects with CBD (5 mg i.v.), but not placebo, diminished the emergence of psychotic symptoms 30 min after i.v. administration of Δ^9 -THC (105). In contrast, a naturalistic study of cannabis users failed to show any differences in the prevalence of psychotic-like symptoms between subjects who reported smoking cannabis containing "low" or "high" levels of CBD; however the authors mention a number of confounding factors, including the lack of adjustment for alcohol consumption that could help explain this apparent inconsistency (656). An internet-based, cross-sectional study of 1 877 individuals who had a consistent history of cannabis use reported that individuals who had consumed cannabis with a higher CBD to THC ratio reported experiencing fewer psychotic episodes; however, the authors noted that the observed effects were subtle (113). Furthermore, the study was hampered by a number of important methodological issues suggesting the conclusions should be interpreted with caution. More recently, a four-week, double-blind, parallel-group, randomized, active-controlled clinical trial comparing CBD (200 mg, q.i.d., up to a total daily amount of 800 mg) to amisulpride (a dopamine D_2/D_3 receptor antagonist used in the treatment of schizophrenia) reported that both drugs were associated with a significant clinical improvement in symptoms with no significant difference between the two treatments (698). Treatment with CBD was well tolerated with significantly fewer side effects compared to those associated with anti-psychotic treatment (e.g. the presence of extra-pyramidal symptoms and lower prolactin release). In addition, CBD did not appear to significantly affect either hepatic or cardiac functions (698). Cannabidiol treatment, but not amisulpride, was also associated with an increase in serum levels of anandamide (698).

While there is some indication for a potential therapeutic role for CBD itself in the treatment of patients with pre-existing schizophrenia or psychosis or those who develop psychotic symptoms as a result of cannabis use, the extent to which CBD (at the levels typically found in cannabis) is able to ameliorate psychotic symptoms has not been firmly established and in fact, much of the cannabis consumed typically contains relatively low levels of CBD (60). For example, the CBD content of cannabis typically varies between 0.1 and 0.5%, although CBD levels of up to 8.8% (in hashish) have been noted (113). Therefore, a 1 g joint could contain between 1 mg (0.1%) and 88 mg (8.8%) of CBD—levels which are much lower than those usually administered in clinical trials (600 - 1500 mg/day) (699).

In conclusion, consumption of cannabis or other psychoactive cannabinoids (e.g. dronabinol, nabilone) should be treated with considerable caution in this patient population as these substances are believed to trigger psychotic episodes, lower the age of onset of symptoms, and contribute to a negative long-term prognosis in vulnerable individuals. Additionally, the therapeutic potential of CBD alone in the treatment of schizophrenia/psychosis, while promising, requires further study.

4.8.6 Alzheimer's disease and dementia

While still controversial, a widely accepted theory underlying the pathophysiology of Alzheimer's disease (AD) is the deposition of amyloid- β ($A\beta$) protein in specific brain regions leading to localized neuroinflammatory responses and accumulation of intra-cellular neurofibrillary tangles (composed of hyperphosphorylated tau protein); these events result in neuronal cell death with accompanying loss of functional synapses and changes in neurotransmitter levels (700). These pathological processes are thought to give rise to disease-associated symptoms such as memory deficits, and cognitive and motor impairments (700).

The endocannabinoid system and Alzheimer's disease

There is some evidence to suggest a role for the endocannabinoid system in the pathophysiology of AD (700,701). One *in vivo* study reported elevation in the levels of the endocannabinoid 2-arachidonoylglycerol (2-AG) in response to intra-cerebral administration of $A\beta_{1-42}$ peptide in animals (702). Another study using post-mortem brain samples from AD patients showed decreased anandamide levels with increasing $A\beta_{1-42}$ levels, but no association with $A\beta_{40}$ levels, amyloid plaque load, or tau protein phosphorylation (703).

Pre-clinical data

Pre-clinical studies suggest the endocannabinoid system protects against excitotoxicity, oxidative stress, and inflammation—all key pathological events associated with the development of AD (704). However, limited information exists regarding the use of cannabis or cannabinoids in the treatment of AD. Results from *in silico* and *in vitro* experiments suggest Δ^9 -THC could bind and competitively inhibit acetylcholinesterase (AChE), which in the context of AD functions as a molecular chaperone accelerating the formation of amyloid fibrils and

forming stable complexes with A β (705). Δ^9 -THC blocked the amyloidogenic effect of AChE, thereby diminishing A β aggregation (705). Other *in vitro* studies suggest that cannabidiol may have neuroprotective, antioxidant, and anti-apoptotic effects, as well as preventing tau protein hyperphosphorylation in cellular models of AD (706,707,708). Endocannabinoids have also been shown to prevent A β -induced lysosomal permeabilization and subsequent neuronal apoptosis *in vitro* (704). In pre-clinical animal models of AD, cannabidiol dose-dependently and significantly inhibited reactive gliosis and subsequent neuroinflammatory responses in A β -injected mice, at doses of 2.5 mg/kg/day and 10 mg/kg/day i.p., during a seven-day course of treatment (709). Another study using both *in vitro* and *in vivo* models of AD reported opposing roles for the CB₁ and CB₂ receptors in this context: CB₁ receptor agonism and CB₂ receptor antagonism were both associated with blunted A β -induced reactive astroglia and attenuation of neuroinflammatory marker expression (710).

Clinical data

There are very few clinical studies of cannabis or cannabinoids for the treatment of AD. One double-blind, placebo-controlled, six-week, crossover study of 12 patients suffering from Alzheimer-type dementia reported that 5 mg of dronabinol (Δ^9 -THC) daily was associated with a decrease in disturbed behaviour (410). However, adverse reactions such as fatigue, somnolence, and euphoria (presumably unwanted) were reported in dronabinol-treated patients. One open-label pilot study of six patients suggested an evening dose of 2.5 mg dronabinol (Δ^9 -THC) reduced nocturnal motor activity and agitation in those who were severely demented (711). In one case-report, a patient suffering from dementia of the Alzheimer-type who had been treated unsuccessfully with donepezil, memantine, gabapentin, trazodone, and citalopram was given nabilone (initially 0.5 mg at bedtime, and then twice per day) with immediate reduction in the severity of agitation and resistiveness and eventual improvement in various behavioural symptoms following six weeks of continuous treatment (712). It is unclear if the beneficial effects observed in these three studies are related to the non-specific sedative effects of Δ^9 -THC or nabilone, or to a specific cannabinoid-dependent therapeutic mechanism of action. It is also worth noting that one cross-sectional study reported that prolonged use of ingested or inhaled cannabis was associated with poorer performance on various cognitive domains (e.g. information processing speed, working memory, executive function, and visuospatial perception) in patients with multiple sclerosis (178). Similar adverse effects of cannabis/cannabinoids on cognition could potentially apply in the context of Alzheimer-type dementia.

A Cochrane database systematic review of cannabinoids for the treatment of dementia concluded that there was insufficient clinical evidence to suggest cannabinoids as being effective in the improvement of disturbed behavior in dementia or in the treatment of other symptoms of dementia (713).

4.8.7 Inflammation

The role of the endocannabinoid system in inflammation is complex as the endocannabinoid system has been implicated in both pro- and anti-inflammatory processes (701). Endocannabinoids, such as anandamide and 2-arachidonoylglycerol (2-AG), are known to be produced and released by activated immune cells and to act as immune cell chemoattractants promoting or directing the inflammatory response (714). On the other hand, cannabinoids can also suppress the production of pro-inflammatory cytokines and chemokines and thus may have therapeutic applications in diseases with an underlying inflammatory component (714,715). For information on other diseases with an inflammatory component such as the arthritides or inflammatory bowel disease, please consult sections 4.7 and 4.8.8.2, respectively, of this document.

4.8.7.1 Inflammatory skin diseases (dermatitis, psoriasis, pruritus)

The skin possesses an endocannabinoid system (41). CB₁ and CB₂ receptors are expressed in a number of skin cells including epidermal keratinocytes, cutaneous nerves and nerve fibres, sebaceous cells, myoepithelial cells of eccrine sweat glands, sweat gland ducts, mast cells, and macrophages (716). The endocannabinoid system and certain associated signaling pathways (e.g. PPAR γ , TRPV1) appear to regulate the balance between keratinocyte proliferation, differentiation, and apoptosis; together, these systems may play a role in cutaneous homeostasis but also in diseases such as psoriasis, which is characterized by keratinocyte proliferation and inflammation (41,717,718,719).

Pre-clinical and clinical studies

The results from pre-clinical studies on the role of cannabinoids in the modulation of cutaneous allergic reactions are mixed. Some studies suggest a protective role for certain cannabinoids, while others suggest an antagonistic role (reviewed in (41)). In clinical studies, experimentally-induced histamine-triggered pruritus was reduced by peripheral administration of the potent synthetic CB₁/CB₂ cannabinoid receptor agonist HU-

210, and the accompanying increases in skin blood flow and neurogenic mediated flare responses were attenuated (720). In another study, topically applied HU-210 significantly reduced the perception of localized pain in human subjects following locally restricted application of capsaicin to the skin, and reduced subsequent heat hyperalgesia and touch-evoked allodynia without any psychomimetic effects (721). On the other hand, there have also been some case-reports of contact urticaria following exposure to cannabis flowers, and extreme sensitization to Δ^9 -THC and cannabidiol has also been documented in an animal model of contact dermatitis (722,723). Therefore, while it is possible that some cannabinoids (e.g. HU-210) may have therapeutic value in the treatment of certain inflammatory skin conditions (such as psoriasis, pruritus, and dermatitis), it is also possible for some cannabinoids to trigger adverse skin reactions. Much further research is required in this area.

4.8.8 Gastrointestinal system disorders (irritable bowel syndrome, inflammatory bowel disease, hepatitis, pancreatitis, metabolic syndrome/obesity)

Historical and anecdotal reports suggest that cannabis has been used to treat a variety of gastrointestinal disorders (e.g. diarrhea, inflammation, and pain of gastrointestinal origin) (724,725,726).

The endocannabinoid system and gastrointestinal disorders

The expression of both the CB₁ and CB₂ receptors has been detected in the enteric nervous system (enteric sensory neurons, nerve fibers and terminals), whereas the human colonic epithelium, colonic epithelial cells lines, and stomach parietal cells appear to only express the CB₁ receptor (28,29). CB₂ receptor expression appears to be upregulated in sections of the colon in patients with inflammatory bowel disease (31). In contrast, the expression and localization of endocannabinoid synthesizing enzymes have not been well determined (31). However, studies in animals indicate that the endocannabinoid degradative enzymes FAAH and MAGL can be found in the gastrointestinal tract (31). For example, FAAH is expressed in the stomach and in the large and small intestines, and has also been localized to the cell bodies of the myenteric plexus (31). MAGL expression has been detected in the muscle and mucosal layers of the duodenum and the ileum, as well as in the proximal and distal colon, and in the nerve cell bodies and nerve fibers of the enteric nervous system (727). There also appears to be some regional variation in the levels of endocannabinoids in the gut; 2-arachidonoylglycerol (2-AG) appears to be more abundant in the ileum than the colon, whereas the opposite is true of anandamide (31). CB₁ and CB₂ receptors appear to be expressed in the pancreas (30), whereas the CB₁, but not the CB₂ receptor, is expressed in the liver under normal conditions (32,33).

Cannabinoids appear to have many functions in the digestive system including the inhibition of gastric acid production, gastrointestinal motility, and secretion and ion transport, and the attenuation of visceral sensation and inflammation (reviewed in (31)). Perturbations in the levels of various components of the endocannabinoid system have been noted in experimental models of gastrointestinal disorders, as well as in clinical studies (reviewed in (31)). The sections below summarize the information regarding the uses of cannabis and cannabinoids in the treatment of various disorders of the gastrointestinal system.

4.8.8.1 Irritable bowel syndrome

Irritable bowel syndrome (IBS) is the most common functional gastrointestinal disorder encountered in clinical medicine (728). It is a spectrum of disorders characterized by the presence of chronic abdominal pain and/or discomfort and alterations in bowel habits (728,729). Symptom patterns can be divided into diarrhea predominant (D-IBS), constipation predominant (C-IBS), and a mixed pattern (M-IBS) (729,730). While the pathophysiology of IBS remains unclear, the disorder is thought to be caused by dysregulation of the 'brain-gut axis' in response to psychological or environmental stressors or to physical stressors such as infection or inflammation, and is characterized by altered gut motility and visceral hypersensitivity (728,729). There is also some emerging evidence that suggests an association between genetic alterations in genes coding for certain endocannabinoid system proteins (e.g. *FAAH* and *CNRI*) and the pathophysiology of IBS (731,732,733).

Pre-clinical data

A few pre-clinical studies in animal models of IBS have been carried out to date. Two studies have employed mechanically-induced colorectal distension to trigger an acute visceral pain response in rodents as a model of IBS-associated visceral hypersensitivity. One study in rats showed that intra-peritoneal injection of different synthetic cannabinoid receptor agonists inhibited pain-related responses to experimentally-induced colorectal distension when administered *prior* to the experimental stimulus (734). Intravenous administration of

different synthetic cannabinoid receptor agonists also appeared to inhibit the overall pain-related responses to experimentally-induced colorectal distension in rats, as well as in mice, when administered *after* the experimental stimulus (735). In another study, subcutaneous administration of CB₁ or CB₂-selective agonists was reported to reduce the enhanced small intestinal transit observed in a mouse model of post-inflammatory IBS (736).

Clinical data with dronabinol

There are only a handful of clinical studies examining the effects of cannabinoids in human experimental models of IBS and in patients with IBS.

One double-blind, randomized, placebo-controlled, parallel-group study examined the effects of dronabinol on gastrointestinal transit, gastric volume, satiation, and post-prandial symptoms in a group of healthy volunteers (737). A 5 mg dose of dronabinol was associated with a significant delay in gastric emptying in female subjects, but not male subjects (737). No significant differences in either small bowel or colonic transit were observed between subjects administered dronabinol or placebo in any gender group (737). The 5 mg dose of dronabinol was used because a 7.5 mg dose caused intolerable side effects in more than half of the subjects (737). Adverse effects associated with the consumption of a 5 mg dose of dronabinol included dizziness/light-headedness, dry mouth, disturbed mental concentration, and nausea (737).

A subsequent double-blind, randomized, placebo-controlled, parallel-group study carried out by the same group investigated the effects of dronabinol on colonic sensory and motor functions of healthy human volunteers (738). Administration of a 7.5 mg dose of dronabinol significantly increased colonic compliance, especially in females, and reduced pre- and post-prandial phasic colonic motility and pressure (738). Colonic compliance is defined as the change in distensibility of the colon in response to a change in applied intracolonic pressure and it is used as a measure of colonic viscoelastic properties and as an indicator of colonic motor/contractile activity (738,739,740). Decreased compliance is typically associated with urgency and diarrhea, while increased compliance is typically associated with constipation (739,741). An increase in colonic compliance in this setting could indicate a return towards proper colonic function. In contrast to the results seen in the pre-clinical rodent studies, dronabinol increased the sensory rating of pain but did not affect the sensory rating of gas, or the thresholds for first sensation of either gas or pain during experimentally-induced random phasic distensions (738).

A double-blind, randomized, parallel-group study investigated the effects of escalating doses of dronabinol on colonic sensory and motor functions in a population of mostly female patients diagnosed with IBS according to Rome III criteria (IBS-C, IBS-D, or IBS-A (i.e. *alternating* between diarrhea and constipation)) (742). Only the highest dose of dronabinol tested (5 mg) was associated with a small, but statistically significant, increase in colonic compliance (742). Furthermore, the effect on colonic compliance appeared to be more pronounced in the IBS-D/A sub-group compared to IBS-C. No significant differences were observed on fasting or post-prandial colonic tone in response to dronabinol at any dose. However, the highest dose of dronabinol (5 mg) was associated with a significant reduction in the proximal left colon motility index, with a trend towards decreased colon motility indices (742). Treatment effects were significant on the proximal colon motility index in patients with IBS-D/A, but not in IBS-C, and only for the highest dose (742). Sensation thresholds and sensation scores for gas and pain during experimentally-induced ramp distensions did not differ significantly among the different treatment groups (742). The effects of genotype and dronabinol dose interaction on gas and pain sensation ratings, as well as on proximal fasting and distal fasting motility indices were also investigated. The results from these preliminary pharmacogenetic studies raise the possibility that the effects of dronabinol on colonic compliance and proximal colonic motility may be influenced by genetic variations in the *FAAH* and *CNR1* genes, but further studies are required to substantiate this hypothesis (742).

A subsequent double-blind, randomized, placebo-controlled, parallel-group study in a population of mostly female patients with IBS-D (Rome III criteria) further investigated gene-treatment interactions on colonic motility in this sub-set of IBS patients (743). Neither the 2.5 mg b.i.d. nor the 5 mg b.i.d. doses of dronabinol had any statistically significant effects on gastric, small bowel, or colonic transit (743). The effects on colonic transit were also examined as a function of genotype-by-treatment dose interaction. While treatment with dronabinol appeared to decrease colonic transit in subjects carrying the *CNR1* rs806378 C/T/T polymorphism, these effects were not statistically significant. Adverse effects were reported not to differ significantly between treatment groups.

4.8.8.2 Inflammatory bowel diseases (Crohn's disease, ulcerative colitis)

Inflammatory bowel diseases (IBD) include Crohn's disease and ulcerative colitis (744). Crohn's disease is characterized by patchy, intra-mural inflammation which may affect any part of the gastrointestinal tract (745). Symptoms include abdominal pain, diarrhea and weight loss as well as systemic symptoms of malaise, anorexia, and/or fever (745). Crohn's disease may cause intestinal obstruction due to strictures, fistulae, or abscesses (745). Ulcerative colitis is characterized by diffuse mucosal inflammation limited to the colon (745). Symptoms commonly include bloody diarrhea, colicky abdominal pain, urgency, or tenesmus (745). Both diseases are associated with an equivalent increased risk of colonic carcinoma (745).

The endocannabinoid system and IBD

Endocannabinoid system changes have been observed in the gastrointestinal tracts of experimental animal models of IBD, as well as in those of IBD patients (31,744). These changes include changes in the levels of endocannabinoids, cannabinoid receptors, and endocannabinoid synthesizing and degrading enzymes (28,31,744,746,747,748).

Pre-clinical data

Pre-clinical experiments in animal models of IBD suggest cannabinoids and endocannabinoids may limit intestinal inflammation and disease severity via activation of CB receptors (749,750,751,752,753,754).

Acute colitis

Mice bearing a genetic deletion of the CB₁ receptor had a stronger colonic inflammatory response (749) following rectal administration of dinitrobenzene sulfonic acid (DNBSA), an established method of inducing an acute colitis-like phenotype in mice (755). In contrast to wild-type mice, histological examination of the colons of CB₁ knockout mice treated with DNBSA revealed disruption of epithelial structure, with extensive hemorrhagic necrosis and neutrophil infiltration into the mucosa, and with acute inflammation extending into the sub-mucosa and muscle layer (749). Pharmacological blockade of the CB₁ receptor in wild-type mice produced similar effects accompanied by thickening of the bowel wall, inflammatory infiltrates, and an increase in lymphoid-follicle size associated with adherence to surrounding tissues (749). Furthermore, in contrast to CB₁ knockout mice, wild-type mice retained a significantly greater body weight following DNBSA treatment (749). Treatment of wild-type mice with the potent synthetic CB₁ and CB₂ receptor agonist HU-210, prior to and after DNBSA insult, significantly reduced the macroscopic colonic inflammatory response (749). Mice bearing a genetic deletion of the FAAH enzyme also displayed an attenuated inflammatory response to DNBSA compared to wild-type littermates (749).

An analogous study found that CB₁ and CB₂ receptor knockout mice and CB₁/CB₂ receptor double knockout mice showed increased extent of colonic inflammation, increased loss of crypt architecture, increased hyperemia/edema, and an increased degree of infiltration of inflammatory cells compared to wild-type mice following trinitrobenzene sulfonic acid (TNBSA)-induced acute colitis (753). All three knockout strains exhibited severe transmural colitis, with severe loss of epithelium, thickening of the bowel wall, and inflammatory infiltrates compared to wild-type mice (753). Genetic deletion of either or both CB receptors was also associated with significantly increased mRNA levels of various pro-inflammatory cytokines compared to wild-type mice in mice treated with TNBSA (753).

TNBSA-induced acute colitis in mice was associated with a significant upregulation of CB₂ receptor mRNA levels in the proximal and distal colons of treated mice (756). Intra-peritoneal administration of CB₂ receptor agonists, prior to and following TNBSA-induced colitis, was associated with a reduction in the macroscopic damage (e.g. reduced ulceration, reduction in colonic adhesions, and reduced colonic shortening) (756). Conversely, administration of a CB₂ receptor antagonist aggravated TNBSA-induced colitis (756).

Acute colitis and cannabidiol

Intra-peritoneal injection of cannabidiol (5 - 10 mg/kg) prior to DNBSA-induced acute colitis was associated with a significant attenuation of body weight loss caused by DNBSA (757). Cannabidiol (CBD) also reduced the wet weight/colon length ratio of inflamed colonic tissue, a marker of the severity and extent of the inflammatory response (757). Furthermore, CBD (5 - 10 mg/kg) significantly reduced macroscopic damage associated with DNBSA administration (mild edema, hyperemia, and small bowel adhesions) as well as microscopic damage (epithelium erosion, and mucosal and sub-mucosal infiltration of inflammatory cells

with edema) (757). Lastly, treatment with CBD significantly attenuated the observed increases in some biological markers associated with inflammation and oxidative stress, as well as attenuating the observed increases in the colonic levels of anandamide and 2-AG (757).

Another study reported that intra-peritoneal (10 mg/kg) or intra-rectal (20 mg/kg) pre-treatment with CBD, again administered *prior* to induction of colitis by TNBSA, caused a significant improvement of the colitis score and a decrease in the myeloperoxidase activity (a measure of neutrophil accumulation in colonic tissue) (758). No such differences were observed for orally administered CBD. Histological examination of colonic tissue further revealed decreased destruction of the epithelial lining, a reduction in colon thickness, and less infiltration of immunocytes compared to vehicle-treated mice (758). In contrast to the study by Borrelli (757), no differences in body weight were observed between vehicle-treated and CBD-treated mice that had developed colitis (758).

The effects of intra-peritoneal injections of THC, CBD, and a combination of THC and CBD on TNBSA-induced acute colitis in rats have been investigated (754). In one experiment, treatment with 10 mg/kg of THC alone, a combination of 5 mg/kg THC and 10 mg/kg CBD, a combination of 10 mg/kg THC and 10 mg/kg CBD, or sulfasalazine alone was associated with a statistically significant decrease in the macroscopic damage score (MDS) (754). The MDS is a linear scale measuring the extent of macroscopic damage to the colon and includes markers such as the presence or absence of hyperemia, ulceration, inflammation, adhesions, damage length, and diarrhea (754). Furthermore, treatment of rats (with experimentally-induced colitis) with CBD alone did not affect body weight. However, treatment with 5 or 20 mg/kg THC alone, or a combination of 10 mg/kg THC and 10 mg/kg CBD, resulted in a significant reduction of body weight gain in rats with experimentally-induced colitis in comparison with the vehicle group (754). Myeloperoxidase activity, a measure of inflammation, was significantly decreased in CBD-treated rats and in rats treated with 10 or 20 mg/kg THC, or 5 mg/kg THC and 10 mg/kg CBD (754). Treatment with 10 mg/kg CBD, 10 mg/kg THC, 10 mg/kg THC and 10 mg/kg CBD, or sulfasalazine alone was also associated with decreased disturbances in colonic motility resulting from TNBSA-induced colitis (754).

In a different experimental mouse model of acute colitis, the CB₁ receptor-selective agonist ACEA and the synthetic CB₂ receptor-selective agonist JWH-133, when injected intra-peritoneally prior to and after colonic insult, significantly reduced colon weight gain, colon shrinkage, colon inflammatory damage score, and diarrhea (751).

Inhibition of the 2-AG degrading enzyme monoacylglycerol lipase (MAGL) in mice by intra-peritoneal administration of a MAGL inhibitor *prior* to induction of acute colitis by TNBSA was associated with decreased macroscopic and histological colon alterations, as well as decreased colonic expression of pro-inflammatory cytokines (759). Inhibition of MAGL was also associated with a reduction in colitis-related systemic and central inflammation in the liver and the CNS (759). Co-administration of either CB₁ or CB₂ receptor-selective antagonists completely abolished the protective effect in the colon afforded by MAGL inhibition, and partially reversed the protective anti-inflammatory effects associated with MAGL inhibition in the liver (759).

Chronic colitis

Intra-peritoneal administration of the synthetic CB₂ receptor-specific agonist JWH-133 significantly attenuated colitis-associated body weight loss, inflammation, leukocyte infiltration, and tissue damage in a mouse model of spontaneous chronic colitis (760). This CB₂ receptor specific agonist also reduced T-cell proliferation, increased T-cell apoptosis, and increased the numbers of mucosal and systemic mast cells (760).

Ileitis

The effect of cannabichromene on inflammation-induced hypermotility in a mouse model of intestinal ileitis has been studied (761). Ileitis is characterized by disruption of the mucosa, infiltration of lymphocytes into the sub-mucosa, increased myeloperoxidase activity, and vascular permeability (761). Administration of cannabichromene (15 mg/kg i.p.) following croton oil-induced intestinal inflammation was associated with a decrease in the expression of CB₁ and CB₂ receptor mRNA in the jejunum, but not in the ileum (761). Cannabichromene did not affect upper gastrointestinal transit, colonic propulsion, or whole gut transit in untreated mice, but did reduce intestinal motility in croton oil-treated mice at 10 and 20 mg/kg i.p. (761). Cannabichromene also dose-dependently and significantly inhibited contractions induced by acetylcholine, as

well as electrical field stimulation, *in vitro* in ilea isolated from control mice and croton oil-treated mice (761). The inhibitory effect of cannabichromene appeared to be cannabinoid receptor-independent (761).

Clinical studies with THC

A double-blind, randomized, placebo-controlled, crossover study examining the effects of 5 and 10 mg Δ^9 -THC in visceral sensitivity reported that Δ^9 -THC did not alter baseline rectal perception to experimentally-induced distension or sensory thresholds of discomfort after sigmoid stimulation compared to placebo, in either healthy controls or IBD patients (762). However, the authors did note a bias in the patient selection criteria which could have explained the apparent lack of effect.

Surveys and clinical studies with cannabis

Findings from a cross-sectional survey of 291 patients with IBD (Crohn's disease or ulcerative colitis) suggested that the vast majority of those patients reported using cannabis to relieve abdominal pain and to improve appetite (157). In contrast to patients with Crohn's disease, a greater proportion of patients with ulcerative colitis reported using cannabis to improve diarrheal symptoms (157). In general, patients reported being more likely to use cannabis for symptom relief if they had a history of abdominal surgery, chronic analgesic use, alternative/complementary medicine use, and a lower SIBDQ (short inflammatory bowel disease questionnaire) score (157). Both ulcerative colitis and Crohn's disease patients reported using cannabis to improve stress levels and sleep (157). The mean duration of cannabis use (current or previous) was seven years. The majority of cannabis users reported using once per month or less, but 16% reported using cannabis daily or several times per day (157). The vast majority (77%) of users reported smoking the cannabis as a joint without tobacco, 18% of users smoked it with tobacco, 3% used a water pipe, and 1% reported oral ingestion (157). Approximately one-third of patients in this study reported significant side effects associated with the use of cannabis such as paranoia, anxiety, and palpitations. Other commonly reported side effects included feeling "high", dry mouth, drowsiness, memory loss, hallucinations, and depression (157).

A retrospective, observational study of 30 patients with Crohn's disease examined disease activity, use of medication, need for surgery, and hospitalization before and after cannabis use (248). The average duration of disease was 11 years (range: 1 - 41 years). Twenty patients suffered from inflammation of the terminal ileum, five had inflammation of the proximal ileum, and eight had Crohn's disease of the colon. The indication for cannabis was lack of response to conventional treatment in the majority of the patients, and chronic intractable pain in most of the other patients (248). Most patients smoked cannabis as joints (0.5 g cannabis/joint), a few inhaled the smoke through water, and one patient consumed cannabis orally (248). Of those who smoked cannabis, most smoked between one and three joints per day. One patient smoked seven joints per day. The average duration of cannabis use was two years (range: 2 months - 9 years). All patients reported that consuming cannabis had a positive effect on their disease activity (248). The scores on the Harvey-Bradshaw index (an index of Crohn's disease activity) were significantly decreased following cannabis use, and the use of other medications (e.g. 5-ASA, corticosteroids, thiopurine, methotrexate, and TNF antagonist) also appeared to be significantly reduced following use of cannabis (248). The study was limited by design and small size.

A preliminary, observational, open-label, prospective, single-arm trial in a group of 13 patients suffering from Crohn's disease or ulcerative colitis reported that treatment with inhaled cannabis over a three-month period improved subjects' quality of life, caused a statistically significant increase in subjects' weight, and improved the clinical disease activity index in patients with Crohn's disease (189). Patients reported a statistically significant improvement in their perception of their general health status, their ability to perform daily activities, and their ability to maintain a social life (189). Patients also reported a statistically significant reduction in physical pain, as well as improvement in mental distress (189). No serious adverse events were noted. Study limitations included study design, subject selection bias, the lack of a proper control group and placebo, small number of subjects, and the inability to establish a dose-response effect (189).

Note: for sections 4.8.8.3, 4.8.8.4, and 4.8.8.5 below, no clinical studies examining the role of cannabis in the treatment of these disorders have been carried out to date.

4.8.8.3 Diseases of the liver (hepatitis, fibrosis, steatosis, ischemia-reperfusion injury, hepatic encephalopathy)

CB₁ receptors are expressed at low levels in the whole liver, hepatocytes, stellate cells, and hepatic vascular endothelial cells, but increased CB₁ receptor expression has been detected in the context of diseases such as hepatocellular carcinoma and primary biliary cirrhosis (reviewed in (763)). CB₂ receptors are undetectable in normal liver but, like the CB₁ receptors, they are upregulated in pathological conditions; these include non-alcoholic fatty liver disease (NAFLD), liver fibrosis, regenerating liver, and hepatocellular carcinoma (reviewed in (763)). Increases in the concentrations of the endocannabinoids anandamide and 2-AG in the liver appear to vary depending on the pathophysiological condition in question (33).

Steatosis and fibrosis

Mounting evidence suggests an important role for the endocannabinoid system in the pathophysiology of a multitude of diseases affecting the liver (33). In general, the CB₁ and CB₂ receptors appear to play *opposing* roles in the liver: activation of the CB₁ receptors is implicated in the progression and worsening of alcoholic and metabolic steatosis, liver fibrogenesis, and circulatory failure associated with cirrhosis; stimulation of the CB₂ receptors, in general, appears to confer beneficial effects in alcoholic fatty liver, hepatic inflammation, liver injury, liver regeneration, and fibrosis (reviewed in (33) and see also (249,250,251,764)). Conversely, antagonism of the CB₁ receptor appears to attenuate liver fibrosis in animal models by interfering with the production of several pro-fibrotic, pro-inflammatory, as well as anti-inflammatory mediators secreted in the liver during chronic liver injury and the wound healing process (249,765).

In vitro studies indicate that CBD may also play a protective role in attenuating liver fibrosis induced by acute liver injury or by chronic alcohol exposure (766). CBD dose-dependently triggered the apoptosis of cultured, activated hepatic stellate cells isolated from the livers of rats chronically exposed to an ethanol diet (766). The activation of hepatic stellate cells in response to liver injury is considered a key cellular event underlying hepatic fibrogenesis (766). Furthermore, CBD dose-dependently promoted the selective apoptosis of activated hepatic stellate cells, but not control hepatic stellate cells or primary hepatocytes, by triggering an endoplasmic reticulum-associated cellular stress response leading to apoptosis; this effect was independent of CB receptor activation (766).

Ischemia-reperfusion injury and hepatic encephalopathy

Pre-clinical studies also indicate a protective role for CBD in hepatic ischemia/reperfusion injury, and hepatic encephalopathy, in mice and rats (767,768,769). Pre-treatment of mice with 3 or 10 mg/kg body weight CBD (i.p.), 2 h before induction of ischemia-reperfusion in liver, dose-dependently attenuated serum transaminase elevations at 2 and 6 h of reperfusion compared to vehicle (767). CBD administered immediately following the induction of ischemia, or at 90 min of reperfusion, still attenuated hepatic injury measured at 6 h of reperfusion, though to a lesser extent than when administered prior to the induction of the ischemia-reperfusion injury (767). Pre-treatment with CBD also significantly reduced the signs of coagulation necrosis observed 24 h after ischemia-reperfusion, significantly attenuated hepatic cell apoptosis, significantly decreased the expression of pro-inflammatory chemokines and cytokines, attenuated neutrophil infiltration into the injury site, and decreased the expression of markers of tissue and cellular injury (767). Similar beneficial findings in a rat model of ischemia-reperfusion injury were reported in a different study; however, CBD (5 mg/kg, i.v.) was administered *after* ischemia-reperfusion injury (768). CBD treatment resulted in significant reductions in serum transaminase levels, hepatic lipid peroxidation, and the attenuation of various markers of tissue or cellular injury associated with ischemia-reperfusion (768). Administration of Δ^8 -tetrahydrocannabivarin (3 or 10 mg/kg, i.p.) 2 h *before* induction of hepatic ischemia-reperfusion injury dose-dependently attenuated serum transaminase elevations at 2 and 6 h of reperfusion compared to vehicle (770). Administration of Δ^8 -tetrahydrocannabivarin *post-ischemia* attenuated, although to a lesser degree, the hepatic injury measured at 6 h of reperfusion (770). Pre-treatment with Δ^8 -tetrahydrocannabivarin also significantly reduced the extent of coagulation necrosis in the liver, attenuated neutrophil infiltration, decreased the expression of hepatic pro-inflammatory chemokines and cytokines, reduced the hepatic levels of markers of oxidative stress, and decreased the extent of hepatocyte cell death following ischemia-reperfusion injury (770).

Intra-peritoneal administration of CBD (5 mg/kg, i.p.) improved neurological, locomotor, and cognitive functions in a mouse model of fulminant hepatic encephalopathy (769). CBD also attenuated the degree of astrogliosis, but did not affect the extent and severity of necrotic lesions in the liver (769). CBD partially restored whole brain 5-HT levels, as well as the levels of markers of liver function (ammonia, bilirubin, AST, ALT) in affected mice (769).

4.8.8.4 Metabolic syndrome, obesity, diabetes

The endocannabinoid system and energy metabolism

Increasing evidence suggests an important role for the endocannabinoid system in the regulation of energy balance; dysregulation of the system is associated with the development of metabolic syndrome and obesity, and may also increase the risk of developing atherosclerosis and type-2 diabetes (11,17,771). Pre-clinical studies carried out in animal models of obesity and clinical studies performed in obese humans report increased endocannabinoid tone in adipose tissue, liver, pancreas, and in the hypothalamus compared to controls (772).

The regulation of energy balance by the endocannabinoid system appears to occur both centrally (in the CNS, particularly in the hypothalamus) and peripherally in multiple organs such as the white adipose tissue, skeletal muscle, pancreas, liver, and small intestine (11,17,771,773). In general, overactivity of the endocannabinoid system is associated with increased nutrient intake, enhanced energy storage, and reduced energy expenditure (17). Endocannabinoid tone appears to be modulated by hormones and peptides including leptin, insulin, ghrelin, and corticosteroids (17). Endocannabinoids, in turn, appear to modulate the release of neurotransmitters and neuropeptides such as opioids, serotonin, and GABA, which are known to play a role in regulating appetite mainly through central mechanisms (774).

Pre-clinical data

THC and the role of the CB₁ receptor

In pre-clinical *in vitro* studies, THC significantly inhibited basal and catecholamine-triggered lipolysis in a differentiated mouse adipocyte cell line in a concentration-dependent manner and caused dose-dependent accumulation of lipid droplets in these cells (23). In mice, activation of the CB₁ receptor resulted in increased *de novo* fatty acid synthesis in the liver and increased formation and storage of triglycerides in the adipose tissue (11,775,776,777). In rats, central stimulation of the CB₁ receptor was associated with the development of hepatic and adipose tissue insulin resistance (772). Mice lacking overall CB₁ receptor gene expression were hypophagic and were leaner than wild-type mice regardless of diet, had lower plasma insulin levels, did not develop diet-induced insulin resistance or obesity, and had enhanced leptin sensitivity (391,775,778). In mice, targeted deletion of the CB₁ receptor in the forebrain-projecting neurons in the hypothalamus and in the nucleus of the solitary tract, and partial deletion in sympathetic neurons were associated with a lean phenotype and resistance to diet-induced obesity and increases in plasma levels of leptin, insulin, glucose, free fatty acids, and triglycerides; these effects resulted from an increase in lipid oxidation and thermogenesis as a consequence of enhanced sympathetic tone and a decrease in energy absorption (779). Similarly, partial targeted deletion of the CB₁ receptor gene in the adult mouse hypothalamus lead to a significant decrease in body weight gain triggered by an increase in energy expenditure, rather than a decrease in food intake (777).

Targeted deletion of the CB₁ receptor gene in mouse liver is associated with the development of diet-induced obesity, but retention of glucose, insulin and leptin sensitivity and lipid indices; targeted hepatic re-expression of the CB₁ receptor gene in CB₁ receptor gene knockout mice was associated with glucose intolerance and insulin resistance in response to a high-fat diet, but maintenance of proper body weight (780,781). Studies with CB₁ antagonists/inverse agonists strongly suggest that antagonism/inverse agonism at the CB₁ receptor is associated with reduced caloric intake, weight loss, improvement or reversal of hepatic steatosis, and restoration of insulin and glucose sensitivity and normal lipid indices in various animal models of diet-induced obesity (391,782,783,784,785,786,787,788). Clinical studies with the CB₁ antagonist rimonabant have strongly supported the data gathered from animal studies (789,790,791,792,793,794,795).

Taken together, the above findings suggest an important role for the CB₁ receptor, both centrally and

peripherally, in regulating energy balance; stimulation of the CB₁ receptor promotes energy storage and lipogenesis, whereas CB₁ receptor antagonism has the opposite effects. Consistent with these findings, cannabis and prescription cannabinoids (dronabinol, nabilone) are known to increase appetite and body weight and have been used clinically to treat HIV/AIDS-associated anorexia-cachexia, and possibly also cancer-associated cachexia (see sections 4.3.1 and 4.3.2, respectively). Yet curiously, despite these beneficial effects on body weight in clinical disorders, a number of studies have so far failed to find an association between overweight/obesity and consumption of cannabis in the general population (796,797). In fact, the prevalence of obesity appeared to be significantly lower in cannabis users than in non-users, and the proportion of obese individuals also appeared to decrease with frequency of cannabis use according to a cross-sectional analysis of two U.S. epidemiological studies (797).

Role of the CB₂ receptor

The CB₂ receptor also appears to also play an important role in energy balance (798). Pre-clinical studies in mice indicate that the CB₂ receptor is expressed in epididymal adipose tissue in lean mice, and the levels of this receptor appear to increase in the non-parenchymal cell fractions of adipose tissue and liver in genetically obese mice or in wild-type mice fed a high-fat diet (798). Furthermore, systemic administration of a CB₂ receptor-selective agonist to lean or obese mice, or exposure of cultured fat pads to the same agonist, was associated with upregulation of a subset of genes linked to inflammation in the adipose tissue but not the liver (798). Conversely, administration of a CB₂-selective antagonist reduced inflammation both in adipose tissue and liver of obese animals (798). Under a high-fat diet, mice lacking the CB₂ receptor displayed a slower body weight progression and were more insulin sensitive than wild-type mice (798). CB₂ knockout mice on a high-fat diet also exhibited minimal hepatic steatosis compared to wild-type mice (798). Mice deficient in CB₂ receptor expression also exhibited increased food intake and body weight with age compared to wild-type mice (799). The CB₂ receptor knockout mice did not develop insulin resistance and showed enhanced insulin-stimulated glucose uptake in skeletal muscle (799). Taken together, these results suggest an important and complex role for the CB₂ receptor in energy balance and obesity, although further studies are needed to better understand its role.

Other cannabinoids

Pure Δ^9 -tetrahydrocannabivarin (THCV) administered i.p. (3 mg/kg, 10 mg/kg, or 30 mg/kg) in mice suppressed feeding and significantly reduced body weight gain, but this effect appeared to be blocked when a botanical extract containing both Δ^9 -THCV and Δ^9 -THC was used (92). Inclusion of cannabidiol into the botanical extract, as a way of attenuating the proposed hyperphagic effects of THC in this study, resulted in a trend towards decreased food intake in treated mice, but the effect did not reach statistical significance (92). Lean and obese rats injected with a cannabis extract (on alternate days, for 28 days) containing a THC : CBN : CBD ratio of 1.0 : 1.2 : 0.4 (5 mg/kg Δ^9 -THC) exhibited a significant reduction in weight gain during the study period, but the cannabis extract treatment was not associated with any changes in either insulin or glucose levels (800).

4.8.8.5 Diseases of the pancreas (diabetes, pancreatitis)

Although there appears to be a general lack of consensus as well as insufficient information regarding the exact expression, distribution, and function of the various endocannabinoid system components in the pancreas among different species, the pancreas does appear to have at least some, and in certain cases many, of the individual elements of the endocannabinoid system (774,801,802).

Function of the endocannabinoid system in the pancreas

Two studies using primary human islet cells suggest that the CB₁ and CB₂ receptors are expressed in these cells, and that stimulation of the CB₁ receptor is associated with secretion of insulin and glucagon while stimulation of the CB₂ receptor is associated with either increased or decreased insulin secretion (801,803) (and also reviewed in (774)). More recently, the endocannabinoid 2-arachidonoylglycerol (2-AG) has been implicated in the regulation of both insulin and glucagon secretion in human pancreas (802).

Intra-muscular administration of cannabis resin (containing 6.3% Δ^9 -THC, 3.2% cannabidiol, and 1.9% cannabinol) at increasing doses (Δ^9 -THC at 2.5, 5.0, and 10 mg/kg) to dogs was associated with a progressive increase in plasma glucose levels which reached maximum values 90 min after administration, with a return to baseline values 180 min after administration (804). Injection of anandamide or a CB₁ receptor-selective agonist in rats was associated with acute glucose intolerance, whereas administration of a CB₁ receptor

inverse agonist attenuated this effect (805). In humans, intravenous injection of 6 mg of Δ^9 -THC to healthy, non-obese male volunteers was associated with acute impairment of glucose tolerance in response to glucose challenge with no change in plasma insulin levels (806).

Survey data

A cross-sectional study of 10 896 adults, ages 20 - 59, who were participants in the National Health and Nutrition Examination Survey III (NHANES), a nationally representative sample of the U.S. population, reported that cannabis use was independently associated with a decreased prevalence of diabetes mellitus, and that cannabis users had lower odds of developing diabetes mellitus compared to non-users (807). The lowest prevalence of diabetes mellitus was seen in current, light cannabis users, but current heavy users and past users also had a lower prevalence of diabetes mellitus than non-cannabis users (807). Due to limitations in study methodology (e.g. cross-sectional nature of the study, self-report bias, and inconsistent sampling methodology) as well as the possibility of additional and uncontrolled confounding factors, the authors indicate that it is not yet possible to conclude that cannabis use does not lead to diabetes mellitus, nor that cannabis should be considered a treatment for this disorder (807).

Cannabis, the endocannabinoid system, and acute and chronic pancreatitis

Acute, heavy cannabis use has been linked to the development of acute pancreatitis (253,254,255,256). Acute pancreatitis is a potentially lethal disorder involving inflammation, cell death, and complex neuroimmune interactions; the management of chronic pancreatitis remains clinically challenging with no definite cure and supportive measures are the only treatment available (808,809). Pancreatic tissue isolated from patients with *acute* pancreatitis has been reported to have a marked upregulation of CB₁ and CB₂ receptors in the acini and ducts as well as elevated levels of the endocannabinoid anandamide but not 2-AG (808). In a subsequent study, an increase in the expression levels of CB₁ and CB₂ receptors, and a decrease in the levels of endocannabinoids (anandamide and 2-AG) were noted in tissue samples isolated from patients suffering from *chronic* pancreatitis compared to pancreatic tissues isolated from healthy subjects (809). In addition, in contrast to the findings obtained for acute pancreatitis (808), tissues isolated from patients with chronic pancreatitis appeared to have decreased levels of anandamide and 2-AG (809). Activation of CB₁ and CB₂ receptors in chronic pancreatitis-derived pancreatic stellate cells was also associated with the induction of a quiescent-cell phenotype as well as the downregulation of extracellular matrix protein production and inflammatory cytokine production (809).

Pre-clinical data and acute or chronic pancreatitis

There are only a handful of reports on the effects of cannabinoids in experimental animal models of acute or chronic pancreatitis, and the findings from these reports are conflicting. Thus, the use of cannabinoids in the treatment of acute or chronic pancreatitis remains unclear. Information gathered from pre-clinical animal studies is summarized below.

Elevations in the plasma levels of anandamide have been noted in a rat model of severe acute pancreatitis (810), and administration of the CB₁ receptor antagonist AM251 after induction of pancreatitis appeared to improve the course of the disease (810). In another study, administration of anandamide *prior* to induction of pancreatic damage further aggravated the usual course of the disease, whereas pre-treatment with the CB₁ receptor antagonist AM251 prevented the development of cerulein-induced pancreatitis and when administered *after* injury also appeared to reverse cerulein-induced pancreatic damage (811). Similarly, mice treated with the CB₁ receptor antagonist rimonabant *prior* to cerulein-induced pancreatitis exhibited significantly decreased pancreatic damage as well as decreased production of inflammatory cytokines (812). Subcutaneous administration of a synthetic CB₁/CB₂ receptor agonist, both prior to as well as after induction of acute pancreatitis in mice, attenuated the abdominal pain, inflammation, and tissue pathology associated with pancreatitis (808). In contrast, a different study reported that pre-treatment of rats with a synthetic CB₁/CB₂ receptor agonist *before* induction of experimentally-induced pancreatitis attenuated the extent of tissue damage and the release of inflammatory cytokines, whereas administration of the same agonist *after* the induction of pancreatitis had the opposite effects and appeared to aggravate the course of the disease (813). These contradictory findings may be due to differences in experimental methods, differences in timing of drug administration, differences in the types of agonists and antagonists that were used, differences in the route of administration, and differences in animal species.

4.8.9 Anti-neoplastic properties

A number of studies have implicated the endocannabinoid system in the pathophysiology of cancer. In general, endocannabinoids seem to have a protective effect against carcinogenesis, and proper regulation of local endocannabinoid tone is likely an important factor in controlling the malignancy of different cancers (814). When compared with healthy tissues, the levels of endocannabinoids appear to be elevated in glioblastomas, meningiomas, pituitary adenomas, prostate and colon carcinomas, and endometrial sarcomas (746,815,816,817,818,819). The expression levels of cannabinoid receptors are also differentially regulated in normal versus malignant cells, with increased or decreased levels of these receptors varying with cancer type (reviewed in (814)). Such differences in the levels of endocannabinoids and in the patterns of expression levels of cannabinoid receptors across different cancer types reflect the complex role of the endocannabinoid system in cancer and are likely to pose challenges to potential therapeutic approaches. Nonetheless, a number of pre-clinical studies have shown that endocannabinoids, certain synthetic cannabinoid agonists, and some phytocannabinoids can inhibit tumour growth and progression of numerous types of cancers through various mechanisms including promotion of apoptosis, cell-cycle arrest/growth inhibition, and prevention of metastasis through inhibition of tumour invasion, migration, and neo-angiogenesis (reviewed in (814,820)).

In general, the anti-neoplastic effects of Δ^9 -THC appear to be biphasic: lower doses (under 100 nM), comparable to those typically seen in clinical or therapeutic settings, are considered pro-proliferative; higher doses (above 100 nM) are thought to be anti-proliferative (821), although exceptions have been noted. Furthermore, cannabinoid concentrations above 100 nM, that is two orders of magnitude above the average affinity of these receptors for cannabinoids, are likely to produce off-target, CB receptor-independent effects (822). As a point of reference, single oral doses of dronabinol (Δ^9 -THC) of 2.5, 5, and 10 mg have been associated with mean peak Δ^9 -THC plasma concentrations of 0.65, 1.83, and 6.22 ng/mL, respectively (174). These concentrations correspond to concentrations of 0.002, 0.006, and 0.02 μ M (or 2, 6, and 20 nM) Δ^9 -THC. Doubling of these daily oral doses is associated with mean peak Δ^9 -THC plasma concentrations of 1.3, 2.9, and 7.9 ng/mL Δ^9 -THC (174), respectively, corresponding to 0.004, 0.009, and 0.03 μ M (or 4, 9, and 30 nM) Δ^9 -THC. Continuous dosing for seven days with 20 mg doses of dronabinol (total daily doses of 40 - 120 mg dronabinol) gave mean plasma Δ^9 -THC concentrations of ~20 ng/mL or ~0.06 μ M (60 nM) Δ^9 -THC (288). Smoking a 1 g joint containing 12.5% Δ^9 -THC can be assumed, based on the literature, to yield peak plasma Δ^9 -THC concentrations between 50 and 100 ng/mL or more (see section 3.1 "Smoking", subsection "Plasma concentrations of Δ^9 -THC following smoking"). Such Δ^9 -THC plasma concentrations correspond to 0.16 and 0.32 μ M (or 160 and 320 nM) Δ^9 -THC, respectively. Plasma concentrations of Δ^9 -THC are known to vary widely across individuals, and diminish more rapidly by the smoking route than by oral administration. With respect to doses expressed in mg/kg of body weight, a daily oral dose of 2.5 mg of dronabinol (Δ^9 -THC) can be estimated to correspond to a dose of approximately 0.04 mg/kg (assuming a body weight of 70 kg), whereas a daily oral dose of 40 mg of dronabinol would correspond to a dose of approximately 0.6 mg/kg of dronabinol. Smoking a 1 g joint containing 12.5% Δ^9 -THC would correspond to a hypothetical dose of 1.8 mg/kg Δ^9 -THC.

The following paragraphs summarize the main findings from a number of pre-clinical *in vitro* and *in vivo* studies of cannabinoids in neoplastic diseases. Clinical data are presented at the end of this section.

Pre-clinical data

In vitro studies suggest that Δ^9 -THC decreases cell proliferation and increases cell death in human glioblastoma multiforme cell lines, with CB receptor activation accounting for only part of the observed effects (823). In the case of astrocytomas, higher concentrations were deemed to be clinically preferable because this would bypass CB receptor activation and induce apoptosis in all astrocytoma cell sub-populations (824). In the case of breast cancer, Δ^9 -THC reduced human breast cancer cell proliferation at concentrations of 4 - 10 μ M (i.e. 4 000 - 10 000 nM), with more aggressive estrogen receptor-negative tumour cells being more sensitive to the effects of THC (825). In contradistinction, another study showed that Δ^9 -THC (50 μ M (i.e. 50 000 nM) *in vitro* or 50 mg/kg *in vivo*) enhanced breast cancer growth and metastasis (826). Furthermore, Δ^9 -THC, CBD, and CBN all stimulated breast cancer cell proliferation at concentrations ranging from 5 - 20 μ M (i.e. 5 000 - 20 000 nM) (827), but this effect appeared to depend to some extent on the hormonal milieu (with lower estrogen levels promoting, and higher estrogen levels inhibiting growth). On the other hand, cannabinoids such as cannabigerol, cannabichromene, cannabidiolic acid, and THC acid as well as cannabinoid-based extracts enriched in either Δ^9 -THC or CBD inhibited cell proliferation (in the micromolar range) in a number of different breast cancer cell lines (828). In *in vitro* studies examining the role of cannabinoids in lung cancer, Δ^9 -THC (10 - 15 μ M) (i.e. 10 000 - 15 000 nM) attenuated growth factor-induced migration and invasion of non-small cell lung cancer cell lines (829). In the case of colorectal cancer, Δ^9 -THC at concentrations of 2.5 μ M (i.e. 2 500 nM) and above (range: 7.5

- 12.5 μM) (i.e. 7 500 – 12 500 nM) were associated with a decrease in colorectal cancer cell survival, whereas lower concentrations (100 nM - 1 μM) had no effect (830). Taken together, these and other *in vitro* studies suggest cannabinoids can have complex biological effects in the context of malignancies. Differences in experimental conditions, cancer cell type, CB-receptor expression, hormonal levels, and the existence of CB-receptor dependent and independent regulatory mechanisms all appear to affect the control of growth, proliferation, and invasion of cancer cells in response to cannabinoids. Furthermore, these findings also suggest that the effective inhibitory concentrations of Δ^9 -THC seen *in vitro* are between ~ 10 and 7 500 times higher than the concentrations of Δ^9 -THC seen clinically, depending on the route of administration.

A pre-clinical *in vivo* study in rats showed that intra-tumoural administration of Δ^9 -THC caused significant regression of intra-cranial malignant gliomas, and an accompanying increase in animal survival time without any neurotoxicity to healthy tissues (831). Furthermore, no substantial change was observed in certain behavioural measures suggesting that the effect of Δ^9 -THC was limited to diseased neural tissues (831). Other studies showed that peritumoural administration of 0.5 mg Δ^9 -THC /day, twice per week, for 90 days, significantly slowed focal breast tumour growth, blocked tumour generation, decreased total tumour burden, delayed the appearance of subsequent tumours, and impaired tumour vascularization in the ErbB2-positive metastatic breast cancer mouse model (832). Δ^9 -THC, at doses of 5 mg/kg/day, administered intra-peritoneally or intra-tumourally also dramatically decreased the growth and metastasis as well as the vascularization of xenografted non-small cell lung cancer cell lines in immunodeficient mice (829). CBD (5 mg/kg) or CBD-rich extract (6.5 mg/kg) administered intra-tumourally or intra-peritoneally, twice per week, to breast-cancer-cell-xenografted athymic mice significantly decreased both tumour volume and the number of metastatic nodules (828). Other investigators showed that intra-peritoneal administration of CBD at 1 or 5 mg/kg/day significantly reduced the growth and metastasis of an aggressive breast cancer cell line in immune-competent mice (833). Importantly, the primary tumour acquired resistance to the inhibitory properties of CBD by day 25 of treatment (833). Taken together, these studies suggest that cannabinoids such as Δ^9 -THC and CBD can, under a specific set of circumstances, have anti-neoplastic effects in various animal models of cancer at certain doses or concentration ranges.

Combining cannabinoids with other chemotherapeutic agents

Pre-clinical *in vitro* and *in vivo* studies investigating the effects of combining cannabinoids with frequently used chemotherapeutic agents have also been performed. One *in vitro* study showed that combining sub-maximal doses of Δ^9 -THC (0.75 μM) with cisplatin or doxorubicin reduced the viability of an astrocytoma cell line in a synergistic manner (834). Likewise, combining sub-maximal doses of Δ^9 -THC with temozolomide reduced the viability of several human glioma cell lines and primary cultures of glioma cells derived from human glioblastoma multiforme biopsies *in vitro* (835). Complementing these findings, an *in vivo* study showed that combined treatment with Δ^9 -THC (15 mg/kg/day) and temozolomide (5 mg/kg/day) reduced the growth of glioma tumour xenografts in mice in a synergistic manner (835).

Clinical data

There is only one report of a clinical study of Δ^9 -THC to treat cancer (836). In this non-placebo controlled pilot study, nine patients with glioblastoma multiforme who had failed standard surgical and radiation therapy, had clear evidence of tumour progression, and had a minimum Karnofsky score of 60 were treated with 20 - 40 μg Δ^9 -THC intra-tumourally per day (with doses of up to 80 - 180 μg Δ^9 -THC per day). Median treatment duration was 15 days (836). While intra-tumoural administration of Δ^9 -THC appeared to be well tolerated, the effect of Δ^9 -THC on patient survival was not significantly different from that observed in other studies using chemotherapeutic agents such as temozolomide or carmustine (837,838). Nevertheless, *in vitro*, Δ^9 -THC inhibited the proliferation and decreased the viability of tumour cells isolated from glioblastoma biopsies, most likely through a combination of cell-cycle arrest and apoptosis (836,839). In addition, results from a separate *in vitro* study suggest that CBD enhanced the inhibitory effects of Δ^9 -THC on human glioblastoma cell proliferation and survival (839).

Despite the evidence presented in these and other studies, there is a general consensus that Δ^9 -THC would not be considered the most appropriate CB agonist in anti-tumoural strategies, especially if administered systemically, because of its high hydrophobicity, relatively low agonist potency, and its well-known psychoactive properties (814,840,841). Much remains to be known regarding factors such as the expression levels of the cannabinoid receptors in different cancers, the effects of different cannabinoids on different cancer cell types, the identification of factors that confer resistance to cannabinoid treatment, as well as the most efficient approaches for enhancing cannabinoid anti-tumoural activity whether alone or in combination with other therapies (828,840). Furthermore, the apparent biphasic effect of cannabinoids further highlights the need for more comprehensive dose-response studies (842).

4.8.10 Emerging Potential Therapeutic Uses

There are a few pre-clinical reports which suggest that administration of a low dose of THC, a CB₁ receptor antagonist, or a CB₂ receptor agonist may reduce the progression of atherosclerosis in mouse models of the disease (843,844,845). Oral administration of THC (1 mg/kg/day) has been associated with significant inhibition of disease progression in the apolipoprotein E (ApoE) knockout mouse, a mouse model of atherosclerosis (843). The beneficial effect of THC in this study was mediated by the CB₂ receptor, likely through its inhibitory effects on immune system cells (macrophages and T-cells) located in or near atherosclerotic lesions (843). These findings were supported by another study which showed that intra-peritoneal administration of a synthetic CB₁/CB₂ receptor agonist significantly reduced aortic plaque area in the ApoE knockout mouse (845). Administration of the CB receptor agonist reduced macrophage infiltration into the atherosclerotic plaque, and reduced the expression of vascular cellular adhesion molecule-1 (VCAM-1), intercellular adhesion molecule-1 (ICAM-1), and P-selectin in the aorta, as well as reducing macrophage adhesion (845). Again, the observed beneficial effects appeared to be mediated by activation of the CB₂ receptor (845). A separate study confirmed the atheroprotective effects of selective CB₂ receptor activation by demonstrating increased vascular leukocyte infiltration in atherosclerotic plaques in mice lacking both the ApoE and CB₂ receptors compared to ApoE knockout mice, and decreased atherosclerotic plaque formation and reduced vascular superoxide release in ApoE knockout mice treated with a CB₂ receptor selective agonist (846). In contrast to these findings, a different study showed that activation or deletion of the CB₂ receptor did not modulate atherogenesis in the LDL receptor knockout mouse model of atherosclerosis (847). Another study suggested that the CB₂ receptor, while not affecting the size of atherosclerotic lesions in LDL receptor knockout mice, did increase lesional macrophage accumulation and smooth muscle cell infiltration, as well as reduce lesional apoptosis and alter the extra-cellular matrix of lesions (848). The findings from this study suggested that while the CB₂ receptor did not play a significant role in the initial formation of atherosclerotic lesions, it did play a role in modulating the progression of the disease (848). On the other hand, activation of the CB₁ receptor is associated with the release of reactive oxygen species and endothelial cell death (849), and CB₁ receptor blockade by rimonabant in ApoE knockout mice was associated with a significant reduction in the relative size of aortic atherosclerotic lesions (844). In conclusion, it appears that in the case of atherosclerosis, the CB₁ and CB₂ receptors play opposing roles—the CB₁ receptor appears to be atherogenic, whereas the CB₂ receptor appears to be anti-atherogenic (844,846,849,850,851) although some controversy still remains regarding the exact role played by the CB₂ receptor (852). Cannabidiol has also been shown to potently inhibit the activity of the enzyme 15-lipoxygenase, which has been implicated in the pathophysiology of atherogenesis (850,853). Further studies are needed in this area.

5.0 Precautions

The contraindications that apply to those considering using prescription cannabinoid-based therapies (such as nabilone (Cesamet®), nabiximols (Sativex®) or dronabinol (Marinol®)) also apply to those considering using cannabis. Currently, no clinical guidelines exist with respect to monitoring patients who are taking cannabis for therapeutic purposes.

The risk/benefit ratio of using cannabis should be carefully evaluated in patients with the following medical conditions because of individual variation in response and tolerance to its effects, as well as the difficulty in dosing noted in section 3.0:

- Cannabis should not be used in any person under the age of 18, or in any patient who has a history of hypersensitivity to any cannabinoid or to smoke. The adverse effects of cannabis use on mental health are greater during development, particularly during adolescence, than in adulthood (146,686,690) (see also section 7.7.3).
- Cannabis should not be used in patients with severe cardio-pulmonary disease because of occasional hypotension, possible hypertension, syncope, or tachycardia (117,233,234).
- Smoked cannabis is not recommended in patients with respiratory insufficiency such as asthma or chronic obstructive pulmonary disease (243).
- Cannabis should not be used in patients with severe liver or renal disease. Patients with ongoing chronic hepatitis C should be strongly advised to abstain from daily cannabis use, as this has been shown to be a predictor of steatosis severity in these individuals (32,854).
- Cannabis should not be used in patients with a personal history of psychiatric disorders (especially schizophrenia), or a familial history of schizophrenia.
- Cannabis should be used with caution in patients with a history of substance abuse, including alcohol abuse, because such individuals may be more prone to abuse cannabis, which itself, is a frequently abused substance (675,855,856).
- Patients with mania or depression and using cannabis or a cannabinoid should be under careful psychiatric monitoring (139,143,857).
- Cannabis should be used with caution in patients receiving concomitant therapy with sedative-hypnotics or other psychoactive drugs because of the potential for additive or synergistic CNS depressant or psychoactive effects (169,170,171) (also see section 7.7). Cannabis may also exacerbate the CNS depressant effects of alcohol and increase the incidence of adverse effects (see section 7.7). Patients should be advised of the negative effects of cannabis/cannabinoids on memory and to report any mental or behavioural changes that occur after using cannabis (178,181).
- Cannabis is not recommended in women of childbearing age not on a reliable contraceptive, as well as those planning pregnancy, those who are pregnant, or women who are breastfeeding (see sections 6.0 and 7.4).

6.0 Warnings

Cannabis is one of the most widely abused illicit drugs, and can produce physical and psychological dependence (122,156,210,858,859). The drug has complex effects in the CNS and can cause cognitive and memory impairment, changes in mood, altered perception, and decreased impulse control (152,180,860,861). Patients should be supervised when administration is initiated.

Dosing: In the case of smoked/vapourized cannabis, the dose required to achieve therapeutic effects and avoid adverse effects is difficult to estimate and is affected by the source of the plant material, its processing, and by different smoking techniques. These techniques include depth of inhalation, duration of breath-holding and the number and frequency of puffs, as well as how much of the cigarette is smoked or how much plant material is vapourized. Smoking or vapourization should proceed slowly and cautiously in a gradual fashion and should cease if the patient begins to experience the following effects: disorientation, dizziness, ataxia, agitation, anxiety, tachycardia and orthostatic hypotension, depression, hallucinations, or psychosis. There is also insufficient information regarding oral dosing, but the patient should be made aware that the effects following oral administration only begin to be felt 30 min to 1 h or more after ingestion, and that consumption of cannabis-based products (e.g. cookies, baked goods) should proceed slowly, and that edibles should be consumed only in small amounts at a time in order to gauge the effects and to prevent overdosing.

Psychosis: Anyone experiencing an acute psychotic reaction to cannabis or cannabinoids should promptly stop taking the drug and seek immediate medical attention. A psychotic reaction is defined as a loss of contact with reality characterized by one or more of the following: changes in thinking patterns (difficulty concentrating, memory loss, and/or disconnected thoughts), delusions (fixed false beliefs not anchored in reality), hallucinations (seeing, hearing, tasting, smelling or feeling something that does not exist in reality), changes in mood (intense bursts of emotion, absence of, or blunted emotions), very disorganized behaviour or speech, and thoughts of death and suicide (341).

Occupational hazards: Patients using cannabis should be warned not to drive or to perform hazardous tasks, such as operating heavy machinery, because impairment of mental alertness and physical coordination resulting from the use of cannabis or cannabinoids may decrease their ability to perform such tasks (182). Depending on the dose, impairment can last for over 24 h after last use because of the long half-life of Δ^9 -THC (62,131,290,862,863). Furthermore, impairment may be exacerbated with co-consumption of other CNS depressants (e.g. benzodiazepines, barbiturates, opioids, anti-histamines, muscle relaxants, or ethanol) (114,170,174,864,865,866).

Pregnancy: Pre-clinical studies suggest that endocannabinoid tone plays a critical role in fertilization, oviductal transport, implantation, and fetal/placental development (reviewed in (867)). One pilot clinical study suggested that high circulating levels of anandamide were associated with an increased incidence of miscarriage (868). Thus, there is a risk that maternal exposure to cannabis or cannabinoids could potentially adversely affect conception and/or maintenance of pregnancy. In addition, the use of cannabis during pregnancy should be avoided as there is some evidence of long-term developmental problems in children exposed to cannabis *in utero* (869,870). Men, especially those on the borderline of infertility and intending to start a family, are cautioned against using cannabis since exposure to cannabis or THC could potentially reduce the success rates of intended pregnancies (see section 7.4).

Lactation: Cannabinoids are excreted in human milk and may be absorbed by the nursing baby (871,872). Because of potential risks to the child, nursing mothers should not use cannabis.

6.1 Tolerance, dependence, and withdrawal symptoms

Tolerance, psychological, and physical dependence can occur with prolonged use of cannabis (118,210). Tolerance to cardiovascular effects occurs quickly, but dependence is slower to develop and appears more likely with higher, more frequent dosing (219,220). See section 2.4 for further information on tolerance, dependence, and withdrawal symptoms.

6.2 Drug interactions

The most clinically significant interactions may occur when cannabis is taken with other CNS depressant drugs such as sedative-hypnotics or alcohol (114,169,170,171,864,865,866,873,874). An overdose can occur if a patient is smoking/vapourizing cannabis and consuming orally administered cannabinoids, whether from prescription cannabinoid medications (e.g. dronabinol, nabilone), or from consumption of teas, baked goods or other products (174,290).

Xenobiotic-mediated inhibition or potentiation of cannabinoid metabolism

Δ^9 -THC is oxidized by the xenobiotic-metabolizing cytochrome P450 (CYP) mixed-function oxidases 2C9, 2C19, and 3A4 (62). Therefore substances that inhibit these CYP isoenzymes such as certain anti-depressants (e.g. fluoxetine, fluvoxamine, and nefazodone), proton pump inhibitors (e.g. cimetidine and omeprazole), macrolides (e.g. clarithromycin and erythromycin), anti-mycotics (e.g. itraconazole, fluconazole, ketoconazole, miconazole), calcium antagonists (e.g. diltiazem, verapamil), HIV protease inhibitors (e.g. ritonavir), amiodarone, and isoniazid can potentially increase the bioavailability of Δ^9 -THC as well as the chance of experiencing THC-related side effects (289,875,876). On the other hand, drugs that accelerate Δ^9 -THC metabolism via 2C9 and 3A4 isozymes such as rifampicin, carbamazepine, phenobarbital, phenytoin, primidone, rifabutin, troglitazone, and Saint John's Wort may conversely decrease the bioavailability of THC and hence its effectiveness if used in a therapeutic context (289,876).

Cannabinoid-mediated regulation of drug metabolism and drug transport

THC, CBD, and CBN are known to inhibit CYP isozymes such as CYP1A1, 1A2, and 1B1 (58). Cannabis may therefore increase the bioavailability of drugs metabolized by these enzymes. Such drugs include amitriptyline, phenacetin, theophylline, granisetron, dacarbazine, and flutamide (58). THC, carboxy- Δ^9 -THC, CBD, and CBN all stimulate, and in some cases even inhibit, the activity of the drug transporter P-glycoprotein *in vitro* (56). This suggests a potential additional role for these cannabinoids in affecting the therapeutic drug efficacy and toxicity of co-

administered drugs (56). Clinicians should therefore be aware other medications that the patient is taking and carefully monitor patients using other drugs along with cannabis or cannabinoids.

Cannabinoid-opioid interaction

Patients taking fentanyl (or related opioids) and anti-psychotic medications (clozapine or olanzapine) may also be at risk of experiencing adverse effects if co-consuming cannabis/cannabinoids (322,323,324,503,877). In one study, subjects reported an increase in the intensity and duration of the “high” when oxycodone was combined with inhalation of vapourized cannabis; this effect was not observed when morphine was combined with inhalation of vapourized cannabis (187). In that study, inhalation of vapourized cannabis was associated with a statistically significant decrease in the maximum concentration (C_{max}) of sustained-release morphine sulfate, and the time to C_{max} for morphine was also delayed, although the delay was not statistically significant (187). There were no changes in the AUC for morphine metabolites, or in the ratio of morphine metabolites to parent morphine (187). In contrast to the effects seen with morphine sulfate, inhalation of vapourized cannabis was not associated with any changes in oxycodone pharmacokinetics (187).

Evidence from pharmacogenetic studies

Pharmacogenetic studies have suggested that patients homozygous for the *CYP2C9**3 allele appear to have impaired THC metabolism and may show greater intoxication than **1*/**3* heterozygotes or **1*/**1* homozygotes (318).

Data from clinical studies

A significant proportion of published clinical studies of cannabis or prescription cannabinoid medications have used patient populations that were taking concomitant medications for a variety of disorders such as neuropathic pain of various etiologies (142,168,172,186,187,261,292,364,494,501,502,503), cancer-related pain (112,349,509), fibromyalgia (158,261,353,354), pain and spasticity associated with multiple sclerosis (188,262,291,361,428,504), and symptoms associated with Huntington’s or Parkinson’s disease (586,595). Examples of commonly-used medications seen in clinical trials of cannabis or prescription cannabinoid medications (e.g. dronabinol, nabilone and nabiximols) include non-steroidal anti-inflammatory drugs (e.g. acetaminophen, COX-2 inhibitors), metamizol, topical steroids, muscle relaxants, short- and long-acting opioids (e.g. codeine, morphine, hydromorphone, oxycodone, oxycontin, tramadol, fentanyl, methadone), ketamine, anti-convulsants (e.g. gabapentin, pregabalin), anti-depressants (e.g. tricyclics, selective-serotonin re-uptake inhibitors, serotonin-norepinephrine re-uptake inhibitors, serotonin-antagonist re-uptake inhibitors), and anxiolytics. According to the cited clinical studies, concomitant use of cannabis or prescription cannabinoid medications with other medications was reported to be well tolerated, and many of the observed adverse effects were those typically associated with the psychotropic effects of cannabis and cannabinoids (e.g. transient impairment of sensory and perceptual functions, abnormal thinking, disturbance in attention, dizziness, confusion, sedation, fatigue, euphoria, dysphoria, depression, paranoia, hallucinations, dry mouth, anxiety, hypotension, tachycardia, headache, throat irritation).

6.3 Drug screening tests

Because of the long half-life of elimination of cannabinoids and their metabolites, drug tests screening for cannabinoids can be positive for weeks after last cannabis/cannabinoid use (878,879) depending on the sensitivities of the tests used.

7.0 Adverse Effects

There is generally far more information available in the medical literature on the adverse effects associated with recreational cannabis use than there is with therapeutic cannabis use. Accordingly, much of the information presented below regarding the adverse effects of cannabis use comes from studies carried out among recreational users. Much less information on the adverse effects associated with the use of cannabis for therapeutic purposes comes from clinical studies, mainly because of the small number of such studies that have been carried out to date. Furthermore, while there is some information on the short-term adverse effects associated with the use of cannabis for therapeutic purposes, much less information exists on the long-term consequences of cannabis use for therapeutic purposes because all of the available clinical studies were short-term. A Canadian systematic review of the adverse effects of prescription cannabinoid medications concluded that the rate of non-serious adverse events was almost two-fold higher among those patients using prescription cannabinoid medications compared to controls (880). The most frequently cited adverse events associated with the use of prescription cannabinoid medications were nervous system disorders, psychiatric disorders, gastrointestinal disorders, and vascular and cardiac disorders (880). An additional consideration in the evaluation of adverse effects associated with cannabis use is the concomitant use of tobacco and alcohol as well as other drugs, whether they are non-prescription, prescription, or illicit drugs (122,881,882,883,884) (and also see section 6.2).

7.1 Carcinogenesis and mutagenesis

Qualitatively, cannabis smoke condensates have been shown to contain many of the same chemicals as tobacco smoke (70). Furthermore, a number of *in vitro* studies have provided strong evidence that smoke from burning cannabis is carcinogenic (reviewed in (118)). More recently, the cytotoxic and mutagenic potential of cannabis smoke condensates were compared to their tobacco counterparts (68). In contrast to tobacco smoke condensates, those derived from cannabis smoke appeared to be more cytotoxic and mutagenic, while the opposite was true with respect to cytogenetic damage (68). In addition, for either cannabis or tobacco smoke, the particulate phase was substantially more cytotoxic than the gas phase. Together, these studies suggest that cannabis smoke cannot be deemed "safer" than tobacco smoke.

Despite some persuasive *in vitro* data, the epidemiological evidence for a link between cannabis smoking and cancer remains inconclusive because of conflicting results from a limited number of studies. One epidemiological study in relatively young clients of a health maintenance organization (HMO) found an increased incidence of prostate cancer in those men who smoked cannabis and other non-tobacco materials (238). No other associations were found between cannabis use and other cancers; however, the study was limited by the demographics of the HMO clientele and the very low cannabis exposure threshold employed in the study to define "users". A case-control study suggested that cannabis smoking may increase the risk of head and neck cancer (Odds Ratio = 2.6; Confidence Interval = 1.1 - 6.6), with a strong dose-response pattern compared to non-smoking controls (239). However, the authors note a number of limitations with their study such as underreporting, inaccurate cannabis dose reporting, assay sensitivity, and low power. A large population-based case-control study, carried out in the year 2006, of 1 212 incident cancer cases and 1 040 cancer-free matched controls did not find a significant relationship between long-term cannabis smoking and cancers of the lung and upper aerodigestive tract (240). However, a smaller case-control study carried out in 2008 in young adults (≤ 55 years of age), examined 79 cases of lung cancer and 324 controls and reported that the risk of lung cancer increased by 8% (95% Confidence Interval = 2 - 15%) for each "joint-year" (defined as the smoking of one joint per day for one year) after adjusting for cigarette smoking (241). Despite the conflicting evidence surrounding the carcinogenic potential of cannabis smoke in humans, it is advisable to limit the degree to which cannabis is smoked. Further well-controlled epidemiological studies are required to better establish whether there is causality between cannabis smoking and carcinogenesis in human populations. Lastly, in the case of cancer patients, the potential risks of carcinogenesis and mutagenesis associated with smoking cannabis must be weighed against any potential therapeutic benefits for this patient population; routes of administration other than smoking (e.g. vapourization, oral administration) may warrant consideration. Because vapourization is a lower-temperature process compared with pyrolysis (i.e. smoking), vapourization appears to be associated with the formation of a smaller quantity of toxic by-products such as carbon monoxide, polycyclic aromatic hydrocarbons (PAHs), and tar, as well as a more efficient extraction of Δ^9 -THC from the cannabis material (273,281,282,283,284).

7.2 Respiratory tract

Differences in the smoking techniques used by cannabis vs. tobacco smokers are reported to result in three-fold higher levels of tar, and five-fold higher levels of carbon monoxide being retained in the lungs during cannabis smoking compared to tobacco smoking (885). A systematic comparison of the mainstream smoke composition from cannabis (Health Canada product) and tobacco cigarettes (prepared in the same way and consumed in an identical manner),

under two different sets of smoking conditions ("standard" and "extreme") has been reported (70). The "standard" condition reflects typical tobacco cigarette smoking conditions, whereas the "extreme" condition approaches that typically seen in cannabis smoking (70). Ammonia in mainstream cannabis smoke was 20-fold greater than that found in tobacco smoke, and oxides of nitrogen and hydrogen cyanide were three to five times higher in cannabis smoke vs. tobacco smoke. Carbon monoxide was significantly lower in mainstream cannabis smoke, under both smoking conditions. Tar was statistically significantly higher in mainstream cannabis smoke but only under the "extreme" smoking condition.

Mucosal biopsy specimens taken from chronic cannabis smokers, who reported smoking only cannabis, showed a number of histopathologic changes including basal cell hyperplasia, stratification, goblet cell hyperplasia, cell disorganization, inflammation, basement membrane thickening, and squamous cell metaplasia (242). However, the study employed a small number of subjects and relied on the accuracy and integrity of the subjects' recall to establish smoking status as well as frequency and duration of smoking. Epidemiological studies have found mild changes in pulmonary function in heavy cannabis smokers, including reduction of the forced expiratory volume in 1 second (FEV₁), an increase in airway resistance, and a decrease in airway conductance (244,245,246). Heavy chronic cannabis smokers presented with symptoms of bronchitis, including wheezing, production of phlegm and chronic cough, and long-term cannabis smoking may be a risk factor for chronic obstructive pulmonary disease in later life (122,886). All changes were most evident in heavy chronic users, defined as those who smoked more than three joints per day for 25 years (238,887), although evidence of measurable respiratory symptoms (e.g. decreased FEV₁/FVC ratio) was also observed in young, cannabis-dependent individuals whose smoking behaviour was comparable to tobacco smokers consuming 1 - 10 cigarettes/day (888). The potential risk of developing chronic obstructive respiratory disease, with long-term use and/or dependence, has been claimed to be potentially as great as among tobacco users (888). However, a recently published longitudinal study collecting repeated measurements of pulmonary function and smoking over a period of 20 years, in a cohort of 5 115 men and women in four U.S. cities (the CARDIA study), suggested a more complex picture. The study found a non-linear association between marijuana smoking and pulmonary function (247). By comparison, tobacco smoking (current and lifetime) was linearly associated with lower FEV₁ and FVC (247). Low levels of cumulative marijuana smoking were not associated with adverse effects on pulmonary function. Instead, at this level, marijuana smoking was associated with an increase in the FEV₁ and FVC values (247). At up to seven "joint-years" (a "joint-year" defined as smoking one joint/day, 365 days/year) of lifetime exposure there was no evidence of decreased pulmonary function. However, heavy chronic marijuana smoking (> ~30 joint-years or > ~25 smoking episodes per month) was associated with an accelerated decline in pulmonary function (FEV₁ but not FVC) (247).

Further research is needed to clarify the complex changes in lung function found in cannabis smokers, and to determine if there is a cause and effect relationship between cannabis smoking and the development of lung disease. Smoking cannabis may also increase the risk of developing respiratory infections in chronic users (889) through exposure to infectious organisms such as fungi and molds which can be found in the plant material (890), or alternatively by decreasing natural host defenses (891). However, further epidemiological research is also required to establish a causal relationship between cannabis smoking and respiratory infections. Vapourization of cannabis may be considered an alternative to smoking, although research is required to determine if there are any adverse effects of vapourization on lung health/function. For additional information on vapourization please consult sections 1.1.1, 1.1.2, 2.2.1.2, 3.4, 4.6.2.3, and Table 6.

7.3 Immune system

Pre-clinical studies

Evidence from *in vivo* and *in vitro* studies suggests complex and apparently dichotomous roles for the endocannabinoid system on immune system function (24). First, CB₁ and CB₂ receptors are known to be expressed in various immunocytes (B cells, monocytes, neutrophils, T lymphocytes, macrophages, mast cells), with CB₂ receptor expression generally being more abundant than CB₁ receptor expression; the ratio of CB₂ to CB₁ receptor expression ranges between 10 - 100 : 1 respectively, depending on the immune cell type in question (24,25). Second, immune cells also have the ability to synthesize, secrete, transport and catabolize endocannabinoids (24). Third, while stimulation of the CB₂ receptor appears to be generally associated with immunosuppressive effects, activation of the CB₁ receptor appears to be associated with an opposing immunostimulatory effect (24). Fourth, whereas certain cannabinoids have been shown to modulate the release of pro- or anti-inflammatory cytokines, pro-inflammatory cytokines (such as TNF- α) have, in turn, been reported to affect the functioning of the endocannabinoid system by upregulating the expression of both CB₁ and CB₂ receptor mRNA and protein levels (25). Thus, there appears to be some level of cross-talk between

the endocannabinoid and immune systems. Fifth, as is the case in other situations, Δ^9 -THC appears to have a biphasic effect on immune system function. Low doses of Δ^9 -THC seem to have stimulatory or pro-inflammatory effects, while higher doses appear to have inhibitory or immunosuppressive effects (266). Both Δ^9 -THC and CBD have been reported to modulate cell-mediated and humoral immunity, through CB receptor-dependent and CB receptor-independent mechanisms (266,892,893). Cannabinoids target various cellular signaling and transcriptional pathways resulting in the inhibition of pro-inflammatory cytokine release (e.g. IL-1 β , IL-6, IFN- β), and/or stimulation of anti-inflammatory cytokine release (e.g. IL-4, IL-5, IL-10, IL-13) (25,266). CBD also appears to induce a shift in Th1/Th2 immunobalance (892). While under certain circumstances, cannabinoids may appear to have broad anti-inflammatory and immunosuppressive functions which could be of benefit in pathological conditions having inflammatory characteristics, such beneficial functions may become problematic in the context of essential defensive responses to infections (24). For example, *in vitro* as well as *in vivo* experiments suggest cannabinoids have an impact on virus-host cell interactions (894): cannabinoid treatment was associated with increased viral replication of HSV-2, HIV-1, KSHV, influenza, and VSV viruses, or was associated with increases in surrogate measures of infection in these experimental models (895,896,897,898,899,900).

Taken together, the available information suggests that differences in the observed effects of cannabinoids on immune system function (i.e. immunosuppressive vs. immunostimulatory) may be explained by differences in the routes/methods of administration (smoked, oral, or other route), the length of exposure to the cannabinoid(s), the dose and type of cannabinoid used and which receptors are preferentially targeted, but also by differences between species, the experimental protocols and outcome measures that were used, and for clinical studies the health status/medical condition of the human subjects (266).

Clinical studies

The effects of cannabis smoking on the human immune system have been studied, but to a very limited degree. A major concern with HIV-positive cannabis smokers, or patients undergoing cancer chemotherapy, is that they might be more vulnerable than other cannabis smokers to the immunosuppressive effects of cannabis or that they risk exposure to infectious organisms associated with cannabis plant material (378). A group of studies has partially addressed the former concern. In one study, HIV-positive patients on stable anti-retroviral therapy were randomized to smoked cannabis or oral dronabinol and showed no changes in CD4+ and CD8+ T-cell, B cell, or NK cell counts and a number of other parameters compared with placebo, over a 21-day study period (901). A longitudinal study of 481 HIV-infected men who used cannabis and who were followed over an average five-year period found that while cannabis use was generally associated with a higher CD4+ cell count in infected men and controls, no clinically meaningful associations, adverse or otherwise, between cannabis use and T-cell counts and percentages could be established (902). Cannabis use was also not associated with an increased rate of progression to AIDS in HIV-infected individuals (903). In another study, smoking cannabis was associated with lower plasma concentrations of the protease inhibitors indinavir and nelfinavir; dronabinol or placebo had no effect (322). However, the decreased protease inhibitor levels were not associated with an elevated viral load, or changes in CD4+ or CD8+ cell counts (390).

In humans, smoking cannabis was also associated with poorer outcome in patients with chronic hepatitis C (882,904). Although pre-clinical studies strongly suggest that cannabinoids have broad immunomodulatory effects, and raise the possibility that cannabinoids may affect the ability of immunosuppressed patients to successfully resist or combat infections, it is unclear at this time if the immunomodulatory effects seen both pre-clinically and clinically translate into any clinically significant adverse outcomes.

Clear predictions concerning the effects of cannabinoids in those individuals who suffer from a dysregulated immune system are difficult to make because of the relative lack of available, comprehensive information on the subject. The clinician must therefore weigh the potential benefits of using cannabis and/or cannabinoids against the possible risks of using these substances on a case-by-case basis.

A recent cross-sectional study examined the association between cannabis use status and adherence to anti-retroviral therapy as well as the association between cannabis use status, HIV symptoms, and side effects associated with anti-retroviral therapy among a sample of HIV-positive individuals (905). The study reported that those subjects who had a cannabis use disorder (according to DSM-IV criteria and a Marijuana Smoking History Questionnaire score indicating daily cannabis or use more than once per day) had a significantly lower adherence to treatment than those who reported using cannabis once per week or more, but less than daily or not at all (905). Those who had a cannabis use disorder also had a higher viral load than those who used cannabis less than daily but at least once per week, as well as those who did not use at all; absolute CD4 count was not significantly different between groups (905). Furthermore, those

subjects with a cannabis use disorder reported significantly more frequent and severe HIV symptoms and/or medication side effects than those who used cannabis less than daily (but at least once per week), or those who reported not using cannabis at all (905). One limitation to this study was its cross-sectional nature, precluding the ability to establish a cause-and-effect relationship.

7.4 Reproductive and endocrine systems

Role of the endocannabinoid system in sexual physiology

The CB₁ receptor is widely expressed in various brain structures such as the striatum, hippocampus, and the cerebellum, as well as the amygdala, the midbrain, and the cerebral cortex—all structures that play various roles in regulating different aspects of sexual behaviour and function (269). For example, CB₁ receptors within the striatum and cerebellum may regulate motor activity and function; CB₁ receptors located within corticolimbic structures (e.g. prefrontal cortex, amygdala and hippocampus) may regulate stress responsivity and emotional behaviour; CB₁ receptors located within the dorsal raphe and ventral tegmental area may regulate genital reflexes, sexual motivation and inhibition; and lastly, CB₁ receptors expressed within the hypothalamus and the pituitary gland may modulate the functioning of the hypothalamic-pituitary-gonadal axis either directly through modulation of gonadotropin-releasing hormone or indirectly through other modulators (269,270).

CB₁ receptor-mediated modulation of the hypothalamic-pituitary axis results in the suppression of luteinizing hormone, thyroid stimulating hormone, growth hormone, and prolactin release from the pituitary gland, while the effects on follicle stimulating hormone are seemingly unclear but point to a probable suppression of release (268,906). In animals, these effects are accompanied by changes in reproductive function and behaviour including decreases in plasma testosterone levels, degenerative changes in spermatocytes and spermatids, anovulation, and potential reduction in copulatory behaviour (268,270). Aside from the roles of the cannabinoid receptors in the brain, the male and female reproductive systems also contain an endocannabinoid system, and increasing experimental evidence suggests important roles for the endocannabinoid system in regulating various reproductive functions such as folliculogenesis, spermatogenesis, ovulation, fertilization, oviductal transport, implantation, embryo development, pregnancy, and labour (reviewed in (37)).

Effects of cannabis on human sexual behaviour

There is a relative paucity of data with regards to the effects of cannabis or cannabinoids on human sexual behaviour. One review article has summarized the few available studies on the subject (269). It concluded that in general, the effects of cannabis on sexual functioning and behaviour appear to be dose-dependent. For women, the available information suggests beneficial effects on sexual behaviour and functioning (e.g. reported increases in sensitivity to touch and relaxation and a corresponding increase in sexual responsiveness) at low to moderate doses, and potentially opposite responses at higher doses (269). For men, the available information suggests that cannabis intake at low to moderate doses may facilitate sexual desire and activity, but that at higher doses or with more frequent or chronic use it may inhibit sexual motivation as well as erectile function (269). Results obtained from animal studies appear to mirror some of these findings, although exceptions have also been noted (269). Although the effects of cannabis on human sexual behaviour are still not well understood, some of its reported beneficial effects have been speculatively linked to its psychoactive properties (e.g. increase in tactile sensitivity/perception or slowing of temporal perception) or alternatively, to a loss of inhibitions and an increased state of relaxation (269).

Studies investigating the effects of cannabis consumption on testosterone levels in men have yielded conflicting results (269). While some investigators have found that acute or chronic cannabis consumption significantly lowered plasma testosterone levels in a dose-dependent manner, others have apparently failed to find similar effects (269). Differences in the reported effects of cannabis on testosterone levels among the various studies have been, in part, attributed to differences in the experimental protocols employed (269).

Effects on sperm and testicular health

The effects of cannabis and Δ^9 -THC on human sperm have been investigated both *in vivo* and *in vitro* (907,908,909). A significant decline in sperm count, concentration and motility, and an increase in abnormal sperm morphology were observed in men who smoked cannabis (8 - 20 cigarettes/day) for four weeks (907). In an *in vitro* study, sperm motility and acrosome reactions were decreased in both the 90% and 45% sperm fractions, the 90% fraction being the one with the best fertilizing potential and the 45% fraction being a poorer sub-population (909). Decreased sperm motility was observed in both fractions at Δ^9 -THC concentrations mimicking those attained recreationally (0.32 and 4.8 μ M), and in the 45% fraction at Δ^9 -THC concentrations typically seen therapeutically (0.032 μ M). Inhibition of the acrosome

reaction was only observed at the highest Δ^9 -THC concentration tested (4.8 μ M) in the 90% fraction, while the 45% fraction displayed decreased acrosome reactions at all three Δ^9 -THC concentrations tested. Such effects carry the possibility of impairing crucial sperm functions and male fertility, especially in those males already on the borderline of infertility (909).

A recently published, population-based, case-control study reported that compared with men who never used cannabis, those who had reported ever-using had a nearly two-fold increased risk of developing testicular germ-cell tumours of any histologic type (Odds Ratio = 1.94, 95% Confidence Interval: 1.02 - 3.68) and a greater than two-fold increased risk of non-seminoma or mixed germ-cell tumours (Odds Ratio = 2.42, 95% Confidence Interval: 1.08 - 5.42) (910). Men who reported using cannabis less than once per week appeared to have an elevated risk of developing testicular germ-cell tumours compared to those men who reported using cannabis more frequently. Men who reported using cannabis for a period under 10 years were also more than twice as likely to develop such tumours as those reporting \geq 10 years of use (910).

Effects on foetal development and child development

Results from human epidemiological studies examining short-term neonatal outcomes among women who smoked cannabis during pregnancy are equivocal; some report reduced neonatal birth weight and length (911,912,913,914) or a slightly increased risk of sudden infant death (915), while others report no effect (916,917,918). On the other hand, there appear to be some long-term effects on the development of children born to mothers who used cannabis during pregnancy. Two longitudinal investigations carried out over a time span of 20 years (reviewed in (869)) suggest that such *in utero* exposure impacts negatively on attentional behaviour and visual analysis and hypothesis testing, but not on standardized derived IQ scores. These findings were confirmed by a third study (870). These behavioural effects also appeared to have an adverse influence on aspects of executive function in later years.

Evidence suggests that cannabinoids accumulate in the breast milk of mothers who smoke cannabis and are transferred to newborns through breastfeeding (871,919). In a case-control study (920), exposure to cannabis from the mother's milk during the first month post-partum appeared to be associated with a decrease in infant motor development at one year of age.

7.5 Cardiovascular system

The most consistent acute physiological effect of smoking cannabis is dose-related tachycardia (121,226,232). While this is not usually considered dangerous for healthy young users, it may be dangerous to those already suffering from cardiac disorders or angina (118,921). Inhalation of cannabis smoke reduces the amount of exercise required to cause an angina attack by 50% (922), and has been associated with a five-fold increased risk of myocardial infarction in the first hour following smoking (232). This may be caused by a Δ^9 -THC-related increase in cardiac output, myocardial oxygen demand, catecholamine levels, and carboxyhemoglobin as well as postural hypotension (226,227,923). While tachycardia is observed in both occasional and chronic users, tolerance develops relatively quickly with the degree of tachycardia diminishing with use. After about 8 to 10 days of constant dosing with 10 mg of Δ^9 -THC per day (equivalent to 80 - 100 mg of cannabis containing 10% Δ^9 -THC), bradycardia (924) with a decrease in supine blood pressure was observed (925).

Cannabis is also known to cause peripheral vasodilatation, postural hypotension, and characteristic conjunctival reddening after smoking (926).

AIDS patients may be at an increased risk of experiencing adverse cardiovascular outcomes caused by interactions between cannabis and anti-retroviral drugs, such as ritonavir, which has been associated with adverse cardiovascular events (927).

There have been a number of case-reports of arteritis associated with long-standing, chronic, daily cannabis smoking (928,929,930,931). Case-reports have also suggested an association between chronic, daily cannabis smoking and multi-focal intracranial stenosis (932) and stroke (236,237).

7.6 Gastrointestinal system and Liver

7.6.1 Hyperemesis

There are an increasing number of case-reports being published regarding the “cannabis hyperemesis syndrome” (CHS). CHS is a condition observed in people chronically using cannabis on a daily basis, often for years, and is characterized by severe, intractable episodes of nausea and cyclic vomiting accompanied by abdominal pain (typically epigastric or periumbilical); these symptoms are relieved by compulsive hot water bathing or showering (194,195,196,197,198,199,200,201,202,203,204). The pathophysiology of CHS is not well understood (202). Treatment of patients presenting with this syndrome has been reported to include: recommending cessation of cannabis use, rehydration, and psychological counselling (200,202). The efficacy of anti-emetics such as metoclopramide, ondansetron, prochlorperazine, and promethazine in relieving the symptoms of nausea and vomiting in patients with CHS appears to be debatable (198,200,201,204). A recent case-report suggests that lorazepam (1 mg i.v., followed by 1 mg tablets b.i.d.) may provide some benefit in alleviating the symptoms of CHS, at least in the short-term (933).

7.6.2 Liver

A number of studies have strongly implicated the endocannabinoid system in chronic liver disease (934,935,936,937,938). Studies in patients with chronic hepatitis C have found a significant association between daily cannabis smoking and moderate to severe fibrosis (904), as well as cannabis smoking being a predictor of fibrosis progression (882). Another study showed that daily cannabis use was a predictor of steatosis severity in these individuals (854). Steatosis is an independent predictor of fibrosis progression and an established factor of poor response to anti-viral therapy (939). The authors recommend that patients with ongoing chronic hepatitis C be strongly advised to abstain from daily cannabis use.

In contrast, another study showed that modest cannabis use (defined as anything less than daily use in this study) was associated with an increase in the duration of time that patients remained on anti-retroviral treatment (252). This effect was postulated to contribute, at least in part, to an increase in the percentage of patients demonstrating a sustained virological response (i.e. the absence of detectable levels of hepatitis C virus RNA six months after completion of therapy) (252).

7.7 Central nervous system

The most frequently reported adverse events encountered with cannabinoids involve the central nervous system (CNS). Commonly reported CNS events in controlled clinical trials with dronabinol (Marinol®) and nabilimols (Sativex®) are intoxication-like reactions including drowsiness, dizziness, and transient impairment of sensory and perceptual functions (174,290). A “high” (easy laughing, elation, heightened awareness), which could be unwanted or unpleasant for patients, was reported in 24% of the patients receiving Marinol® as an anti-emetic, and in 8% of patients receiving it as an appetite stimulant (174). Other adverse events occurring at a rate of > 1% for Marinol® include anxiety/nervousness, confusion, and depersonalization (174). Dizziness, euphoria, paranoia, somnolence, abnormal thinking ranged from 3 - 10% (174). The rates of amnesia, ataxia, and hallucinations were > 10% when used as an anti-emetic at higher doses (174). Dizziness is the most common intoxication effect with Sativex®, reported initially in 35% of patients titrating their dose; the reported incidence of this effect in long-term use is approximately 25% (940). All other intoxication-like effects are reported by less than 5% of users (with the exception of somnolence, 7%) (940). Other events reported for Sativex® include disorientation and dissociation. Many, if not all, of the above-noted CNS effects also occur with cannabis.

7.7.1 Cognition

The acute effects of cannabis use on cognition have been reviewed by Lundqvist (235). Cannabis impairs cognition involving faculties such as short-term memory, attention, concentration, executive functioning and visuo-perception (180,941,942). The digit span task has been used to estimate the effects of cannabis on recent memory, but results have been inconsistent. Differences may be due to the dosage used, the smoking procedure, or whether the digit span task assesses forward or backward recall (943). Cannabis intoxication significantly impairs the ability to learn and recall word lists or short stories (944).

The long-term effects of cannabis on cognition remain controversial. Some studies report a positive association between cannabis consumption and cognitive deficits (945,946,947), or suggest that cognitive deficits persist after abstinence (180,941,948,949). Other studies did not find an association between cannabis use and long-term cognitive decline (948,949). Methodological limitations and the absence of powerful

effects have contributed to difficulties in assessing the effects of chronic use, and may help explain the discrepancies among studies (950,951). Nonetheless, studies generally suggest that chronic cannabis users suffer varying degrees of cognitive impairment that have the potential to be long-lasting (127). Prolonged use of ingested or inhaled cannabis in patients with multiple sclerosis was associated with poorer performance on various cognitive domains (e.g. information processing speed, working memory, executive function, and visuospatial perception), according to a cross-sectional study (178). A recently published, prospective, longitudinal study investigating the association between persistent cannabis use and neuropsychological functioning in a birth cohort of 1 037 individuals followed over a period of 20 years found that persistent cannabis use beginning in adolescence was associated with statistically significant global neuropsychological decline across a number of domains of functioning (952). Furthermore, cessation of cannabis use, for a period of one year or more, did not appear to fully restore neuropsychological functioning among adolescent-onset persistent cannabis users (952).

7.7.2 Psychomotor performance

Although no studies have been carried out to date examining the effects of cannabis or psychoactive cannabinoid exposure on psychomotor performance in individuals using these substances solely for medical purposes, it is well known that exposure to such substances impairs psychomotor performance (118) and patients must be warned not to drive or operate complex machinery after smoking or eating cannabis or consuming psychoactive cannabinoid medications (e.g. dronabinol, nabilone, nabiximols).

A double-blind, placebo-controlled, crossover study comparing the effects of a medium dose of dronabinol (20 mg) and of two hemp milk decoctions, containing medium (16.5 mg) or high doses (45.7 mg) of THC, reported severe impairment on several performance skills required for safe driving (953). A "moderate" dose (21 mg of THC) was associated with impairments in motor and perceptual skills necessary for safe driving (954). In one study, performance impairment appeared to be less significant among heavy cannabis users compared to occasional users, potentially because of the development of tolerance or compensatory behaviour (169). It has been suggested that, unlike alcohol, cannabis users are aware of their level of intoxication and compensate by becoming hyper-cautious; in tasks such as driving, this kind of behaviour results in decreased speed, decreased frequency of overtaking, and an increase in following distance (955,956). Others disagree with this assertion ((957) and also see (176)).

A recent double-blind, placebo-controlled, randomized, three-way, crossover design study suggested that administration of dronabinol dose-dependently impaired driving performance in both occasional (defined as using a cannabinoid between 5 and 36 times per year) and heavy cannabis users (defined as using 1 - 3 joints per day, > 160 times per year) (958). However, the magnitude of the impairment appeared to be less in heavy users, possibly due to tolerance (958). The authors indicate that driving impairments after dronabinol were of clinical relevance and comparable to drivers operating their vehicles at a blood-alcohol concentration of greater than 0.8 mg/mL (0.08 g%) (958). Approximately 25% of the "heavy users" demonstrated impairment equivalent to, or worse than, that reported for drivers with a blood-alcohol concentration of 0.5 mg/mL (0.05 g%). Driving impairments after dronabinol use were evident even though THC plasma concentrations were relatively low (varying between 2 and 10 ng/mL) (175,958).

A recent case-control study estimating accident risk for a variety of substances including alcohol, medicines, and illegal drugs found that the odds ratio for accident risk for all the THC concentrations measured (1 to > 5 ng/mL) was statistically significant (959). At whole-blood concentrations of ≥ 2 ng/mL THC, the risk of having an accident was significantly increased (959). One study found that the risk of responsibility for fatal traffic crashes, while driving under the influence of cannabis, increased with increasing blood concentrations of THC such that there was a significant dose-effect relationship between risk of responsibility for fatal traffic crashes and blood concentrations of THC. The study showed that the odds ratio of having a fatal crash increased from 2.18 if blood concentrations ranged between 0 and 1 ng/mL of THC, to 4.72 if blood THC concentrations were ≥ 5 ng/mL (960). The findings from this study further support the notion of a causal relationship between cannabis use and crashes (960). Another study suggested that drivers who were judged (by a police physician) as being impaired had higher blood THC concentrations than drivers judged not to be impaired (median: 2.5 ng/mL vs. 1.9 ng/mL) (961). Using a binary logistic regression model, the odds ratio for being judged impaired appeared to increase with increasing drug concentrations from 2.9 ng/mL onwards (961). Serum THC concentrations between 2 and 5 ng/mL have been identified as a threshold above which THC-induced impairment of skills related to driving become apparent (133,959). Performance impairment

after cannabis intake was reported to be highest during the first hour after smoking, and between 1 - 2 h after oral intake, and declining after 3 - 4 h (or longer in the case of oral ingestion) (862,961).

A recent meta-analysis of observational studies examining acute cannabis consumption and motor vehicle collision risk reported that driving under the influence of cannabis was associated with a significantly increased risk of motor vehicle collisions compared with unimpaired driving, with an odds ratio of 1.92 (95% Confidence Interval = 1.35 - 2.73; $p = 0.0003$) (175). Collision risk estimates were higher in case-control studies and studies of fatal collisions, than in culpability studies and studies of non-fatal collisions (175). It has been reported that individuals who drive within 1 h of using cannabis are nearly twice as likely to be involved in motor vehicle accidents as those who do not consume cannabis (954). For this meta-analysis, only observational studies with a control or comparison group, including cohort (historical prospective), case-control, and culpability designs were included, and experimental laboratory or simulator studies were excluded (175). Furthermore, only studies that assessed acute or recent cannabis use were examined. This meta-analysis supports the findings of other studies which suggest that cannabis use impairs the performance of the cognitive and motor tasks that are required for safe driving, thereby increasing the risk of collision (175). Although driving simulator studies have reported a dose-response effect, in which elevated concentrations of THC were associated with increased crash risk, dose-response effects could not be established in this study (175).

A double-blind, counter-balanced, placebo-controlled driving simulator study reported that driving performance was more impaired in subjects who co-consumed alcohol and low or high doses of THC by smoking cannabis cigarettes (176). The level of THC detected in the blood was higher when cannabis was consumed along with alcohol than when consumed alone (176). It also appeared that regular cannabis users displayed more driving errors than non-regular cannabis users (176).

A recent systematic review and meta-analysis concluded that, after adjusting for study quality, cannabis use was associated with a seven-fold estimated risk of being involved in a fatal accident, benzodiazepine use was associated with a two-fold estimated risk of a fatal accident, and opiate use with a three-fold estimated risk of a fatal accident (177). In contrast, cannabis use was associated with a 1.5-fold estimated risk of having an accident that only caused injury, benzodiazepine use was associated with a 0.71-fold estimated risk, whereas opiates were associated with a 21-fold estimated risk of having an accident that only caused injury (177).

7.7.3 Psychiatric effects

7.7.3.1 Acute psychotic reactions

Cannabis and cannabinoid use has been linked to episodes of acute psychosis in both regular and drug-naïve users (122,145,962). In one report, two healthy patients who had participated in a randomized controlled trial (RCT) measuring the effects of orally administered cannabinoids (including dronabinol or cannabis decoctions) on psychomotor performance displayed acute psychotic reactions following exposure to cannabis (145). The subjects had no psychiatric history or concomitant drug use, but were "occasional" regular cannabis users. In another RCT, 22 healthy subjects, also with a history of occasional cannabis use, no concomitant drug use, and with no psychiatric disorders received intravenous doses of Δ^9 -THC paralleling peak plasma THC levels achieved by smoking cannabis cigarettes containing 1 - 3.5% Δ^9 -THC (140). Drug administration was associated with a range of acute, transient, behavioural, and cognitive effects including suspiciousness, paranoid and grandiose delusions, conceptual disorganization, and illusions. Depersonalization, derealization, distorted sensory perceptions, altered bodily perceptions, feelings of unreality, and extreme slowing of time were also reported. Furthermore, blunted affect, reduced rapport, lack of spontaneity, psychomotor retardation, and emotional withdrawal were observed. Another study reported similar results (963).

7.7.3.2 Anxiety, Depression and Bipolar Disorder

Anxiety and depression

Cannabis is known to cause an acute and short-lasting episode of anxiety, often resembling a panic attack; this is more often encountered in naïve cannabis users and those who consume higher doses of cannabis or THC (> 5 mg oral Δ^9 -THC), and also when cannabis is consumed in novel or stressful environments (147,155). While clinical trials of cannabis, or oral Δ^9 -THC, to treat anxiety or depression show either a lack of improvement or worsening of these conditions (964,965,966,967) there is some evidence that cannabis or cannabinoids may be useful in treating anxiety or depression secondary to other disorders (e.g. chronic pain, post-traumatic stress disorder). For more information on potential therapeutic uses of cannabis or cannabinoids to treat anxiety and depression, please consult section 4.8.5.1.

Research on the topic of cannabis and depression is relatively scarce and conflicting. A 2003 review reported that the co-morbidity level between heavy or problematic cannabis use and depression, in surveys of the general population, exceeds what would be expected by chance (968). The authors also identify a modest association between early-onset regular or problematic use and later depression. However, limitations in the available research on cannabis and depression, including limitations in study design, as well as limitations in the ability to measure cannabis use, and limitations in the ability to measure depression were also highlighted. A U.S. study of adults using longitudinal national survey data (n = 8 759) found that the odds of developing depression in past-year cannabis users was 1.4 times higher than the odds of non-users developing depression (969). However, after adjusting for group differences, the association was no longer significant. In a 2008 study, the same group looked at the relationship between cannabis use and depression among youth using a longitudinal cohort of 1 494 adolescents. Similar to the adult study, the results did not support the causal relationship between adolescent-onset cannabis use problems and early adult depression (970). In contrast, another U.S. study based on the results of the National Epidemiological Survey on Alcohol and Related Conditions (n = 43 093) found that major depression was significantly associated with lifetime cannabis disorders and dependence (971). A 2007 study using data from the Netherlands Mental Health Survey and Incidence Study found a modest increased risk of a first depressive episode (Odds Ratio = 1.62; 1.06 - 2.48), after controlling for strong confounding factors (972). Of greater significance in this study was the strong increased risk of bipolar disorder (Odds Ratio = 4.98; 1.80 - 13.81) with cannabis use (see below for further information on cannabis and bipolar disorder). There was a dose-response relationship associated with the risk of 'any mood disorder' for almost daily and weekly users, but not for less frequent users. A survey of 248 French high school students found that cannabis users had significantly higher rates of suicidal behaviours and depressive and anxious symptoms compared to non-users (973). Another study suggested a putative positive association between exposure to cannabis and protracted suicidal thoughts or attempts in young people, although the study suffered from a number of limitations (974).

Bipolar disorder

Cannabis is one of the most frequently abused drugs in people diagnosed with bipolar disorder (148,975,976,977,978). A number of studies have examined the relationship between cannabis use and bipolar disorder, its effect on disease course, and its effect on treatment compliance.

One three-year, prospective study involving 4 815 subjects attempted to determine if baseline cannabis use increased the risk for development of manic symptoms, if the association between cannabis use and mania was independent of the emergence of psychotic symptoms, and if baseline mania predicted cannabis use at follow-up (975). The authors found that cannabis use at baseline was associated with follow-up mania (Odds Ratio = 5.32, 95% Confidence Interval: 3.59, 7.89). After adjusting for confounding factors, the association persisted although it was reduced (Odds Ratio = 2.70, 95% Confidence Interval: 1.54, 4.75). The risk of developing manic symptoms appeared to increase with increased baseline frequency of cannabis use (975). The effect size was largest for those who used cannabis 3 - 4 days/week, followed by those who used daily and 1 - 2 days/week, and lastly for those who used 1 - 3 days/month (975). The authors reported that manic

symptoms at baseline did not predict cannabis use during follow-up. The results suggested that use of cannabis increased the risk of developing subsequent manic symptoms and that this effect was dose-dependent (975).

Another group of investigators conducted a five-year, prospective, cohort study examining three groups of patients: one where a cannabis use disorder preceded the onset of bipolar disorder, another where bipolar disorder preceded a cannabis use disorder, and one group with bipolar disorder only (976). The authors found that cannabis use was associated with more time in affective (manic or mixed) episodes and with rapid cycling, but a causal relationship between cannabis use and bipolar disorder could not be established (976).

A separate prospective study which followed a group of type I bipolar patients over a 10-year period, beginning from the onset of illness, concluded that there was a strong association between cannabis use and manic/hypomanic episodes or symptoms, and that substance abuse preceded or coincided with, but did not follow, exacerbations of affective illness (979).

A two-year, prospective, observational study on the outcome of pharmacological treatment of mania (the European Mania in Bipolar Longitudinal Evaluation of Medication (EMBLEM) study) followed 3 459 eligible in- and out-patients who were being treated for acute mania in bipolar disorder, assessing patients' current cannabis use as well as the influence of cannabis exposure on clinical and social treatment outcome measures (148). The study concluded that during a one-year treatment period, cannabis users exhibited less treatment compliance and higher levels of overall illness severity, mania, and psychosis compared to non-users (148). Cannabis users also reported experiencing less satisfaction with life (148).

A preliminary study found that patients diagnosed with bipolar disorder with psychotic features were significantly more likely to carry a functional polymorphism in the promoter region of the 5-HT transporter gene and also have a diagnosis of cannabis abuse/dependence, compared to bipolar patients who did not exhibit psychotic symptoms (978). Genetic studies have also raised the possibility of a link between allelic variants of the cannabinoid receptor gene (*CNR1*) and susceptibility to mood disorders (980,981).

The influence of cannabis use on age at onset in both schizophrenia and bipolar disorder (with psychotic symptoms) has been studied using regression analysis (150). The authors of this study found that although cannabis and other substance use was more frequent in patients with schizophrenia than those diagnosed with bipolar disorder, cannabis use was nonetheless associated with a decrease in age at onset in both disorders (150). Cannabis use also preceded first hospitalization in the vast majority of cases (95.4%). Furthermore, the period of most intensive use ("several times per day") preceded first admission in 87.1% of the cases (150). In bipolar patients, cannabis use reduced age at onset by an average of nine years (150). In contrast, in schizophrenic patients, cannabis use reduced age at onset by an average of 1.5 years (150). No significant difference was noted in age at onset between male and female patients in either of the diagnostic groups (150).

Another study investigated which factors were associated with age at onset in bipolar disorder, and also examined the sequence of the onsets of excessive substance use and bipolar disorder (982). A total of 151 patients with bipolar disorder (type I and II) receiving psychiatric treatment participated in the study. The authors found that when compared with alcohol use, excessive cannabis use (defined as either meeting DSM-IV criteria for substance use disorder, or weekly use of cannabis over a period of at least four years) was associated with an earlier age at onset in both primary and secondary bipolar disorder, even after adjusting for possible confounders (982). In addition, the mean age at onset of excessive cannabis use preceded the age at onset of bipolar disease; this was reversed in the alcohol group (982).

One study reported that when compared with controls, patients with bipolar disorder were almost seven times (95% Confidence Interval: 5.41 - 8.52) more likely to report a lifetime history of cannabis use (977). Furthermore, this association appeared to be gender-independent. Those patients who used cannabis after, or in tandem with, their onset of bipolar symptoms had a lower

age at onset of the disorder (17.5 vs. 21.5 yrs) (977). Furthermore, those who used cannabis prior to the onset of a bipolar disease episode were 1.75 times (95% Confidence Interval: 1.05 - 2.91) more likely to report disability attributable to bipolar disorder (977).

Lastly, a retrospective analysis of a large cohort of bipolar I subjects, with or without a history of a cannabis use disorder, reported that bipolar patients with a cannabis use disorder had similar age at onset as patients without such a substance use disorder (983). However, patients with a cannabis use disorder were more likely to have experienced psychosis at some time during the course of their illness compared to patients who never met the criteria for the disorder (983).

7.7.3.3 Schizophrenia and psychosis

The endocannabinoid system has been implicated in the pathogenesis of schizophrenia and psychosis (please see section 4.8.5.5 for more information). Individuals with schizophrenia, or with a family history of this disorder, are likely to be at greater risk of suffering adverse psychiatric effects as a result of using cannabis or psychoactive cannabinoids such as Δ^9 -THC (152). Heavy cannabis use can aggravate psychotic symptoms and cause more relapses, and those individuals who use cannabis are at an increased risk of a poor prognosis (118,138,984,985). Self-reported use of cannabis in adolescence has been associated with an increased risk of developing schizophrenia, and this risk was related to frequency of cannabis exposure (986). A cohort study of over 1 000 children followed from birth to age 26 reported a three-fold increased risk of psychotic disorders in those who used cannabis, and suggested that cannabis exposure among psychologically vulnerable adolescents should be strongly discouraged (987). The relationship between cannabis use and psychotic symptoms was also studied in a cohort of 2 437 young people (ages 14 - 24 yrs) who had greater than average pre-disposition for psychosis, and who had first used cannabis during adolescence (146). The authors found a dose-response relationship between frequency of cannabis use and the risk of psychosis. The effect of cannabis use was also much stronger in those individuals with a pre-disposition for psychosis. A systematic review of evidence pertaining to cannabis use and the occurrence of psychotic or affective mental health outcomes reported an increased risk of any psychotic outcome in individuals who had ever used cannabis compared with non-users (Odds Ratio = 1.41) (141). Furthermore, the findings appeared to show a dose-related effect, with greater risk to individuals who used cannabis most frequently (Odds Ratio = 2.09) (149,150).

In one study, the relationship between age at onset of psychosis and other clinical characteristics in a sample of well-characterized patients diagnosed with bipolar disorder with psychosis, schizoaffective disorder, or schizophrenia, has been investigated (149). The study concluded that lifetime cannabis abuse/dependence was associated with a significantly earlier age at onset of psychosis (3.1 years, 95% Confidence Interval: 1.4 - 4.8) (149). Furthermore, among those patients with lifetime cannabis abuse/dependence, the age at onset of cannabis abuse/dependence preceded the onset of psychotic illness by almost another three years (149). However, patients who had a lifetime cannabis abuse/dependence diagnosis and a lifetime alcohol abuse/dependence diagnosis had a significantly later age at onset of psychosis (149).

Another study looked at the influence of cannabis use on age at onset in both schizophrenia and bipolar disorder (with psychotic symptoms) using regression analysis (150). The authors of this study found that although cannabis and other substance use was more frequent in patients with schizophrenia than those diagnosed with bipolar disorder, cannabis use was nonetheless associated with a decrease in age at onset in both disorders (150). Cannabis use also preceded first hospitalization in the vast majority of cases (95.4%) and furthermore, the period of most intensive use ("several times per day") preceded first admission in 87.1% of the cases (150). In bipolar patients, cannabis use reduced age at onset by an average of nine years (150). In contrast, in schizophrenic patients, cannabis use reduced age at onset by an average of 1.5 years (150). No significant difference was noted in age at onset between male and female patients in either of the diagnostic groups (150).

Although cannabis use increases the risk of psychosis, it is only one factor in a larger constellation of contributing factors (988).

Genetic factors

A number of studies have investigated the influence of potential genetic factors in the development of psychosis and schizophrenia, and more specifically as a function of interaction with cannabis use. Some studies have focused on the role of genetic polymorphisms at the catechol-O-methyltransferase gene (*COMT*) (686,687,688,689,690), while others have focused on polymorphisms at the *AKT1* gene (691,692,693), or the brain-derived neurotrophic factor (*BDNF*) gene (989).

Schizophrenia and the Catechol-O-Methyltransferase gene

Catechol-O-methyltransferase (*COMT*) regulates the breakdown of catecholamines, including neurotransmitters such as dopamine, epinephrine, and norepinephrine (690). A missense mutation at codon 158 in the *COMT* gene, causing a substitution to the methionine (Met) at the positional valine (Val) (Val158Met), results in an enzyme with decreased activity and correspondingly slower dopamine catabolism (990,991). Changes in dopaminergic tone and signaling are known to affect neurophysiological function, and these changes have been implicated in the pathophysiology of schizophrenia (992). Although a large-scale association study and meta-analysis has failed to find a strong association between the Val158Met *COMT* polymorphism and vulnerability to schizophrenia (993), evidence gathered from convergent functional genomic data nevertheless implicates the *COMT* gene (as well as the *CNR1* and 2 genes) in the pathophysiology of schizophrenia (994). Caspi et al. (686) followed an epidemiological birth cohort of 1 037 children longitudinally across the first three decades of life. They concluded that the *COMT* Val/Val homozygous genotype interacted with adolescent-onset cannabis use, but not adult-onset use, to predict the emergence of adult psychosis (686). Subsequent studies confirmed and extended these findings (687,688,689,690,693). Carriers of the Val allele were most sensitive to Δ^9 -THC-induced psychotic experiences (especially if they scored highly on a psychosis liability assessment), and were also more sensitive to the Δ^9 -THC-induced memory and attention impairments compared to carriers of the Met allele (687). Homozygous carriers of the Val allele, but not subjects with the homozygous Met genotype, showed an increase in the incidence of hallucinations after cannabis exposure, but this was conditional on prior psychometric evidence of psychosis liability (688). Those patients who were Val/Met heterozygous also appeared to be more sensitive to the effects of cannabis than Met homozygotes, but less sensitive than Val homozygotes (688). Another study suggested that cannabis use could reduce the (protective) delay effect of the *COMT* Met allele in influencing the age of onset of psychosis (689). These findings were supported, and extended, by a subsequent study which showed that those who started using cannabis earlier had an earlier age at onset of psychiatric disorders, and that carriers of the Val homozygous genotype had an earlier age of onset of psychosis compared to Met carriers (690). The authors of this study concluded that gene-environment interaction (i.e. the combination of the *COMT* Val to Met polymorphism and cannabis use) may modulate the emergence of psychosis in adolescents (690). Taken together, these studies also suggest the presence of a gene-dosage effect, with increasing disease risk among Val/Val homozygotes, moderate risk in Val/Met heterozygotes, and less risk among Met/Met homozygotes.

Schizophrenia and the AKT1 gene

Other studies have focused on the role of *AKT1*, a gene that encodes a protein kinase involved in the dopamine and cannabinoid receptor signaling cascades, and which is involved in regulating cellular metabolism, cell stress, cell-cycle regulation, and apoptosis as well as regulating neuronal cell size and survival (691). In one study, the authors found evidence of a gene-environment interaction between a single nucleotide polymorphism in the *AKT1* gene (rs2494732, C/C homozygous polymorphism) and cannabis use (692). Individuals with the C/C homozygous polymorphism had an approximately two-fold increased risk of being diagnosed with a psychotic disorder after having used cannabis either daily or weekly (692). In contrast, C/T heterozygous individuals had only a slightly increased risk of developing cannabis-related psychosis compared to T/T homozygotes, which served as the controls (692). In another study by the same group, individuals with the rs2494732 C/C homozygous polymorphism exhibited a deficit in sustained attention, but not in verbal memory, even in the absence of current cannabis use (691).

Schizophrenia and the Brain-Derived Neurotrophic Factor gene

One study found that cannabis use, before diagnosis of schizophrenia, was associated with a decrease in the age at onset of a psychotic disorder, decreasing the age at first admission by almost three years (989). Furthermore, a dose-dependent association between cannabis use and age at onset of psychotic symptoms was found, with an earlier onset of psychotic disorder in heavier users (989). A significant association between a younger age of first cannabis use and an earlier onset of psychotic disorder was also found, even after controlling for possible confounders (989). In that study, cannabis use independently predicted age at onset of a psychotic disorder in male patients, whereas in female patients cannabis use was only associated with age at onset of psychotic disorder in those who carried a Met allele mutation in the gene for brain-derived neurotrophic factor (*BDNF*). Female carriers of the mutant Met allele presented with psychotic symptoms seven years earlier than female patients who did not use cannabis and who had a *BDNF* Val/Val genotype (989).

In conclusion, given the evidence suggesting a strong genetic component in the modulation of psychosis, and especially psychosis or schizophrenia precipitated by cannabis use, the taking of a thorough patient medical history, especially one which includes a psychiatric history/evaluation, would be very valuable in determining whether cannabis/cannabinoids represent a sensible and viable therapeutic option.

7.7.3.4 Amotivational syndrome

The term "amotivational syndrome" is generally used to qualify people who exhibit apathy, lack of motivation, social withdrawal, narrowing of interests, lethargy, impaired memory, impaired concentration, disturbed judgement, and impaired occupational achievement (995).

Some investigators suggest that heavy, chronic use of cannabis is linked to the development of such a syndrome (995); de-intoxication results in resolution of symptoms (152,996). Other investigators have not found such a causal relationship (995,997).

8.0 Overdose/Toxicity

LD₅₀ values for rats administered single oral doses of THC, or crude cannabis extract, are approximately 1000 mg/kg (998). Dogs and monkeys are able to tolerate significantly higher oral doses of THC, or cannabis extract, of 3000 mg/kg (or greater in certain cases) (998). The estimated human lethal dose of intravenous THC is 30 mg/kg (2100 mg/70 kg) (174), although there has been no documented evidence of death exclusively attributable to cannabis overdose to date. Significant CNS symptoms are observed with oral doses of 0.4 mg/kg dronabinol (Marinol[®]) (174). Cannabis and THC often produce unwanted physical effects, typically dizziness, sedation, intoxication, transient impairment of sensory and perceptual functions, clumsiness, dry mouth, lowered blood pressure, or increased heart rate (174,999). These adverse effects are generally tolerable and not unlike those seen with other medications (118). The rare acute complications (e.g. panic attacks, psychosis, convulsions, etc.) that present to hospital Emergency Departments can be managed with conservative measures, such as reassurance in a quiet environment, and administration of benzodiazepines, if required (1000). As is stated in the case of overdose with Marinol[®] (174), the signs and symptoms observed with smoked or ingested cannabis are an extension of the psychotomimetic and physiologic effects of THC. Individuals experiencing psychotic reactions should stop using cannabis or cannabinoids immediately and seek prompt medical/psychiatric attention.

Reference List

1. Rodriguez de Fonseca, F., del Arco, I, Bermudez-Silva, F. J., Bilbao, A. and others. (2005). The endocannabinoid system: physiology and pharmacology. *Alcohol Alcohol.* 40: 2-14.
2. Serrano, A. and Parsons, L. H. (2011). Endocannabinoid influence in drug reinforcement, dependence and addiction-related behaviors. *Pharmacol.Ther.* 132: 215-241.
3. Maccarrone, M., Gasperi, V., Catani, M. V., Diep, T. A. and others. (2010). The endocannabinoid system and its relevance for nutrition. *Annu.Rev.Nutr.* 30: 423-440.
4. Aggarwal, S. K. (2012). Cannabinergic Pain Medicine: A Concise Clinical Primer and Survey of Randomized-controlled Trial Results. *Clin.J.Pain.* 29: 162-171.
5. Bradshaw, H. B. and Walker, J. M. (2005). The expanding field of cannabimimetic and related lipid mediators. *Br.J.Pharmacol.* 144: 459-465.
6. De Petrocellis, L. and Di Marzo, V. (2009). An introduction to the endocannabinoid system: from the early to the latest concepts. *Best.Pract.Res.Clin.Endocrinol.Metab.* 23: 1-15.
7. De Petrocellis, L. and Di Marzo, V. (2010). Non-CB1, non-CB2 receptors for endocannabinoids, plant cannabinoids, and synthetic cannabimimetics: focus on G-protein-coupled receptors and transient receptor potential channels. *J.Neuroimmune.Pharmacol.* 5: 103-121.
8. O'Sullivan, S. E. and Kendall, D. A. (2010). Cannabinoid activation of peroxisome proliferator-activated receptors: potential for modulation of inflammatory disease. *Immunobiology.* 215: 611-616.
9. Hansen, H. S. (2010). Palmitoylethanolamide and other anandamide congeners. Proposed role in the diseased brain. *Exp.Neurol.* 224: 48-55.
10. Ben-Shabat, S., Frider, E., Sheskin, T., Tamiri, T. and others. (1998). An entourage effect: inactive endogenous fatty acid glycerol esters enhance 2-arachidonoyl-glycerol cannabinoid activity. *Eur.J.Pharmacol.* 353: 23-31.
11. Quarta, C., Mazza, R., Obici, S., Pasquali, R. and others. (2011). Energy balance regulation by endocannabinoids at central and peripheral levels. *Trends Mol.Med.* 17: 518-526.
12. Battista, N., Di Tommaso M., Bari, M., and Maccarrone, M. (2012). The endocannabinoid system: an overview. *Front.Behav.Neurosci.* 6: 9-
13. Horvath, B., Mukhopadhyay, P., Hasko, G., and Pacher, P. (2012). The endocannabinoid system and plant-derived cannabinoids in diabetes and diabetic complications. *Am.J.Pathol.* 180: 432-442.
14. Hermanson, D. J. and Marnett, L. J. (2011). Cannabinoids, endocannabinoids, and cancer. *Cancer Metastasis Rev.* 30: 599-612.
15. Rouzer, C. A. and Marnett, L. J. (2011). Endocannabinoid oxygenation by cyclooxygenases, lipoxygenases, and cytochromes P450: cross-talk between the eicosanoid and endocannabinoid signaling pathways. *Chem.Rev.* 111: 5899-5921.
16. Pertwee, R. G. (2008). The diverse CB1 and CB2 receptor pharmacology of three plant cannabinoids: delta9-tetrahydrocannabinol, cannabidiol and delta9-tetrahydrocannabivarin. *Br.J.Pharmacol.* 153: 199-215.
17. Di Marzo, V., Piscitelli, F., and Mechoulam, R. (2011). Cannabinoids and endocannabinoids in metabolic disorders with focus on diabetes. *Handb.Exp.Pharmacol.* 75-104.

18. Bab, I. and Zimmer, A. (2008). Cannabinoid receptors and the regulation of bone mass. *Br.J.Pharmacol.* 153: 182-188.
19. Howlett, A. C., Barth, F., Bonner, T. I., Cabral, G. and others. (2002). International Union of Pharmacology. XXVII. Classification of cannabinoid receptors. *Pharmacol.Rev.* 54: 161-202.
20. Pertwee, R. G., Howlett, A. C., Abood, M. E., Alexander, S. P. and others. (2010). International Union of Basic and Clinical Pharmacology. LXXIX. Cannabinoid receptors and their ligands: beyond CB and CB₂. *Pharmacol.Rev.* 62: 588-631.
21. Kraft, B. (2012). Is there any clinically relevant cannabinoid-induced analgesia? *Pharmacology.* 89: 237-246.
22. Guindon, J. and Hohmann, A. G. (2009). The endocannabinoid system and pain. *CNS.Neurol.Disord Drug Targets.* 8: 403-421.
23. Teixeira, D., Pestana, D., Faria, A., Calhau, C. and others. (2010). Modulation of adipocyte biology by delta(9)-tetrahydrocannabinol. *Obesity.(Silver.Spring).* 18: 2077-2085.
24. Greineisen, W. E. and Turner, H. (2010). Immunoactive effects of cannabinoids: considerations for the therapeutic use of cannabinoid receptor agonists and antagonists. *Int.Immunopharmacol.* 10: 547-555.
25. Jean-Gilles, L., Gran, B., and Constantinescu, C. S. (2010). Interaction between cytokines, cannabinoids and the nervous system. *Immunobiology.* 215: 606-610.
26. Rice, W., Shannon, J. M., Burton, F., and Fiedeldej, D. (1997). Expression of a brain-type cannabinoid receptor (CB1) in alveolar Type II cells in the lung: regulation by hydrocortisone. *Eur.J.Pharmacol.* 327: 227-232.
27. Shmist, Y. A., Goncharov, I., Eichler, M., Shneyvays, V. and others. (2006). Delta-9-tetrahydrocannabinol protects cardiac cells from hypoxia via CB2 receptor activation and nitric oxide production. *Mol.Cell Biochem.* 283: 75-83.
28. Wright, K., Rooney, N., Feeney, M., Tate, J. and others. (2005). Differential expression of cannabinoid receptors in the human colon: cannabinoids promote epithelial wound healing. *Gastroenterology.* 129: 437-453.
29. Marquez, L., Suarez, J., Iglesias, M., Bermudez-Silva, F. J. and others. (2009). Ulcerative colitis induces changes on the expression of the endocannabinoid system in the human colonic tissue. *PLoS.One.* 4: e6893.-
30. Linari, G., Agostini, S., Amadoro, G., Ciotti, M. T. and others. (2009). Involvement of cannabinoid CB1- and CB2-receptors in the modulation of exocrine pancreatic secretion. *Pharmacol.Res.* 59: 207-214.
31. Izzo, A. A. and Sharkey, K. A. (2010). Cannabinoids and the gut: new developments and emerging concepts. *Pharmacol.Ther.* 126: 21-38.
32. Purohit, V., Rapaka, R., and Shurtleff, D. (2010). Role of cannabinoids in the development of fatty liver (steatosis). *AAPS.J.* 12: 233-237.
33. Mallat, A., Teixeira-Clerc, F., Deveaux, V., Manin, S. and others. (2011). The endocannabinoid system as a key mediator during liver diseases: new insights and therapeutic openings. *Br.J.Pharmacol.* 163: 1432-1440.
34. Jenkin, K. A., McAinch, A. J., Grinfeld, E., and Hryciw, D. H. (2010). Role for cannabinoid receptors in human proximal tubular hypertrophy. *Cell Physiol Biochem.* 26: 879-886.
35. Gratzke, C., Streng, T., Park, A., Christ, G. and others. (2009). Distribution and function of cannabinoid receptors 1 and 2 in the rat, monkey and human bladder. *J.Urol.* 181: 1939-1948.

36. Tyagi, P., Tyagi, V., Yoshimura, N., and Chancellor, M. (2010). Functional role of cannabinoid receptors in urinary bladder. *Indian J.Urol.* 26: 26-35.
37. Karasu, T., Marczylo, T. H., Maccarrone, M., and Konje, J. C. (2011). The role of sex steroid hormones, cytokines and the endocannabinoid system in female fertility. *Hum.Reprod.Update.* 17: 347-361.
38. Idris, A. I. and Ralston, S. H. (2010). Cannabinoids and bone: friend or foe? *Calcif.Tissue Int.* 87: 285-297.
39. Watkins, B. A., Hutchins, H., Li, Y., and Seifert, M. F. (2010). The endocannabinoid signaling system: a marriage of PUFA and musculoskeletal health. *J.Nutr.Biochem.* 21: 1141-1152.
40. Richardson, D., Pearson, R. G., Kurian, N., Latif, M. L. and others. (2008). Characterisation of the cannabinoid receptor system in synovial tissue and fluid in patients with osteoarthritis and rheumatoid arthritis. *Arthritis Res.Ther.* 10: R43.-
41. Biro, T., Toth, B. I., Hasko, G., Paus, R. and others. (2009). The endocannabinoid system of the skin in health and disease: novel perspectives and therapeutic opportunities. *Trends Pharmacol.Sci.* 30: 411-420.
42. Mackie, K. (2008). Signaling via CNS cannabinoid receptors. *Mol.Cell Endocrinol.* 286: S60-S65.
43. Cabral, G. A. Marijuana and the immune system. *Marijuana and medicine.* Nahas, G. G., Sutin, K. M., Harvey, D. J., and Agurell, S. Totowa: Humana Press, 1999.
44. De Petrocellis, L., Ligresti, A., Moriello, A. S., Allara, M. and others. (2011). Effects of cannabinoids and cannabinoid-enriched Cannabis extracts on TRP channels and endocannabinoid metabolic enzymes. *Br.J.Pharmacol.* 163: 1479-1494.
45. Alger, B. E. (2004). Endocannabinoids: getting the message across. *Proc.Natl.Acad.Sci.U.S.A.* 101: 8512-8513.
46. Bisogno, T. (2008). Endogenous cannabinoids: structure and metabolism. *J.Neuroendocrinol.* 20 Suppl 1: 1-9.
47. Miller, L. K. and Devi, L. A. (2011). The highs and lows of cannabinoid receptor expression in disease: mechanisms and their therapeutic implications. *Pharmacol.Rev.* 63: 461-470.
48. Martin-Sanchez, E., Furukawa, T. A., Taylor, J., and Martin, J. L. (2009). Systematic review and meta-analysis of cannabis treatment for chronic pain. *Pain Med.* 10: 1353-1368.
49. Gowran, A., Noonan, J., and Campbell, V. A. (2011). The multiplicity of action of cannabinoids: implications for treating neurodegeneration. *CNS.Neurosci.Ther.* 17: 637-644.
50. Guindon, J., Lai, Y., Takacs, S. M., Bradshaw, H. B. and others. (2012). Alterations in endocannabinoid tone following chemotherapy-induced peripheral neuropathy: Effects of endocannabinoid deactivation inhibitors targeting fatty-acid amide hydrolase and monoacylglycerol lipase in comparison to reference analgesics following cisplatin treatment. *Pharmacol.Res.* 67: 94-109.
51. Russo, E. B. and Hohmann, A. G. Role of cannabinoids in pain management. *Comprehensive Treatment of Chronic Pain by Medical, Interventional, and Behavioral Approaches: The AMERICAN ACADEMY OF PAIN MEDICINE Textbook on Patient Management* . Deer, T. R. and Leong, M. S. New York: Springer, 2012.
52. Guzman, M. (2003). Cannabinoids: potential anticancer agents. *Nat.Rev.Cancer.* 3: 745-755.
53. Di Marzo, V., Bifulco, M., and De Petrocellis, L. (2004). The endocannabinoid system and its therapeutic exploitation. *Nat.Rev.Drug Discov.* 3: 771-784.
54. Bazzaz, F. A., Dusek, D., Seigler, D. S., and Haney, A. W. (1975). Photosynthesis and cannabinoid content of temperate and tropical populations of *Cannabis sativa*. *Biochemical Systematics and Ecology.* 3: 15-18.

55. Elsohly, M. A. and Slade, D. (2005). Chemical constituents of marijuana: the complex mixture of natural cannabinoids. *Life Sci.* 78: 539-548.
56. Zhu, H. J., Wang, J. S., Markowitz, J. S., Donovan, J. L. and others. (2006). Characterization of P-glycoprotein inhibition by major cannabinoids from marijuana. *J.Pharmacol.Exp.Ther.* 317: 850-857.
57. Balducci, C., Nervegna, G., and Cecinato, A. (2009). Evaluation of principal cannabinoids in airborne particulates. *Anal.Chim.Acta.* 641: 89-94.
58. Yamaori, S., Kushihara, M., Yamamoto, I., and Watanabe, K. (2010). Characterization of major phytocannabinoids, cannabidiol and cannabinol, as isoform-selective and potent inhibitors of human CYP1 enzymes. *Biochem.Pharmacol.* 79: 1691-1698.
59. Hillig, K. W. and Mahlberg, P. G. (2004). A chemotaxonomic analysis of cannabinoid variation in *Cannabis* (Cannabaceae). *Am.J.Bot.* 91: 966-975.
60. Mehmedic, Z., Chandra, S., Slade, D., Denham, H. and others. (2010). Potency Trends of Delta(9)-THC and Other Cannabinoids in Confiscated Cannabis Preparations from 1993 to 2008*. *J.Forensic Sci.* 55: 1209-1217.
61. Whittle, B. A. and Guy, G. W. Development of cannabis-based medicines: risk, benefit and serendipity. *The Medicinal Uses of Cannabis and Cannabinoids.* Guy, G. W., Whittle, B. A., and Robson, P. J. London: Pharmaceutical Press, 2004.
62. Huestis, M. A. (2007). Human cannabinoid pharmacokinetics. *Chem.Biodivers.* 4: 1770-1804.
63. Dussy, F. E., Hamberg, C., Luginbuhl, M., Schwertzmann, T. and others. (2005). Isolation of Delta9-THCA-A from hemp and analytical aspects concerning the determination of Delta9-THC in cannabis products. *Forensic Sci.Int.* 149: 3-10.
64. Ashton, C. H. (2001). Pharmacology and effects of cannabis: a brief review. *Br.J.Psychiatry.* 178: 101-106.
65. Health Canada. Product Information Sheet on Dried Marihuana (*Cannabis*). 2008.
66. Health Canada. Product information sheet on dried marihuana (*Cannabis* spp.). Health Canada website. Health Canada. 2005.
67. Fishedick, J., Van der Kooy, F., and Verpoorte, R. (2010). Cannabinoid receptor 1 binding activity and quantitative analysis of *Cannabis sativa* L. smoke and vapor. *Chem.Pharm.Bull.(Tokyo).* 58: 201-207.
68. Maertens, R. M., White, P. A., Rickert, W., Levasseur, G. and others. (2009). The genotoxicity of mainstream and sidestream marijuana and tobacco smoke condensates. *Chem.Res.Toxicol.* 22: 1406-1414.
69. International Agency for Research on Cancer. IARC monographs on the evaluation of carcinogenic risks to humans: Tobacco smoke and involuntary smoking. 2004.
70. Moir, D., Rickert, W. S., Levasseur, G., Larose, Y. and others. (2008). A comparison of mainstream and sidestream marijuana and tobacco cigarette smoke produced under two smoking conditions. *Chem.Res.Toxicol.* 21: 494-502.
71. Russo, E. B. and McPartland, J. M. (2003). Cannabis is more than simply delta(9)-tetrahydrocannabinol. *Psychopharmacology (Berl).* 165: 431-432.
72. Russo, E. B. (2011). Taming THC: potential cannabis synergy and phytocannabinoid-terpenoid entourage effects. *Br.J.Pharmacol.* 163: 1344-1364.

73. Baker, P. B., Taylor, B. J., and Gough, T. A. (1981). The tetrahydrocannabinol and tetrahydrocannabinolic acid content of cannabis products. *J.Pharm.Pharmacol.* 33: 369-372.
74. Garrett, E. R. and Hunt, C. A. (1974). Physicochemical properties, solubility, and protein binding of delta9-tetrahydrocannabinol. *J.Pharm.Sci.* 63: 1056-1064.
75. Fairbairn, J. W., Liebmann, J. A., and Rowan, M. G. (1976). The stability of Cannabis and its preparations on storage. *J.Pharm.Pharmacol.* 28: 1-7.
76. Thomas, B. F., Parker, V. L., Caddell, L. W., Jones, L. V. and others. Composition and stability of a standard marijuana cigarette. *Marihuana and medicine.* Nahas, C. G., Sutin, K. M., Harvey, D. J., and Agurell, S. Totowa, New Jersey: Humana Press, 1999.
77. Govaerts, S. J., Hermans, E., and Lambert, D. M. (2004). Comparison of cannabinoid ligands affinities and efficacies in murine tissues and in transfected cells expressing human recombinant cannabinoid receptors. *Eur.J.Pharm.Sci.* 23: 233-243.
78. Pertwee, R. G. (2010). Receptors and channels targeted by synthetic cannabinoid receptor agonists and antagonists. *Curr.Med.Chem.* 17: 1360-1381.
79. Hajos, N., Ledent, C., and Freund, T. F. (2001). Novel cannabinoid-sensitive receptor mediates inhibition of glutamatergic synaptic transmission in the hippocampus. *Neuroscience.* 106: 1-4.
80. Darmani, N. A., Janoyan, J. J., Crim, J., and Ramirez, J. (2007). Receptor mechanism and antiemetic activity of structurally-diverse cannabinoids against radiation-induced emesis in the least shrew. *Eur.J.Pharmacol.* 563: 187-196.
81. Abrahamov, A., Abrahamov, A., and Mechoulam, R. (1995). An efficient new cannabinoid antiemetic in pediatric oncology. *Life Sci.* 56: 2097-2102.
82. Izzo, A. A., Borrelli, F., Capasso, R., Di, Marzo, V and others. (2009). Non-psychotropic plant cannabinoids: new therapeutic opportunities from an ancient herb. *Trends Pharmacol.Sci.* 30: 515-527.
83. Institute of Medicine. *Cannabinoids and animal physiology. Marijuana and medicine: Assessing the science base.* Joy, J. E., Watson, S. J., and Benson, J. A. Washington, DC: National Academy Press, 1999.
84. Musty, R. E. *Natural cannabinoids: interactions and effects. The medicinal uses of cannabis and cannabinoids.* Guy, G. W., Whittle, B. A., and Robson, P. J. London: Pharmaceutical Press, 2004.
85. Ruhaak, L. R., Felth, J., Karlsson, P. C., Raftar, J. J. and others. (2011). Evaluation of the cyclooxygenase inhibiting effects of six major cannabinoids isolated from *Cannabis sativa*. *Biol.Pharm.Bull.* 34: 774-778.
86. Cascio, M. G., Gauson, L. A., Stevenson, L. A., Ross, R. A. and others. (2010). Evidence that the plant cannabinoid cannabigerol is a highly potent alpha2-adrenoceptor agonist and moderately potent 5HT1A receptor antagonist. *Br.J.Pharmacol.* 159: 129-141.
87. Brown, A. J. (2007). Novel cannabinoid receptors. *Br.J.Pharmacol.* 152: 567-575.
88. Parker, L. A., Rock, E., and Limebeer, C. (2010). Regulation of nausea and vomiting by cannabinoids. *Br.J.Pharmacol.* 163: 1411-1422.
89. Thomas, A., Stevenson, L. A., Wease, K. N., Price, M. R. and others. (2005). Evidence that the plant cannabinoid Delta9-tetrahydrocannabivarin is a cannabinoid CB1 and CB2 receptor antagonist. *Br.J.Pharmacol.* 146: 917-926.

90. Bolognini, D., Costa, B., Maione, S., Comelli, F. and others. (2010). The plant cannabinoid Delta9-tetrahydrocannabinol can decrease signs of inflammation and inflammatory pain in mice. *Br.J.Pharmacol.* 160: 677-687.
91. Hill, A. J., Weston, S. E., Jones, N. A., Smith, I. and others. (2010). Delta-Tetrahydrocannabinol suppresses in vitro epileptiform and in vivo seizure activity in adult rats. *Epilepsia.* 51: 1522-1532.
92. Riedel, G., Fadda, P., McKillop-Smith, S., Pertwee, R. G. and others. (2009). Synthetic and plant-derived cannabinoid receptor antagonists show hypophagic properties in fasted and non-fasted mice. *Br.J.Pharmacol.* 156: 1154-1166.
93. Pertwee, R. G. (2005). Pharmacological actions of cannabinoids. *Handb.Exp.Pharmacol.* 1-51.
94. Jones, G. and Pertwee, R. G. (1972). A metabolic interaction in vivo between cannabidiol and 1 - tetrahydrocannabinol. *Br.J.Pharmacol.* 45: 375-377.
95. Karniol, I. G., Shirakawa, I., Kasinski, N., Pfeferman, A. and others. (1974). Cannabidiol interferes with the effects of delta 9 - tetrahydrocannabinol in man. *Eur.J.Pharmacol.* 28: 172-177.
96. Zuardi, A. W., Finkelfarb, E., Bueno, O. F., Musty, R. E. and others. (1981). Characteristics of the stimulus produced by the mixture of cannabidiol with delta 9-tetrahydrocannabinol. *Arch.Int.Pharmacodyn. Ther.* 249: 137-146.
97. Zuardi, A. W., Shirakawa, I., Finkelfarb, E., and Karniol, I. G. (1982). Action of cannabidiol on the anxiety and other effects produced by delta 9-THC in normal subjects. *Psychopharmacology (Berl).* 76: 245-250.
98. Zuardi, A. W. and Karniol, I. G. (1983). Effects on variable-interval performance in rats of delta 9-tetrahydrocannabinol and cannabidiol, separately and in combination. *Braz.J.Med.Biol.Res.* 16: 141-146.
99. Reid, M. J. and Bornheim, L. M. (2001). Cannabinoid-induced alterations in brain disposition of drugs of abuse. *Biochem.Pharmacol.* 61: 1357-1367.
100. Fadda, P., Robinson, L., Fratta, W., Pertwee, R. G. and others. (2004). Differential effects of THC- or CBD-rich cannabis extracts on working memory in rats. *Neuropharmacology.* 47: 1170-1179.
101. Ilan, A. B., Gevins, A., Coleman, M., Elsohly, M. A. and others. (2005). Neurophysiological and subjective profile of marijuana with varying concentrations of cannabinoids. *Behav.Pharmacol.* 16: 487-496.
102. Nadulski, T., Pragst, F., Weinberg, G., Roser, P. and others. (2005). Randomized, double-blind, placebo-controlled study about the effects of cannabidiol (CBD) on the pharmacokinetics of Delta9-tetrahydrocannabinol (THC) after oral application of THC versus standardized cannabis extract. *Ther.Drug Monit.* 27: 799-810.
103. Varvel, S. A., Wiley, J. L., Yang, R., Bridgen, D. T. and others. (2006). Interactions between THC and cannabidiol in mouse models of cannabinoid activity. *Psychopharmacology (Berl).* 186: 226-234.
104. Fusar-Poli, P., Crippa, J. A., Bhattacharyya, S., Borgwardt, S. J. and others. (2009). Distinct effects of {delta}9-tetrahydrocannabinol and cannabidiol on neural activation during emotional processing. *Arch.Gen.Psychiatry.* 66: 95-105.
105. Bhattacharyya, S., Morrison, P. D., Fusar-Poli, P., Martin-Santos, R. and others. (2010). Opposite effects of delta-9-tetrahydrocannabinol and cannabidiol on human brain function and psychopathology. *Neuropsychopharmacology.* 35: 764-774.
106. Winton-Brown, T. T., Allen, P., Bhattacharyya, S., Borgwardt, S. J. and others. (2011). Modulation of auditory and visual processing by delta-9-tetrahydrocannabinol and cannabidiol: an fMRI study. *Neuropsychopharmacology.* 36: 1340-1348.

107. Karschner, E. L., Darwin, W. D., McMahon, R. P., Liu, F. and others. (2011). Subjective and physiological effects after controlled Sativex and oral THC administration. *Clin.Pharmacol.Ther.* 89: 400-407.
108. Karschner, E. L., Darwin, W. D., Goodwin, R. S., Wright, S. and others. (2011). Plasma cannabinoid pharmacokinetics following controlled oral delta9-tetrahydrocannabinol and oromucosal cannabis extract administration. *Clin.Chem.* 57: 66-75.
109. Zuardi, A. W., Hallak, J. E., and Crippa, J. A. (2012). Interaction between cannabidiol (CBD) and (9)-tetrahydrocannabinol (THC): influence of administration interval and dose ratio between the cannabinoids. *Psychopharmacology (Berl)*. 219: 247-249.
110. Klein, C., Karanges, E., Spiro, A., Wong, A. and others. (2011). Cannabidiol potentiates Delta(9)-tetrahydrocannabinol (THC) behavioural effects and alters THC pharmacokinetics during acute and chronic treatment in adolescent rats. *Psychopharmacology (Berl)*. 218: 443-457.
111. Wachtel, S. R., Elsohly, M. A., Ross, S. A., Ambre, J. and others. (2002). Comparison of the subjective effects of Delta(9)-tetrahydrocannabinol and marijuana in humans. *Psychopharmacology (Berl)*. 161: 331-339.
112. Johnson, J. R., Burnell-Nugent, M., Lossignol, D., Ganay-Motan, E. D. and others. (2010). Multicenter, double-blind, randomized, placebo-controlled, parallel-group study of the efficacy, safety, and tolerability of THC:CBD extract and THC extract in patients with intractable cancer-related pain. *J.Pain Symptom.Manage.* 39: 167-179.
113. Schubart, C. D., Sommer, I. E., van Gastel, W. A., Goetgebuer, R. L. and others. (2011). Cannabis with high cannabidiol content is associated with fewer psychotic experiences. *Schizophr.Res.* 130: 216-221.
114. Kumar, R. N., Chambers, W. A., and Pertwee, R. G. (2001). Pharmacological actions and therapeutic uses of cannabis and cannabinoids. *Anaesthesia.* 56: 1059-1068.
115. British Medical Association. Therapeutic uses of cannabis. Amsterdam: Harwood Academic Publishers, 1997.
116. Hill, M. N., Froese, L. M., Morrish, A. C., Sun, J. C. and others. (2006). Alterations in behavioral flexibility by cannabinoid CB1 receptor agonists and antagonists. *Psychopharmacology (Berl)*. 187: 245-259.
117. Zuurman, L., Ippel, A. E., Moin, E., and van Gerven, J. M. (2009). Biomarkers for the effects of cannabis and THC in healthy volunteers. *Br.J.Clin.Pharmacol.* 67: 5-21.
118. Institute of Medicine. First, do no harm: consequences of marijuana use and abuse. Marijuana and medicine: Assessing the science base. Joy, J. E., Watson, S. J., and Benson, J. A. Washington, DC: National Academy Press, 1999.
119. Beaconsfield, P., Ginsburg, J., and Rainsbury, R. (1972). Marijuana smoking. Cardiovascular effects in man and possible mechanisms. *N.Engl.J.Med.* 287: 209-212.
120. Perez-Reyes, M. (1990). Marijuana smoking: factors that influence the bioavailability of tetrahydrocannabinol. *NIDA Res.Monogr.* 99: 42-62.
121. Aryana, A. and Williams, M. A. (2007). Marijuana as a trigger of cardiovascular events: speculation or scientific certainty? *Int.J.Cardiol.* 118: 141-144.
122. Hall, W. and Solowij, N. (1998). Adverse effects of cannabis. *Lancet.* 352: 1611-1616.
123. Ameri, A. (1999). The effects of cannabinoids on the brain. *Prog.Neurobiol.* 58: 315-348.
124. Barnett, G., Licko, V., and Thompson, T. (1985). Behavioral pharmacokinetics of marijuana. *Psychopharmacology (Berl)*. 85: 51-56.

125. Kelly, T. H., Foltin, R. W., and Fischman, M. W. (1993). Effects of smoked marijuana on heart rate, drug ratings and task performance by humans. *Behav.Pharmacol.* 4: 167-178.
126. Fant, R. V., Heishman, S. J., Bunker, E. B., and Pickworth, W. B. (1998). Acute and residual effects of marijuana in humans. *Pharmacol.Biochem.Behav.* 60: 777-784.
127. Hollister, L. E. (1998). Health aspects of cannabis: revisited. *Int.J.Neuropsychopharmacol.* 1: 71-80.
128. Miller, L. L. *Marihuana: Acute effects on human memory. Marihuana and medicine.* Nahas, C. G., Sutin, K. M., Harvey, D. J., and Agurell, S. Totowa, New Jersey: Humana Press, 1999.
129. Hart, C. L., Ilan, A. B., Gevins, A., Gunderson, E. W. and others. (2010). Neurophysiological and cognitive effects of smoked marijuana in frequent users. *Pharmacol.Biochem.Behav.* 96: 333-341.
130. Crean, R. D., Crane, N. A., and Mason, B. J. (2011). An Evidence Based Review of Acute and Long-Term Effects of Cannabis Use on Executive Cognitive Functions. *J.Addict.Med.* 5: 1-8.
131. Heishman, S. J., Huestis, M. A., Henningfield, J. E., and Cone, E. J. (1990). Acute and residual effects of marijuana: profiles of plasma THC levels, physiological, subjective, and performance measures. *Pharmacol.Biochem.Behav.* 37: 561-565.
132. Curran, H. V., Brignell, C., Fletcher, S., Middleton, P. and others. (2002). Cognitive and subjective dose-response effects of acute oral Delta 9-tetrahydrocannabinol (THC) in infrequent cannabis users. *Psychopharmacology (Berl)*. 164: 61-70.
133. Ramaekers, J. G., Moeller, M. R., van, Ruitenbeek P., Theunissen, E. L. and others. (2006). Cognition and motor control as a function of Delta9-THC concentration in serum and oral fluid: limits of impairment. *Drug Alcohol Depend.* 85: 114-122.
134. O'Kane, C. J., Tutt, D. C., and Bauer, L. A. (2002). Cannabis and driving: a new perspective. *Emerg.Med.(Fremantle.)*. 14: 296-303.
135. Hanstee, R. W., Miller, R. D., Lonero, L., Reid, L. D. and others. (1976). Effects of cannabis and alcohol on automobile driving and psychomotor tracking. *Ann.N.Y.Acad.Sci.* 282: 240-256.
136. Leirer, V. O., Yesavage, J. A., and Morrow, D. G. (1991). Marijuana carry-over effects on aircraft pilot performance. *Aviat.Space Environ.Med.* 62: 221-227.
137. Smiley, A. *Marijuana: On-road and driving-simulator studies. The Health Effects of Cannabis.* Kalant, H., Corrigan, W., Hall, W., and Smart, R. Toronto, Canada: Centre of Addiction and Mental Health, 1999.
138. van Os, J., Bak, M., Hanssen, M., Bijl, R. V. and others. (2002). Cannabis use and psychosis: a longitudinal population-based study. *Am.J.Epidemiol.* 156: 319-327.
139. D'Souza, D. C., Abi-Saab, W. M., Madonick, S., Forselius-Bielen, K. and others. (2005). Delta-9-tetrahydrocannabinol effects in schizophrenia: implications for cognition, psychosis, and addiction. *Biol.Psychiatry.* 57: 594-608.
140. D'Souza, D. C., Perry, E., MacDougall, L., Ammerman, Y. and others. (2004). The psychotomimetic effects of intravenous delta-9-tetrahydrocannabinol in healthy individuals: implications for psychosis. *Neuropsychopharmacology.* 29: 1558-1572.
141. Moore, T. H., Zammit, S., Lingford-Hughes, A., Barnes, T. R. and others. (2007). Cannabis use and risk of psychotic or affective mental health outcomes: a systematic review. *Lancet.* 370: 319-328.

142. Abrams, D. I., Jay, C. A., Shade, S. B., Vizoso, H. and others. (2007). Cannabis in painful HIV-associated sensory neuropathy: a randomized placebo-controlled trial. *Neurology*. 68: 515-521.
143. Henquet, C., van Os J., Kuepper, R., Delespaul, P. and others. (2010). Psychosis reactivity to cannabis use in daily life: an experience sampling study. *Br.J.Psychiatry*. 196: 447-453.
144. Korver, N., Nieman, D. H., Becker, H. E., van de Fliert, J. R. and others. (2010). Symptomatology and neuropsychological functioning in cannabis using subjects at ultra-high risk for developing psychosis and healthy controls. *Aust.N.Z.J.Psychiatry*. 44: 230-236.
145. Favrat, B., Menetrey, A., Augsburger, M., Rothuizen, L. E. and others. (2005). Two cases of "cannabis acute psychosis" following the administration of oral cannabis. *BMC.Psychiatry*. 5: 17-22.
146. Henquet, C., Krabbendam, L., Spauwen, J., Kaplan, C. and others. (2005). Prospective cohort study of cannabis use, predisposition for psychosis, and psychotic symptoms in young people. *BMJ*. 330: 11-15.
147. Crippa, J. A., Zuardi, A. W., Martin-Santos, R., Bhattacharyya, S. and others. (2009). Cannabis and anxiety: a critical review of the evidence. *Hum.Psychopharmacol*. 24: 515-523.
148. van Rossum, I., Boomsma, M., Tenback, D., Reed, C. and others. (2009). Does cannabis use affect treatment outcome in bipolar disorder? A longitudinal analysis. *J.Nerv.Ment.Dis*. 197: 35-40.
149. Ongur, D., Lin, L., and Cohen, B. M. (2009). Clinical characteristics influencing age at onset in psychotic disorders. *Compr.Psychiatry*. 50: 13-19.
150. De Hert, M., Wampers, M., Jendricko, T., Franic, T. and others. (2011). Effects of cannabis use on age at onset in schizophrenia and bipolar disorder. *Schizophr.Res*. 126: 270-276.
151. Harder, S. and Rietbrock, S. (1997). Concentration-effect relationship of delta-9-tetrahydrocannabinol and prediction of psychotropic effects after smoking marijuana. *Int.J.Clin.Pharmacol.Ther*. 35: 155-159.
152. Johns, A. (2001). Psychiatric effects of cannabis. *Br.J.Psychiatry*. 178: 116-122.
153. Boyce, A. and McArdle, P. (2007). Long-term effects of cannabis. *Pediatrics and child health*. 18: 37-41.
154. Zammit, S., Moore, T. H., Lingford-Hughes, A., Barnes, T. R. and others. (2008). Effects of cannabis use on outcomes of psychotic disorders: systematic review. *Br.J.Psychiatry*. 193: 357-363.
155. Moreira, F. A., Grieb, M., and Lutz, B. (2009). Central side-effects of therapies based on CB1 cannabinoid receptor agonists and antagonists: focus on anxiety and depression. *Best.Pract.Res.Clin.Endocrinol.Metab*. 23: 133-144.
156. Vandrey, R. and Haney, M. (2009). Pharmacotherapy for cannabis dependence: how close are we? *CNS.Drugs*. 23: 543-553.
157. Lal, S., Prasad, N., Ryan, M., Tangri, S. and others. (2011). Cannabis use amongst patients with inflammatory bowel disease. *Eur.J.Gastroenterol.Hepatol*. 23: 891-896.
158. Fiz, J., Duran, M., Capella, D., Carbonell, J. and others. (2011). Cannabis use in patients with fibromyalgia: effect on symptoms relief and health-related quality of life. *PLoS.One*. 6: e18440.-
159. Tramer, M. R., Carroll, D., Campbell, F. A., Reynolds, D. J. and others. (2001). Cannabinoids for control of chemotherapy induced nausea and vomiting: quantitative systematic review. *BMJ*. 323: 16-21.
160. Corcoran, C. M., Kimhy, D., Stanford, A., Khan, S. and others. (2008). Temporal association of cannabis use with symptoms in individuals at clinical high risk for psychosis. *Schizophr.Res*. 106: 286-293.

161. Schierenbeck, T., Riemann, D., Berger, M., and Hornyak, M. (2008). Effect of illicit recreational drugs upon sleep: cocaine, ecstasy and marijuana. *Sleep Med.Rev.* 12: 381-389.
162. Hunault, C. C., Mensinga, T. T., Bocker, K. B., Schipper, C. M. and others. (2009). Cognitive and psychomotor effects in males after smoking a combination of tobacco and cannabis containing up to 69 mg delta-9-tetrahydrocannabinol (THC). *Psychopharmacology (Berl)*. 204: 85-94.
163. Scott, J., Martin, G., Bor, W., Sawyer, M. and others. (2009). The prevalence and correlates of hallucinations in Australian adolescents: results from a national survey. *Schizophr.Res.* 107: 179-185.
164. Page, S. A., Verhoef, M. J., Stebbins, R. A., Metz, L. M. and others. (2003). Cannabis use as described by people with multiple sclerosis. *Can.J.Neurol.Sci.* 30: 201-205.
165. Clark, A. J., Ware, M. A., Yazer, E., Murray, T. J. and others. (2004). Patterns of cannabis use among patients with multiple sclerosis. *Neurology*. 62: 2098-2100.
166. Haney, M., Rabkin, J., Gunderson, E., and Foltin, R. W. (2005). Dronabinol and marijuana in HIV(+) marijuana smokers: acute effects on caloric intake and mood. *Psychopharmacology (Berl)*. 181: 170-178.
167. Haney, M., Gunderson, E. W., Rabkin, J., Hart, C. L. and others. (2007). Dronabinol and marijuana in HIV-positive marijuana smokers. Caloric intake, mood, and sleep. *J.Acquir.Immune.Defic.Syndr.* 45: 545-554.
168. Wilsey, B., Marcotte, T., Tsodikov, A., Millman, J. and others. (2008). A randomized, placebo-controlled, crossover trial of cannabis cigarettes in neuropathic pain. *J.Pain*. 9: 506-521.
169. Sewell, R. A., Poling, J., and Sofuoglu, M. (2009). The effect of cannabis compared with alcohol on driving. *Am.J.Addict.* 18: 185-193.
170. Brannness, J. G., Khiabani, H. Z., and Morland, J. (2010). Impairment due to cannabis and ethanol: clinical signs and additive effects. *Addiction*. 105: 1080-1087.
171. Ronen, A., Chassidim, H. S., Gershon, P., Parnet, Y. and others. (2010). The effect of alcohol, THC and their combination on perceived effects, willingness to drive and performance of driving and non-driving tasks. *Accid.Anal.Prev.* 42: 1855-1865.
172. Ware, M. A., Wang, T., Shapiro, S., Robinson, A. and others. (2010). Smoked cannabis for chronic neuropathic pain: a randomized controlled trial. *CMAJ*. 182: E694-E701.
173. Lynch, M. E. and Campbell, F. (2011). Cannabinoids for Treatment of Chronic Non-Cancer Pain; a Systematic Review of Randomized Trials. *Br.J.Clin.Pharmacol.* 72: 735-744.
174. Abbott Products Inc. Marinol Product Monograph. 2010.
175. Asbridge, M., Hayden, J. A., and Cartwright, J. L. (2012). Acute cannabis consumption and motor vehicle collision risk: systematic review of observational studies and meta-analysis. *BMJ*. 344: e536-
176. Downey, L. A., King, R., Papafotiou, K., Swann, P. and others. (2012). The effects of cannabis and alcohol on simulated driving: Influences of dose and experience. *Accid.Anal.Prev.* 50: 879-886.
177. Elvik, R. (2012). Risk of road accident associated with the use of drugs: A systematic review and meta-analysis of evidence from epidemiological studies (in press). *Accid.Anal.Prev.*
178. Honarmand, K., Tierney, M. C., O'Connor, P., and Feinstein, A. (2011). Effects of cannabis on cognitive function in patients with multiple sclerosis. *Neurology*. 76: 1153-1160.

179. Pope, H. G., Jr., Gruber, A. J., Hudson, J. L., Huestis, M. A. and others. (2002). Cognitive measures in long-term cannabis users. *J.Clin.Pharmacol.* 42: 41S-47S.
180. Solowij, N., Stephens, R. S., Roffman, R. A., Babor, T. and others. (2002). Cognitive functioning of long-term heavy cannabis users seeking treatment. *JAMA.* 287: 1123-1131.
181. Ilan, A. B., Smith, M. E., and Gevins, A. (2004). Effects of marijuana on neurophysiological signals of working and episodic memory. *Psychopharmacology (Berl).* 176: 214-222.
182. Williamson, E. M. and Evans, F. J. (2000). Cannabinoids in clinical practice. *Drugs.* 60: 1303-1314.
183. Weinstein, A., Brickner, O., Lerman, H., Greemland, M. and others. (2008). Brain imaging study of the acute effects of Delta9-tetrahydrocannabinol (THC) on attention and motor coordination in regular users of marijuana. *Psychopharmacology (Berl).* 196: 119-131.
184. Greenwald, M. K. and Stitzer, M. L. (2000). Antinociceptive, subjective and behavioral effects of smoked marijuana in humans. *Drug Alcohol Depend.* 59: 261-275.
185. Wallace, M., Schulteis, G., Atkinson, J. H., Wolfson, T. and others. (2007). Dose-dependent effects of smoked cannabis on capsaicin-induced pain and hyperalgesia in healthy volunteers. *Anesthesiology.* 107: 785-796.
186. Ellis, R. J., Toperoff, W., Vaida, F., van den Brande, G. and others. (2009). Smoked medicinal cannabis for neuropathic pain in HIV: a randomized, crossover clinical trial. *Neuropsychopharmacology.* 34: 672-680.
187. Abrams, D. I., Couey, P., Shade, S. B., Kelly, M. E. and others. (2011). Cannabinoid-opioid interaction in chronic pain. *Clin.Pharmacol.Ther.* 90: 844-851.
188. Corey-Bloom, J., Wolfson, T., Gamst, A., Jin, S. and others. (2012). Smoked cannabis for spasticity in multiple sclerosis: a randomized, placebo-controlled trial. *CMAJ.* 184: 1143-1150.
189. Lahat, A., Lang, A., and Ben-Horin, S. (2012). Impact of cannabis treatment on the quality of life, weight and clinical disease activity in inflammatory bowel disease patients: a pilot prospective study. *Digestion.* 85: 1-8.
190. Lynch, M. E., Young, J., and Clark, A. J. (2006). A case series of patients using medicinal marijuana for management of chronic pain under the Canadian Marijuana Medical Access Regulations. *J.Pain Symptom.Manage.* 32: 497-501.
191. Musty, R. and Rossi, R. (2001). Effects of smoked cannabis and oral delta-9-tetrahydrocannabinol on nausea and emesis after cancer chemotherapy: A review of state clinical trials. *Journal of Cannabis Therapeutics.* 1: 29-42.
192. Soderpalm, A. H., Schuster, A., and de Wit H. (2001). Antiemetic efficacy of smoked marijuana: subjective and behavioral effects on nausea induced by syrup of ipecac. *Pharmacol.Biochem.Behav.* 69: 343-350.
193. Bedi, G., Foltin, R. W., Gunderson, E. W., Rabkin, J. and others. (2010). Efficacy and tolerability of high-dose dronabinol maintenance in HIV-positive marijuana smokers: a controlled laboratory study. *Psychopharmacology (Berl).* 212: 675-686.
194. Sannarangappa, V. and Tan, C. (2009). Cannabinoid hyperemesis. *Intern.Med.J.* 39: 777-778.
195. Donnino, M. W., Cocchi, M. N., Miller, J., and Fisher, J. (2011). Cannabinoid hyperemesis: a case series. *J.Emerg.Med.* 40: e63-e66.
196. Sullivan, S. (2010). Cannabinoid hyperemesis. *Can.J.Gastroenterol.* 24: 284-285.

197. Miller, J. B., Walsh, M., Patel, P. A., Rogan, M. and others. (2010). Pediatric cannabinoid hyperemesis: two cases. *Pediatr. Emerg. Care.* 26: 919-920.
198. Patterson, D. A., Smith, E., Monahan, M., Medvecz, A. and others. (2010). Cannabinoid hyperemesis and compulsive bathing: a case series and paradoxical pathophysiological explanation. *J. Am. Board Fam. Med.* 23: 790-793.
199. Choung, R. S., Locke, G. R., III, Lee, R. M., Schleck, C. D. and others. (2012). Cyclic vomiting syndrome and functional vomiting in adults: association with cannabinoid use in males. *Neurogastroenterol. Motil.* 24: 20-6, e1.
200. Francis, H. (2011). Emerging role of chronic cannabis usage and hyperemesis syndrome. *South. Med. J.* 104: 665-
201. Schmid, S. M., Lapaire, O., Huang, D. J., Jurgens, F. E. and others. (2011). Cannabinoid hyperemesis syndrome: an underreported entity causing nausea and vomiting of pregnancy. *Arch. Gynecol. Obstet.* 284: 1095-1097.
202. Wallace, E. A., Andrews, S. E., Garmany, C. L., and Jelley, M. J. (2011). Cannabinoid hyperemesis syndrome: literature review and proposed diagnosis and treatment algorithm. *South. Med. J.* 104: 659-664.
203. Simonetto, D. A., Oxentenko, A. S., Herman, M. L., and Szostek, J. H. (2012). Cannabinoid hyperemesis: a case series of 98 patients. *Mayo Clin. Proc.* 87: 114-119.
204. Wild, K. and Wilson, H. (2012). Cannabinoid hyperemesis. *Emerg. Med. J.* 29: 67-69.
205. Foltin, R. W., Fischman, M. W., and Byrne, M. F. (1988). Effects of smoked marijuana on food intake and body weight of humans living in a residential laboratory. *Appetite.* 11: 1-14.
206. Mattes, R. D., Engelman, K., Shaw, L. M., and Elsohly, M. A. (1994). Cannabinoids and appetite stimulation. *Pharmacol. Biochem. Behav.* 49: 187-195.
207. Haney, M., Ward, A. S., Comer, S. D., Foltin, R. W. and others. (1999). Abstinence symptoms following oral THC administration to humans. *Psychopharmacology (Berl).* 141: 385-394.
208. Sutton, J. R. and Daeninck, P. (2006). Cannabinoids in the management of intractable chemotherapy-induced nausea and vomiting and cancer-related pain. *J. Support. Oncol.* 4: 531-535.
209. Pisanti, S., Malfitano, A. M., Grimaldi, C., Santoro, A. and others. (2009). Use of cannabinoid receptor agonists in cancer therapy as palliative and curative agents. *Best. Pract. Res. Clin. Endocrinol. Metab.* 23: 117-131.
210. Lichtman, A. H. and Martin, B. R. (2005). Cannabinoid tolerance and dependence. *Handb. Exp. Pharmacol.* 691-717.
211. Gonzalez, S., Cebeira, M., and Fernandez-Ruiz, J. (2005). Cannabinoid tolerance and dependence: a review of studies in laboratory animals. *Pharmacol. Biochem. Behav.* 81: 300-318.
212. Jones, R. T., Benowitz, N., and Bachman, J. (1976). Clinical studies of cannabis tolerance and dependence. *Ann. N. Y. Acad. Sci.* 282: 221-239.
213. Compton, D. R., Dewey, W. L., and Martin, B. R. (1990). Cannabis dependence and tolerance production. *Adv. Alcohol Subst. Abuse.* 9: 129-147.
214. Pertwee, R. Tolerance to and dependence on psychotropic cannabinoids. *The biological bases of drug tolerance and dependence.* Pratt, J. London: Academic Press, 1991.
215. De Vry, J., Jentzsch, K. R., Kuhl, E., and Eckel, G. (2004). Behavioral effects of cannabinoids show differential sensitivity to cannabinoid receptor blockade and tolerance development. *Behav. Pharmacol.* 15: 1-12.

216. D'Souza, D. C., Ranganathan, M., Braley, G., Gueorguieva, R. and others. (2008). Blunted psychotomimetic and amnesic effects of delta-9-tetrahydrocannabinol in frequent users of cannabis. *Neuropsychopharmacology*. 33: 2505-2516.
217. Rog, D. J., Nurmikko, T. J., and Young, C. A. (2007). Oromucosal delta9-tetrahydrocannabinol/cannabidiol for neuropathic pain associated with multiple sclerosis: an uncontrolled, open-label, 2-year extension trial. *Clin.Ther.* 29: 2068-2079.
218. Serpell, M. G., Notcutt, W., and Collin, C. (2012). Sativex long-term use: an open-label trial in patients with spasticity due to multiple sclerosis. *J.Neurol.* 260: 285-295.
219. Hall, W. and Solowij, N. The adverse health and psychological consequences of cannabis dependence. Cannabis dependence. Roffman, R. A. and Stephens, R. S. Cambridge: Cambridge University Press, 2006.
220. Kalant, H. (2004). Adverse effects of cannabis on health: an update of the literature since 1996. *Prog.Neuropsychopharmacol.Biol.Psychiatry.* 28: 849-863.
221. Cooper, Z. D. and Haney, M. (2008). Cannabis reinforcement and dependence: role of the cannabinoid CB1 receptor. *Addict.Biol.* 13: 188-195.
222. Allsop, D. J., Norberg, M. M., Copeland, J., Fu, S. and others. (2011). The Cannabis Withdrawal Scale development: patterns and predictors of cannabis withdrawal and distress. *Drug Alcohol Depend.* 119: 123-129.
223. Renault, P. F., Schuster, C. R., Heinrich, R., and Freeman, D. X. (1971). Marihuana: standardized smoke administration and dose effect curves on heart rate in humans. *Science.* 174: 589-591.
224. Clark, S. C., Greene, C., Karr, G. W., MacCannell, K. L. and others. (1974). Cardiovascular effects of marihuana in man. *Can.J.Physiol Pharmacol.* 52: 706-719.
225. O'Leary, D. S., Block, R. I., Koeppe, J. A., Flaum, M. and others. (2002). Effects of smoking marijuana on brain perfusion and cognition. *Neuropsychopharmacology.* 26: 802-816.
226. Trouve, R. and Nahas, G. Cardiovascular effects of marihuana and cannabinoids. *Marihuana and medicine.* Nahas, C. G., Sutin, K. M., Harvey, D. J., and Agurell, S. Totowa, New Jersey: Humana Press, 1999.
227. Jones, R. T. (2002). Cardiovascular system effects of marijuana. *J.Clin.Pharmacol.* 42: 58S-63S.
228. Hollister, L. E. (1986). Health aspects of cannabis. *Pharmacol.Rev.* 38: 1-20.
229. Miller, R. H., Dhingra, R. C., Kanakis, C., Jr., Leon, F. and others. (1977). The electrophysiological effects of delta-9-tetrahydrocannabinol (cannabis) on cardiac conduction in man. *Am.Heart J.* 94: 740-747.
230. Lindsay, A. C., Foale, R. A., Warren, O., and Henry, J. A. (2005). Cannabis as a precipitant of cardiovascular emergencies. *Int.J.Cardiol.* 104: 230-232.
231. Beaconsfield, P. (1974). Some cardiovascular effects of cannabis. *Am.Heart J.* 87: 143-146.
232. Mittleman, M. A., Lewis, R. A., Maclure, M., Sherwood, J. B. and others. (2001). Triggering myocardial infarction by marijuana. *Circulation.* 103: 2805-2809.
233. Mathew, R. J., Wilson, W. H., and Davis, R. (2003). Postural syncope after marijuana: a transcranial Doppler study of the hemodynamics. *Pharmacol.Biochem.Behav.* 75: 309-318.
234. Gorelick, D. A. and Hershman, S. J. (2006). Methods for clinical research involving cannabis administration. *Methods Mol.Med.* 123: 235-253.

235. Lundqvist, T. (2005). Cognitive consequences of cannabis use: comparison with abuse of stimulants and heroin with regard to attention, memory and executive functions. *Pharmacol.Biochem.Behav.* 81: 319-330.
236. Singh, N. N., Pan, Y., Muengtaweepansa, S., Geller, T. J. and others. (2012). Cannabis-related stroke: case series and review of literature. *J.Stroke Cerebrovasc.Dis.* 21: 555-560.
237. Renard, D., Taieb, G., Gras-Combe, G., and Labauge, P. (2012). Cannabis-related myocardial infarction and cardioembolic stroke. *J.Stroke Cerebrovasc.Dis.* 21: 82-83.
238. Sidney, S., Quesenberry Jr, C. P., Friedman, G. D., and Tekawa, I. S. (1997). Marijuana use and cancer incidence (California, United States). *Cancer Causes Control.* 8: 722-728.
239. Zhang, Z. F., Morgenstern, H., Spitz, M. R., Tashkin, D. P. and others. (1999). Marijuana use and increased risk of squamous cell carcinoma of the head and neck. *Cancer Epidemiol.Biomarkers Prev.* 8: 1071-1078.
240. Hashibe, M., Morgenstern, H., Cui, Y., Tashkin, D. P. and others. (2006). Marijuana use and the risk of lung and upper aerodigestive tract cancers: results of a population-based case-control study. *Cancer Epidemiol.Biomarkers Prev.* 15: 1829-1834.
241. Aldington, S., Harwood, M., Cox, B., Weatherall, M. and others. (2008). Cannabis use and risk of lung cancer: a case-control study. *Eur.Respir.J.* 31: 280-286.
242. Fligel, S. E., Roth, M. D., Kleerup, E. C., Barsky, S. H. and others. (1997). Tracheobronchial histopathology in habitual smokers of cocaine, marijuana, and/or tobacco. *Chest.* 112: 319-326.
243. Tetrault, J. M., Crothers, K., Moore, B. A., Mehra, R. and others. (2007). Effects of marijuana smoking on pulmonary function and respiratory complications: a systematic review. *Arch.Intern.Med.* 167: 221-228.
244. Bloom, J. W., Kaltenborn, W. T., Paoletti, P., Camilli, A. and others. (1987). Respiratory effects of non-tobacco cigarettes. *Br.Med.J.(Clin.Res.Ed).* 295: 1516-1518.
245. Tashkin, D. P., Coulson, A. H., Clark, V. A., Simmons, M. and others. (1987). Respiratory symptoms and lung function in habitual heavy smokers of marijuana alone, smokers of marijuana and tobacco, smokers of tobacco alone, and nonsmokers. *Am.Rev.Respir.Dis.* 135: 209-216.
246. Roth, M. D., Arora, A., Barsky, S. H., Kleerup, E. C. and others. (1998). Airway inflammation in young marijuana and tobacco smokers. *Am.J.Respir.Crit Care Med.* 157: 928-937.
247. Pletcher, M. J., Vittinghoff, E., Kalhan, R., Richman, J. and others. (2012). Association between marijuana exposure and pulmonary function over 20 years. *JAMA.* 307: 173-181.
248. Naftali, T., Lev, L. B., Yablecovitch, D., Half, E. and others. (2011). Treatment of Crohn's disease with cannabis: an observational study. *Isr.Med.Assoc.J.* 13: 455-458.
249. Patsenker, E., Stoll, M., Millonig, G., Agaimy, A. and others. (2011). Cannabinoid receptor type I modulates alcohol-induced liver fibrosis. *Mol.Med.* 17: 1285-1294.
250. Trebicka, J., Racz, I., Siegmund, S. V., Cara, E. and others. (2011). Role of cannabinoid receptors in alcoholic hepatic injury: steatosis and fibrogenesis are increased in CB2 receptor-deficient mice and decreased in CB1 receptor knockouts. *Liver Int.* 31: 860-870.
251. Reichenbach, V., Ros, J., Fernandez-Varo, G., Casals, G. and others. (2012). Prevention of fibrosis progression in CCl4-treated rats: role of the hepatic endocannabinoid and apelin systems. *J.Pharmacol.Exp.Ther.* 340: 629-637.

252. Sylvestre, D. L., Clements, B. J., and Malibu, Y. (2006). Cannabis use improves retention and virological outcomes in patients treated for hepatitis C. *Eur.J.Gastroenterol.Hepatol.* 18: 1057-1063.
253. Grant, P. and Gandhi, P. (2004). A case of cannabis-induced pancreatitis. *JOP.* 5: 41-43.
254. Wargo, K. A., Geveden, B. N., and McConnell, V. J. (2007). Cannabinoid-induced pancreatitis: a case series. *JOP.* 8: 579-583.
255. Bournet, B. and Buscaïl, L. (2008). [Cannabis: a rare cause of acute pancreatitis]. *Gastroenterol.Clin.Biol.* 32: 922-923.
256. Belze, O., Jr., Legras, A., Ehrmann, S., Garot, D. and others. (2011). Cannabis-induced acute pancreatitis. *Am.J.Emerg.Med.* 29: 131-134.
257. Cox, M. L., Haller, V. L., and Welch, S. P. (2007). Synergy between delta9-tetrahydrocannabinol and morphine in the arthritic rat. *Eur.J.Pharmacol.* 567: 125-130.
258. Smith, F. L., Fujimori, K., Lowe, J., and Welch, S. P. (1998). Characterization of delta9-tetrahydrocannabinol and anandamide antinociception in nonarthritic and arthritic rats. *Pharmacol.Biochem.Behav.* 60: 183-191.
259. Blake, D. R., Robson, P., Ho, M., Jubb, R. W. and others. (2006). Preliminary assessment of the efficacy, tolerability and safety of a cannabis-based medicine (Sativex) in the treatment of pain caused by rheumatoid arthritis. *Rheumatology.(Oxford).* 45: 50-52.
260. Schley, M., Legler, A., Skopp, G., Schmelz, M. and others. (2006). Delta-9-THC based monotherapy in fibromyalgia patients on experimentally induced pain, axon reflex flare, and pain relief. *Curr.Med.Res.Opin.* 22: 1269-1276.
261. Weber, J., Schley, M., Casutt, M., Gerber, H. and others. (2009). Tetrahydrocannabinol (Delta 9-THC) Treatment in Chronic Central Neuropathic Pain and Fibromyalgia Patients: Results of a Multicenter Survey. *Anesthesiol.Res.Pract.* 2009: 827290.-
262. Zajicek, J., Fox, P., Sanders, H., Wright, D. and others. (2003). Cannabinoids for treatment of spasticity and other symptoms related to multiple sclerosis (CAMS study): multicentre randomised placebo-controlled trial. *Lancet.* 362: 1517-1526.
263. Nogueira-Filho, Gda R., Cadide, T., Rosa, B. T., Neiva, T. G. and others. (2008). Cannabis sativa smoke inhalation decreases bone filling around titanium implants: a histomorphometric study in rats. *Implant.Dent.* 17: 461-470.
264. Tomida, I., Azuara-Blanco, A., House, H., Flint, M. and others. (2006). Effect of sublingual application of cannabinoids on intraocular pressure: a pilot study. *J.Glaucoma.* 15: 349-353.
265. Tomida, I., Pertwee, R. G., and zuara-Blanco, A. (2004). Cannabinoids and glaucoma. *Br.J.Ophthalmol.* 88: 708-713.
266. Tanasescu, R. and Constantinescu, C. S. (2010). Cannabinoids and the immune system: an overview. *Immunobiology.* 215: 588-597.
267. Nahas, G. G., Frick, H. C., Lattimer, J. K., Latour, C. and others. (2002). Pharmacokinetics of THC in brain and testis, male gametotoxicity and premature apoptosis of spermatozoa. *Hum.Psychopharmacol.* 17: 103-113.
268. Sadeu, J. C., Hughes, C. L., Agarwal, S., and Foster, W. G. (2010). Alcohol, drugs, caffeine, tobacco, and environmental contaminant exposure: reproductive health consequences and clinical implications. *Crit Rev.Toxicol.* 40: 633-652.

269. Gorzalka, B. B., Hill, M. N., and Chang, S. C. (2010). Male-female differences in the effects of cannabinoids on sexual behavior and gonadal hormone function. *Horm.Behav.* 58: 91-99.
270. Brown, T. T. and Dobs, A. S. (2002). Endocrine effects of marijuana. *J.Clin.Pharmacol.* 42: 90S-96S.
271. Shamloul, R. and Bella, A. J. (2011). Impact of cannabis use on male sexual health. *J.Sex Med.* 8: 971-975.
272. Agurell, S., Halldin, M., Lindgren, J. E., Ohlsson, A. and others. (1986). Pharmacokinetics and metabolism of delta 1-tetrahydrocannabinol and other cannabinoids with emphasis on man. *Pharmacol.Rev.* 38: 21-43.
273. Abrams, D. I., Vizoso, H. P., Shade, S. B., Jay, C. and others. (2007). Vaporization as a Smokeless Cannabis Delivery System: A Pilot Study. *Clin.Pharmacol.Ther.* 82: 572-578.
274. McClure, E. A., Stitzer, M. L., and Vandrey, R. (2012). Characterizing smoking topography of cannabis in heavy users. *Psychopharmacology (Berl)*. 220: 309-318.
275. Grotenhermen, F. (2003). Pharmacokinetics and pharmacodynamics of cannabinoids. *Clin.Pharmacokinet.* 42: 327-360.
276. Huestis, M. A. (2005). Pharmacokinetics and metabolism of the plant cannabinoids, delta9-tetrahydrocannabinol, cannabidiol and cannabinol. *Handb.Exp.Pharmacol.* 657-690.
277. Carter, G. T., Weydt, P., Kyashna-Tocha, M., and Abrams, D. I. (2004). Medicinal cannabis: rational guidelines for dosing. *IDrugs.* 7: 464-470.
278. Ohlsson, A., Lindgren, J. E., Wahlen, A., Agurell, S. and others. (1980). Plasma delta-9 tetrahydrocannabinol concentrations and clinical effects after oral and intravenous administration and smoking. *Clin.Pharmacol.Ther.* 28: 409-416.
279. Cooper, Z. D. and Haney, M. (2009). Comparison of subjective, pharmacokinetic, and physiological effects of marijuana smoked as joints and blunts. *Drug Alcohol Depend.* 103: 107-113.
280. Schwoppe, D. M., Bosker, W. M., Ramaekers, J. G., Gorelick, D. A. and others. (2012). Psychomotor performance, subjective and physiological effects and whole blood Delta(9)-tetrahydrocannabinol concentrations in heavy, chronic cannabis smokers following acute smoked cannabis. *J.Anal.Toxicol.* 36: 405-412.
281. Gieringer, D. H. (2001). Cannabis "Vaporization". *Journal of Cannabis Therapeutics.* 1: 153-170.
282. Gieringer, D., St Laurent, J., and Goodrich, S. (2004). Cannabis vaporizer combines efficient delivery of THC with effective suppression of pyrolytic compounds. *Journal of Cannabis Therapeutics.* 4: 7-27.
283. Hazekamp, A., Ruhaak, R., Zuurman, L., van, Gerven J. and others. (2006). Evaluation of a vaporizing device (Volcano) for the pulmonary administration of tetrahydrocannabinol. *J.Pharm.Sci.* 95: 1308-1317.
284. Pomahacova, B., Van der Kooy, F., and Verpoorte, R. (2009). Cannabis smoke condensate III: the cannabinoid content of vaporised Cannabis sativa. *Inhal.Toxicol.* 21: 1108-1112.
285. Walsh, D., Nelson, K. A., and Mahmoud, F. A. (2003). Established and potential therapeutic applications of cannabinoids in oncology. *Support.Care Cancer.* 11: 137-143.
286. Cone, E. J., Johnson, R. E., Paul, B. D., Mell, L. D. and others. (1988). Marijuana-laced brownies: behavioral effects, physiologic effects, and urinalysis in humans following ingestion. *J.Anal.Toxicol.* 12: 169-175.
287. Iversen, L. L. *The pharmacology of THC, the psychoactive ingredient in cannabis. The science of marijuana.* New York, New York: Oxford University Press, 2000.

288. Schwilke, E. W., Schwöpe, D. M., Karschner, E. L., Lowe, R. H. and others. (2009). Delta9-tetrahydrocannabinol (THC), 11-hydroxy-THC, and 11-nor-9-carboxy-THC plasma pharmacokinetics during and after continuous high-dose oral THC. *Clin.Chem.* 55: 2180-2189.
289. Office of Medicinal Cannabis, The Netherlands Ministry of Health Welfare and Sports. Medicinal Cannabis, Information for Health Care Professionals. 2008.
290. GW Pharmaceuticals. Sativex Product Monograph. 2010.
291. Wade, D. T., Makela, P., Robson, P., House, H. and others. (2004). Do cannabis-based medicinal extracts have general or specific effects on symptoms in multiple sclerosis? A double-blind, randomized, placebo-controlled study on 160 patients. *Mult.Scler.* 10: 434-441.
292. Nurmikko, T. J., Serpell, M. G., Hoggart, B., Toomey, P. J. and others. (2007). Sativex successfully treats neuropathic pain characterised by allodynia: a randomised, double-blind, placebo-controlled clinical trial. *Pain.* 133: 210-220.
293. Brenneisen, R., Egli, A., Elsohly, M. A., Henn, V. and others. (1996). The effect of orally and rectally administered delta 9-tetrahydrocannabinol on spasticity: a pilot study with 2 patients. *Int.J.Clin.Pharmacol.Ther.* 34: 446-452.
294. Mattes, R. D., Shaw, L. M., Edling-Owens, J., Engelman, K. and others. (1993). Bypassing the first-pass effect for the therapeutic use of cannabinoids. *Pharmacol.Biochem.Behav.* 44: 745-747.
295. Perlin, E., Smith, C. G., Nichols, A. I., Almirez, R. and others. (1985). Disposition and bioavailability of various formulations of tetrahydrocannabinol in the rhesus monkey. *J.Pharm.Sci.* 74: 171-174.
296. Elsohly, M. A., Little, T. L., Jr., Hikal, A., Harland, E. and others. (1991). Rectal bioavailability of delta-9-tetrahydrocannabinol from various esters. *Pharmacol.Biochem.Behav.* 40: 497-502.
297. Elsohly, M. A., Stanford, D. F., Harland, E. C., Hikal, A. H. and others. (1991). Rectal bioavailability of delta-9-tetrahydrocannabinol from the hemisuccinate ester in monkeys. *J.Pharm.Sci.* 80: 942-945.
298. Valiveti, S., Hammell, D. C., Earles, D. C., and Stinchcomb, A. L. (2004). Transdermal delivery of the synthetic cannabinoid WIN 55,212-2: in vitro/in vivo correlation. *Pharm.Res.* 21: 1137-1145.
299. Valiveti, S., Kiptoo, P. K., Hammell, D. C., and Stinchcomb, A. L. (2004). Transdermal permeation of WIN 55,212-2 and CP 55,940 in human skin in vitro. *Int.J.Pharm.* 278: 173-180.
300. Stinchcomb, A. L., Valiveti, S., Hammell, D. C., and Ramsey, D. R. (2004). Human skin permeation of Delta8-tetrahydrocannabinol, cannabidiol and cannabinol. *J.Pharm.Pharmacol.* 56: 291-297.
301. Harvey, D. J. Absorption, distribution and biotransformation of the cannabinoids. Marijuana and medicine. Nahas, C. G., Sutin, K. M., Harvey, D. J., and Agurell, S. Totowa, New Jersey: Humana Press, 1999.
302. Widman, M., Agurell, S., Ehrnebo, M., and Jones, G. (1974). Binding of (+)- and (minus)-delta-1-tetrahydrocannabinols and (minus)-7-hydroxy-delta-1-tetrahydrocannabinol to blood cells and plasma proteins in man. *J.Pharm.Pharmacol.* 26: 914-916.
303. Garrett, E. R. and Hunt, C. A. (1977). Pharmacokinetics of delta9-tetrahydrocannabinol in dogs. *J.Pharm.Sci.* 66: 395-407.
304. Wahlqvist, M., Nilsson, I. M., Sandberg, F., and Agurell, S. (1970). Binding of delta-1-tetrahydrocannabinol to human plasma proteins. *Biochem.Pharmacol.* 19: 2579-2584.

305. Widman, M., Nilsson, I. M., Agurell, S., Borg, H. and others. (1973). Plasma protein binding of 7-hydroxy-1-tetrahydrocannabinol: an active 1-tetrahydrocannabinol metabolite. *J.Pharm.Pharmacol.* 25: 453-457.
306. Truitt, E. B., Jr. (1971). Biological disposition of tetrahydrocannabinols. *Pharmacol.Rev.* 23: 273-278.
307. Nahas, G. G. (2001). The pharmacokinetics of THC in fat and brain: resulting functional responses to marijuana smoking. *Hum.Psychopharmacol.* 16: 247-255.
308. Schou, J., Prockop, L. D., Dahlstrom, G., and Rohde, C. (1977). Penetration of delta-9-tetrahydrocannabinol and 11-OH-delta-9-tetrahydrocannabinol through the blood-brain barrier. *Acta Pharmacol.Toxicol.(Copenh).* 41: 33-38.
309. Mura, P., Kintz, P., Dumestre, V., Raul, S. and others. (2005). THC can be detected in brain while absent in blood. *J.Anal.Toxicol.* 29: 842-843.
310. Gunasekaran, N., Long, L. E., Dawson, B. L., Hansen, G. H. and others. (2009). Reintoxication: the release of fat-stored delta(9)-tetrahydrocannabinol (THC) into blood is enhanced by food deprivation or ACTH exposure. *Br.J.Pharmacol.* 158: 1330-1337.
311. Lemberger, L. (1973). Tetrahydrocannabinol metabolism in man. *Drug Metab Dispos.* 1: 461-468.
312. Wall, M. E., Sadler, B. M., Brine, D., Taylor, H. and others. (1983). Metabolism, disposition, and kinetics of delta-9-tetrahydrocannabinol in men and women. *Clin.Pharmacol.Ther.* 34: 352-363.
313. Christensen, H. D., Freudenthal, R. I., Gidley, J. T., Rosenfeld, R. and others. (1971). Activity of delta8- and delta9-tetrahydrocannabinol and related compounds in the mouse. *Science.* 172: 165-167.
314. Perez-Reyes, M., Timmons, M. C., Lipton, M. A., Davis, K. H. and others. (1972). Intravenous injection in man of 9-tetrahydrocannabinol and 11-OH-9-tetrahydrocannabinol. *Science.* 177: 633-635.
315. Huestis, M. A., Mitchell, J. M., and Cone, E. J. (1996). Urinary excretion profiles of 11-nor-9-carboxy-delta 9-tetrahydrocannabinol in humans after single smoked doses of marijuana. *J.Anal.Toxicol.* 20: 441-452.
316. Hawks, R. L. (1982). The constituents of cannabis and the disposition and metabolism of cannabinoids. *NIDA Res.Monogr.* 42: 125-137.
317. Martin, B. R. and Cone, E. J. Chemistry and pharmacology of cannabis. *The Health Effects of Cannabis.* Kalant, H., Corrigan, W., Hall, W., and Smart, R. Toronto, Canada: Centre of Addiction and Mental Health, 1999.
318. Sachse-Seeboth, C., Pfeil, J., Sehr, D., Meineke, I. and others. (2009). Interindividual variation in the pharmacokinetics of Delta9-tetrahydrocannabinol as related to genetic polymorphisms in CYP2C9. *Clin.Pharmacol.Ther.* 85: 273-276.
319. Oates, J. A. The science of drug therapy. *Goodman and Gilman's the pharmacological basis of therapeutics.* Brunton, L. L., Lazo, J. S., and Parker, K. L. New York: McGraw-Hill, 2006.
320. Graham, M. J. and Lake, B. G. (2008). Induction of drug metabolism: species differences and toxicological relevance. *Toxicology.* 254: 184-191.
321. Bornheim, L. M., Everhart, E. T., Li, J., and Correia, M. A. (1993). Characterization of cannabidiol-mediated cytochrome P450 inactivation. *Biochem.Pharmacol.* 45: 1323-1331.
322. Kosel, B. W., Aweeka, F. T., Benowitz, N. L., Shade, S. B. and others. (2002). The effects of cannabinoids on the pharmacokinetics of indinavir and nelfinavir. *AIDS.* 16: 543-550.

323. Jusko, W. J., Schentag, J. J., Clark, J. H., Gardner, M. and others. (1978). Enhanced biotransformation of theophylline in marijuana and tobacco smokers. *Clin.Pharmacol.Ther.* 24: 405-410.
324. Zullino, D. F., Delessert, D., Eap, C. B., Preisig, M. and others. (2002). Tobacco and cannabis smoking cessation can lead to intoxication with clozapine or olanzapine. *Int.Clin.Psychopharmacol.* 17: 141-143.
325. Huestis, M. A., Henningfield, J. E., and Cone, E. J. (1992). Blood cannabinoids. I. Absorption of THC and formation of 11-OH-THC and THCCOOH during and after smoking marijuana. *J.Anal.Toxicol.* 16: 276-282.
326. Agurell, S. and Leander, K. (1971). Stability, transfer and absorption of cannabinoid constituents of cannabis (hashish) during smoking. *Acta Pharm.Suec.* 8: 391-402.
327. Wall, M. E. and Perez-Reyes, M. (1981). The metabolism of delta 9-tetrahydrocannabinol and related cannabinoids in man. *J.Clin.Pharmacol.* 21: 178S-189S.
328. Huestis, M. A., Sampson, A. H., Holicky, B. J., Henningfield, J. E. and others. (1992). Characterization of the absorption phase of marijuana smoking. *Clin.Pharmacol.Ther.* 52: 31-41.
329. Johansson, E., Agurell, S., Hollister, L. E., and Haldin, M. M. (1988). Prolonged apparent half-life of delta 1-tetrahydrocannabinol in plasma of chronic marijuana users. *J.Pharm.Pharmacol.* 40: 374-375.
330. Cone, E. J. and Huestis, M. A. (1993). Relating blood concentrations of tetrahydrocannabinol and metabolites to pharmacologic effects and time of marijuana usage. *Ther.Drug Monit.* 15: 527-532.
331. Toennes, S. W., Ramaekers, J. G., Theunissen, E. L., Moeller, M. R. and others. (2008). Comparison of cannabinoid pharmacokinetic properties in occasional and heavy users smoking a marijuana or placebo joint. *J.Anal.Toxicol.* 32: 470-477.
332. Valeant Canada. Cesamet Product Monograph. 2009.
333. Hollister, L. E., Gillespie, H. K., Ohlsson, A., Lindgren, J. E. and others. (1981). Do plasma concentrations of delta 9-tetrahydrocannabinol reflect the degree of intoxication? *J.Clin.Pharmacol.* 21: 171S-177S.
334. Strougo, A., Zuurman, L., Roy, C., Pinquier, J. L. and others. (2008). Modelling of the concentration--effect relationship of THC on central nervous system parameters and heart rate -- insight into its mechanisms of action and a tool for clinical research and development of cannabinoids. *J.Psychopharmacol.* 22: 717-726.
335. Lynch, M. E. and Watson, C. P. (2006). The pharmacotherapy of chronic pain: a review. *Pain Res.Manag.* 11: 11-38.
336. Wu, D. F., Yang, L. Q., Goschke, A., Stumm, R. and others. (2008). Role of receptor internalization in the agonist-induced desensitization of cannabinoid type 1 receptors. *J.Neurochem.* 104: 1132-1143.
337. Maldonado, R. (2002). Study of cannabinoid dependence in animals. *Pharmacol.Ther.* 95: 153-164.
338. Pertwee, R. G. (2009). Emerging strategies for exploiting cannabinoid receptor agonists as medicines. *Br.J.Pharmacol.* 156: 397-411.
339. Hirvonen, J., Goodwin, R. S., Li, C. T., Terry, G. E. and others. (2012). Reversible and regionally selective downregulation of brain cannabinoid CB1 receptors in chronic daily cannabis smokers. *Mol.Psychiatry.* 17: 642-649.
340. Haney, M., Ward, A. S., Comer, S. D., Foltin, R. W. and others. (1999). Abstinence symptoms following smoked marijuana in humans. *Psychopharmacology (Berl).* 141: 395-404.

341. American Psychiatric Association. Substance-related disorders. Diagnostic and statistical manual of mental disorders text revision (DSM-IV-TR). American Psychiatric Association. Washington, D.C.: American Psychiatric Association, 2000.
342. Budney, A. J., Hughes, J. R., Moore, B. A., and Vandrey, R. (2004). Review of the validity and significance of cannabis withdrawal syndrome. *Am.J.Psychiatry*. 161: 1967-1977.
343. Hazekamp, A., Bastola, K., Rashidi, H., Bender, J. and others. (2007). Cannabis tea revisited: a systematic evaluation of the cannabinoid composition of cannabis tea. *J.Ethnopharmacol*. 113: 85-90.
344. Pertwee, R. G. (1974). Tolerance to the effect of delta1-tetrahydrocannabinol on corticosterone levels in mouse plasma produced by repeated administration of cannabis extract or delta1-tetrahydrocannabinol. *Br.J.Pharmacol*. 51: 391-397.
345. Lozano, I. (2001). The therapeutic uses of Cannabis sativa (L.) in Arabic medicine. *Journal of Cannabis Therapeutics*. 1: 63-70.
346. Russo, E. History of cannabis as a medicine. *The Medicinal Uses of Cannabis and Cannabinoids*. Guy, G. W., Whittle, B. A., and Robson, P. J. London: Pharmaceutical Press, 2004.
347. Russo, E. B. (2007). History of cannabis and its preparations in saga, science, and sobriquet. *Chem.Biodivers*. 4: 1614-1648.
348. Fraser, G. A. (2009). The use of a synthetic cannabinoid in the management of treatment-resistant nightmares in posttraumatic stress disorder (PTSD). *CNS.Neurosci.Ther*. 15: 84-88.
349. Portenoy, R. K., Ganae-Motan, E. D., Allende, S., Yanagihara, R. and others. (2012). Nabiximols for opioid-treated cancer patients with poorly-controlled chronic pain: a randomized, placebo-controlled, graded-dose trial. *J.Pain*. 13: 438-449.
350. Ware, M. A., Adams, H., and Guy, G. W. (2005). The medicinal use of cannabis in the UK: results of a nationwide survey. *Int.J.Clin.Pract*. 59: 291-295.
351. World Health Organization (WHO). Cannabis: a health perspective and research agenda. 1997
352. Cami, J., Guerra, D., Ugena, B., Segura, J. and others. (1991). Effect of subject expectancy on the THC intoxication and disposition from smoked hashish cigarettes. *Pharmacol.Biochem.Behav*. 40: 115-119.
353. Skrabek, R. Q., Galimova, L., Ethans, K., and Perry, D. (2008). Nabilone for the treatment of pain in fibromyalgia. *J.Pain*. 9: 164-173.
354. Ware, M. A., Fitzcharles, M. A., Joseph, L., and Shir, Y. (2010). The effects of nabilone on sleep in fibromyalgia: results of a randomized controlled trial. *Anesth.Analg*. 110: 604-610.
355. Kalliomaki, J., Philipp, A., Baxendale, J., Annas, P. and others. (2012). Lack of effect of central nervous system-active doses of nabilone on capsaicin-induced pain and hyperalgesia. *Clin.Exp.Pharmacol.Physiol*. 39: 336-342.
356. World Health Organization (WHO). WHO Definition of Palliative Care. 2012.
357. Green, A. J. and De-Vries, K. (2010). Cannabis use in palliative care - an examination of the evidence and the implications for nurses. *J.Clin.Nurs*. 19: 2454-2462.
358. Gardiner, C. and Ingleton, C. (2010). Commentary on Green AJ & De-Vries K (2010) Cannabis use in palliative care—an examination of the evidence and the implications for nurses. *Journal of Clinical Nursing* 19, 2454-2462. *J.Clin.Nurs*. 19: 3253-3255.

359. Glare, P., Miller, J., Nikolova, T., and Tickoo, R. (2011). Treating nausea and vomiting in palliative care: a review. *Clin.Interv.Aging.* 6: 243-259.
360. Fine, P. G. (2012). Treatment guidelines for the pharmacological management of pain in older persons. *Pain Med.* 13 Suppl 2: S57-S66.
361. Svendsen, K. B., Jensen, T. S., and Bach, F. W. (2004). Does the cannabinoid dronabinol reduce central pain in multiple sclerosis? Randomised double blind placebo controlled crossover trial. *BMJ.* 329: 253-260.
362. Brisbois, T. D., de Kock, I. H., Watanabe, S. M., Mirhosseini, M. and others. (2011). Delta-9-tetrahydrocannabinol may palliate altered chemosensory perception in cancer patients: results of a randomized, double-blind, placebo-controlled pilot trial. *Ann.Oncol.* 22: 2086-2093.
363. Strasser, F., Luftner, D., Possinger, K., Ernst, G. and others. (2006). Comparison of orally administered cannabis extract and delta-9-tetrahydrocannabinol in treating patients with cancer-related anorexia-cachexia syndrome: a multicenter, phase III, randomized, double-blind, placebo-controlled clinical trial from the Cannabis-In-Cachexia-Study-Group. *J.Clin.Oncol.* 24: 3394-3400.
364. Toth, C., Mawani, S., Brady, S., Chan, C. and others. (2012). An enriched-enrolment, randomized withdrawal, flexible-dose, double-blind, placebo-controlled, parallel assignment efficacy study of nabilone as adjuvant in the treatment of diabetic peripheral neuropathic pain. *Pain.* 153: 2073-2082.
365. Selvarajah, D., Gandhi, R., Emery, C. J., and Tesfaye, S. (2010). Randomized placebo-controlled double-blind clinical trial of cannabis-based medicinal product (Sativex[®]) in painful diabetic neuropathy: depression is a major confounding factor. *Diabetes Care.* 33: 128-130.
366. Novotna, A., Mares, J., Ratcliffe, S., Novakova, I. and others. (2011). A randomized, double-blind, placebo-controlled, parallel-group, enriched-design study of nabiximols[®] (Sativex[®]), as add-on therapy, in subjects with refractory spasticity caused by multiple sclerosis. *Eur.J.Neurol.* 18: 1122-1131.
367. Navari, R. M. (2009). Pharmacological management of chemotherapy-induced nausea and vomiting: focus on recent developments. *Drugs.* 69: 515-533.
368. Hornby, P. J. (2001). Central neurocircuitry associated with emesis. *Am.J.Med.* 111 Suppl 8A: 106S-112S.
369. Van Sickle, M. D., Duncan, M., Kingsley, P. J., Mouihate, A. and others. (2005). Identification and functional characterization of brainstem cannabinoid CB2 receptors. *Science.* 310: 329-332.
370. Darmani, N. A. (2001). The cannabinoid CB1 receptor antagonist SR 141716A reverses the antiemetic and motor depressant actions of WIN 55, 212-2. *Eur.J.Pharmacol.* 430: 49-58.
371. Barann, M., Molderings, G., Bruss, M., Bonisch, H. and others. (2002). Direct inhibition by cannabinoids of human 5-HT3A receptors: probable involvement of an allosteric modulatory site. *Br.J.Pharmacol.* 137: 589-596.
372. Rock, E. M., Bolognini, D., Limebeer, C. L., Cascio, M. G. and others. (2012). Cannabidiol, a non-psychotropic component of cannabis, attenuates vomiting and nausea-like behaviour via indirect agonism of 5-HT(1A) somatodendritic autoreceptors in the dorsal raphe nucleus. *Br.J.Pharmacol.* 165: 2620-2634.
373. Rock, E. M., Goodwin, J. M., Limebeer, C. L., Breuer, A. and others. (2011). Interaction between non-psychotropic cannabinoids in marijuana: effect of cannabigerol (CBG) on the anti-nausea or anti-emetic effects of cannabidiol (CBD) in rats and shrews. *Psychopharmacology (Berl).* 215: 505-512.
374. Machado Rocha, F. C., Stefano, S. C., De Cassia, Haiek R., Rosa Oliveira, L. M. and others. (2008). Therapeutic use of Cannabis sativa on chemotherapy-induced nausea and vomiting among cancer patients: systematic review and meta-analysis. *Eur.J.Cancer Care (Engl.).* 17: 431-443.

375. Meiri, E., Jhangiani, H., Vredenburg, J. J., Barbato, L. M. and others. (2007). Efficacy of dronabinol alone and in combination with ondansetron versus ondansetron alone for delayed chemotherapy-induced nausea and vomiting. *Curr.Med.Res.Opin.* 23: 533-543.
376. Kwiatkowska, M., Parker, L. A., Burton, P., and Mechoulam, R. (2004). A comparative analysis of the potential of cannabinoids and ondansetron to suppress cisplatin-induced emesis in the *Suncus murinus* (house musk shrew). *Psychopharmacology (Berl)*. 174: 254-259.
377. Wang, Y., Ray, A. P., McClanahan, B. A., and Darmani, N. A. (2009). The antiemetic interaction of Delta9-tetrahydrocannabinol when combined with tropisetron or dexamethasone in the least shrew. *Pharmacol.Biochem.Behav.* 91: 367-373.
378. Institute of Medicine. The medical value of marijuana and related substances. *Marijuana and medicine: Assessing the science base.* Joy, J. E., Watson, S. J., and Benson, J. A. Washington, DC: National Academy Press, 1999.
379. Health Department of New South Wales, Australia. Working Party on the Use of Cannabis for Medical Purposes. (2000). 2.
380. Herrstedt, J. and Dombernowsky, P. (2007). Anti-emetic therapy in cancer chemotherapy: current status. *Basic Clin.Pharmacol.Toxicol.* 101: 143-150.
381. Canadian Pharmacists Association. Compendium of Pharmaceuticals and Specialties. Ottawa: Canadian Pharmacists Association, 2009.
382. National Comprehensive Cancer Network. NCCN Clinical Practice Guidelines in Oncology. 2010.
383. Clary, P. L. and Lawson, P. (2009). Pharmacologic pearls for end-of-life care. *Am.Fam.Physician.* 79: 1059-1065.
384. Minister of Justice, Government of Canada. Marihuana Medical Access Regulations. 2011.
385. Smit, E. and Crespo, C. J. (2001). Dietary intake and nutritional status of US adult marijuana users: results from the Third National Health and Nutrition Examination Survey. *Public Health Nutr.* 4: 781-786.
386. Di Marzo, V. and Matias, I. (2005). Endocannabinoid control of food intake and energy balance. *Nat.Neurosci.* 8: 585-589.
387. Matias, I., Bisogno, T., and Di, Marzo, V. (2006). Endogenous cannabinoids in the brain and peripheral tissues: regulation of their levels and control of food intake. *Int.J.Obes.(Lond)*. 30 Suppl 1: S7-S12.
388. Tibirica, E. (2010). The multiple functions of the endocannabinoid system: a focus on the regulation of food intake. *Diabetol.Metab Syndr.* 2: 5-10.
389. Farrimond, J. A., Mercier, M. S., Whalley, B. J., and Williams, C. M. (2011). Cannabis sativa and the endogenous cannabinoid system: therapeutic potential for appetite regulation. *Phytother.Res.* 25: 170-188.
390. Abrams, D. I., Hilton, J. F., Leiser, R. J., Shade, S. B. and others. (2003). Short-term effects of cannabinoids in patients with HIV-1 infection: a randomized, placebo-controlled clinical trial. *Ann.Intern.Med.* 139: 258-266.
391. Ravinet-Trillou, C., Delgorge, C., Menet, C., Amone, M. and others. (2004). CB1 cannabinoid receptor knockout in mice leads to leanness, resistance to diet-induced obesity and enhanced leptin sensitivity. *Int.J.Obes.Relat Metab Disord.* 28: 640-648.
392. Timpone, J. G., Wright, D. J., Li, N., Egorin, M. J. and others. (1997). The safety and pharmacokinetics of single-agent and combination therapy with megestrol acetate and dronabinol for the treatment of HIV wasting

syndrome. The DATRI 004 Study Group. Division of AIDS Treatment Research Initiative. *AIDS Res.Hum.Retroviruses*. 13: 305-315.

393. Beal, J. E., Olson, R., Laubenstein, L., Morales, J. O. and others. (1995). Dronabinol as a treatment for anorexia associated with weight loss in patients with AIDS. *J.Pain Symptom.Manage*. 10: 89-97.
394. Beal, J. E., Olson, R., Lefkowitz, L., Laubenstein, L. and others. (1997). Long-term efficacy and safety of dronabinol for acquired immunodeficiency syndrome-associated anorexia. *J.Pain Symptom.Manage*. 14: 7-14.
395. Tchekmedyan, N. S., Zahyna, D., Halpert, C., and Heber, D. (1992). Clinical aspects of nutrition in advanced cancer. *Oncology*. 49 Suppl 2: 3-7.
396. Walsh, D., Donnelly, S., and Rybicki, L. (2000). The symptoms of advanced cancer: relationship to age, gender, and performance status in 1,000 patients. *Support.Care Cancer*. 8: 175-179.
397. Ekert, H., Waters, K. D., Jurk, I. H., Mobilia, J. and others. (1979). Amelioration of cancer chemotherapy-induced nausea and vomiting by delta-9-tetrahydrocannabinol. *Med.J.Aust*. 2: 657-659.
398. Sallan, S. E., Cronin, C., Zelen, M., and Zinberg, N. E. (1980). Antiemetics in patients receiving chemotherapy for cancer: a randomized comparison of delta-9-tetrahydrocannabinol and prochlorperazine. *N.Engl.J.Med*. 302: 135-138.
399. Plasse, T. F., Gorter, R. W., Krasnow, S. H., Lane, M. and others. (1991). Recent clinical experience with dronabinol. *Pharmacol.Biochem.Behav*. 40: 695-700.
400. Nelson, K., Walsh, D., Deeter, P., and Sheehan, F. (1994). A phase II study of delta-9-tetrahydrocannabinol for appetite stimulation in cancer-associated anorexia. *J.Palliat.Care*. 10: 14-18.
401. Regelson, W., Butler, J. R., and Schulz, J. Delta-9-tetrahydrocannabinol as an effective antidepressant and appetite-stimulating agent in advanced cancer patients. *The Pharmacology of marihuana: A monograph of the National Institute on Drug Abuse*. Braude, M. and Szara, S. New York: Raven Press, 1976.
402. Jatoi, A., Windschitl, H. E., Loprinzi, C. L., Sloan, J. A. and others. (2002). Dronabinol versus megestrol acetate versus combination therapy for cancer-associated anorexia: a North Central Cancer Treatment Group study. *J.Clin.Oncol*. 20: 567-573.
403. Mantovani, G., Maccio, A., Madeddu, C., Serpe, R. and others. (2010). Randomized phase III clinical trial of five different arms of treatment in 332 patients with cancer cachexia. *Oncologist*. 15: 200-211.
404. Monteleone, P., Matias, I., Martiadis, V., De, Petrocellis L. and others. (2005). Blood levels of the endocannabinoid anandamide are increased in anorexia nervosa and in binge-eating disorder, but not in bulimia nervosa. *Neuropsychopharmacology*. 30: 1216-1221.
405. Siegfried, Z., Kanyas, K., Latzer, Y., Karni, O. and others. (2004). Association study of cannabinoid receptor gene (CNRI) alleles and anorexia nervosa: differences between restricting and binge/purging subtypes. *Am.J.Med.Genet.B Neuropsychiatr.Genet*. 125B: 126-130.
406. Muller, T. D., Reichwald, K., Bronner, G., Kirschner, J. and others. (2008). Lack of association of genetic variants in genes of the endocannabinoid system with anorexia nervosa. *Child Adolesc.Psychiatry Ment.Health*. 2: 33-39.
407. Lewis, D. Y. and Brett, R. R. (2010). Activity-based anorexia in C57/BL6 mice: effects of the phytocannabinoid, Delta9-tetrahydrocannabinol (THC) and the anandamide analogue, OMDM-2. *Eur.Neuropsychopharmacol*. 20: 622-631.

408. Verty, A. N., Evetts, M. J., Crouch, G. J., McGregor, I. S. and others. (2011). The cannabinoid receptor agonist THC attenuates weight loss in a rodent model of activity-based anorexia. *Neuropsychopharmacology*. 36: 1349-1358.
409. Gross, H., Ebert, M. H., Faden, V. B., Goldberg, S. C. and others. (1983). A double-blind trial of delta 9-tetrahydrocannabinol in primary anorexia nervosa. *J.Clin.Psychopharmacol*. 3: 165-171.
410. Volicer, L., Stelly, M., Morris, J., McLaughlin, J. and others. (1997). Effects of dronabinol on anorexia and disturbed behavior in patients with Alzheimer's disease. *Int.J.Geriatr.Psychiatry*. 12: 913-919.
411. American Medical Association, Council of Scientific Affairs. Medical Marijuana. Internet. American Medical Association, Council of Scientific Affairs. 1997.
412. Aggarwal, S. K., Carter, G. T., Sullivan, M. D., ZumBrunnen, C. and others. (2009). Characteristics of patients with chronic pain accessing treatment with medical cannabis in Washington State. *J.Opioid.Manag*. 5: 257-286.
413. Heutink, M., Post, M. W., Wollaars, M. M., and van Asbeck, F. W. (2011). Chronic spinal cord injury pain: pharmacological and non-pharmacological treatments and treatment effectiveness. *Disabil.Rehabil*. 33: 433-440.
414. Baker, D., Pryce, G., Croxford, J. L., Brown, P. and others. (2000). Cannabinoids control spasticity and tremor in a multiple sclerosis model. *Nature*. 404: 84-87.
415. Centonze, D., Rossi, S., Finazzi-Agro, A., Bernardi, G. and others. (2007). The (endo)cannabinoid system in multiple sclerosis and amyotrophic lateral sclerosis. *Int.Rev.Neurobiol*. 82: 171-186.
416. Di Filippo, M., Pini, L. A., Pelliccioli, G. P., Calabresi, P. and others. (2008). Abnormalities in the cerebrospinal fluid levels of endocannabinoids in multiple sclerosis. *J.Neurol.Neurosurg.Psychiatry*. 79: 1224-1229.
417. Jean-Gilles, L., Feng, S., Tench, C. R., Chapman, V. and others. (2009). Plasma endocannabinoid levels in multiple sclerosis. *J.Neurol.Sci*. 287: 212-215.
418. Pertwee, R. G. (2007). Cannabinoids and multiple sclerosis. *Mol.Neurobiol*. 36: 45-59.
419. Lyman, W. D., Sonett, J. R., Brosnan, C. F., Elkin, R. and others. (1989). Delta 9-tetrahydrocannabinol: a novel treatment for experimental autoimmune encephalomyelitis. *J.Neuroimmunol*. 23: 73-81.
420. Maresz, K., Pryce, G., Ponomarev, E. D., Marsicano, G. and others. (2007). Direct suppression of CNS autoimmune inflammation via the cannabinoid receptor CB1 on neurons and CB2 on autoreactive T cells. *Nat.Med*. 13: 492-497.
421. Pryce, G. and Baker, D. (2007). Control of spasticity in a multiple sclerosis model is mediated by CB1, not CB2, cannabinoid receptors. *Br.J.Pharmacol*. 150: 519-525.
422. Croxford, J. L., Pryce, G., Jackson, S. J., Ledent, C. and others. (2008). Cannabinoid-mediated neuroprotection, not immunosuppression, may be more relevant to multiple sclerosis. *J.Neuroimmunol*. 193: 120-129.
423. Baker, D., Jackson, S. J., and Pryce, G. (2007). Cannabinoid control of neuroinflammation related to multiple sclerosis. *Br.J.Pharmacol*. 152: 649-654.
424. Sanchez, A. J. and Garcia-Merino, A. (2011). Neuroprotective agents: Cannabinoids. *Clin.Immunol*. 142: 57-67.
425. Consroe, P. and Sandyk, R. Therapeutic potential of cannabinoids in neurological disorders. *Marijuana/Cannabinoids as therapeutic agents*. Mechoulam, R. Boca Raton, FL: CRC Press, 1986.

426. Chong, M. S., Wolff, K., Wise, K., Tanton, C. and others. (2006). Cannabis use in patients with multiple sclerosis. *Mult.Scler.* 12: 646-651.
427. Zajicek, J. P., Sanders, H. P., Wright, D. E., Vickery, P. J. and others. (2005). Cannabinoids in multiple sclerosis (CAMS) study: safety and efficacy data for 12 months follow up. *J.Neurol.Neurosurg.Psychiatry.* 76: 1664-1669.
428. Vaney, C., Heinzl-Gutenbrunner, M., Jobin, P., Tschopp, F. and others. (2004). Efficacy, safety and tolerability of an orally administered cannabis extract in the treatment of spasticity in patients with multiple sclerosis: a randomized, double-blind, placebo-controlled, crossover study. *Mult.Scler.* 10: 417-424.
429. Wade, D. T., Makela, P. M., House, H., Bateman, C. and others. (2006). Long-term use of a cannabis-based medicine in the treatment of spasticity and other symptoms in multiple sclerosis. *Mult.Scler.* 12: 639-645.
430. Collin, C., Davies, P., Mutiboko, I. K., and Ratcliffe, S. (2007). Randomized controlled trial of cannabis-based medicine in spasticity caused by multiple sclerosis. *Eur.J.Neurol.* 14: 290-296.
431. Hobart, J. C., Riazi, A., Thompson, A. J., Styles, I. M. and others. (2006). Getting the measure of spasticity in multiple sclerosis: the Multiple Sclerosis Spasticity Scale (MSSS-88). *Brain.* 129: 224-234.
432. Zajicek, J. P., Hobart, J. C., Slade, A., Barnes, D. and others. (2012). Multiple Sclerosis and Extract of Cannabis: results of the MUSEC trial. *J.Neurol.Neurosurg.Psychiatry.* 83: 1125-1132.
433. Killestein, J., Hoogervorst, E. L., Reif, M., Kalkers, N. F. and others. (2002). Safety, tolerability, and efficacy of orally administered cannabinoids in MS. *Neurology.* 58: 1404-1407.
434. Aragona, M., Onesti, E., Tomassini, V., Conte, A. and others. (2009). Psychopathological and cognitive effects of therapeutic cannabinoids in multiple sclerosis: a double-blind, placebo controlled, crossover study. *Clin.Neuropharmacol.* 32: 41-47.
435. Freeman, R. M., Adekanmi, O., Waterfield, M. R., Waterfield, A. E. and others. (2006). The effect of cannabis on urge incontinence in patients with multiple sclerosis: a multicentre, randomised placebo-controlled trial (CAMS-LUTS). *Int.Urogynecol.J.Pelvic.Floor.Dysfunct.* 17: 636-641.
436. Sliwa, J. A., Bell, H. K., Mason, K. D., Gore, R. M. and others. (1996). Upper urinary tract abnormalities in multiple sclerosis patients with urinary symptoms. *Arch.Phys.Med.Rehabil.* 77: 247-251.
437. Consroe, P., Musty, R., Rein, J., Tillery, W. and others. (1997). The perceived effects of smoked cannabis on patients with multiple sclerosis. *Eur.Neurol.* 38: 44-48.
438. Brady, C. M., DasGupta, R., Dalton, C., Wiseman, O. J. and others. (2004). An open-label pilot study of cannabis-based extracts for bladder dysfunction in advanced multiple sclerosis. *Mult.Scler.* 10: 425-433.
439. Rossi, S., Bernardi, G., and Centonze, D. (2010). The endocannabinoid system in the inflammatory and neurodegenerative processes of multiple sclerosis and of amyotrophic lateral sclerosis. *Exp.Neurol.* 224: 92-102.
440. Scotter, E. L., Abood, M. E., and Glass, M. (2010). The endocannabinoid system as a target for the treatment of neurodegenerative disease. *Br.J.Pharmacol.* 160: 480-498.
441. Amtmann, D., Weydt, P., Johnson, K. L., Jensen, M. P. and others. (2004). Survey of cannabis use in patients with amyotrophic lateral sclerosis. *Am.J.Hosp.Palliat.Care.* 21: 95-104.
442. Weber, M., Goldman, B., and Truniger, S. (2010). Tetrahydrocannabinol (THC) for cramps in amyotrophic lateral sclerosis: a randomised, double-blind crossover trial. *J.Neurol.Neurosurg.Psychiatry.* 81: 1135-1140.

443. Gelinas, D. F., Miller, R. G., and Abood, M. (2002). Pilot study of safety and tolerability of delta 9-THC (Marinol) treatment for ALS. *Amyotroph Lateral Scler Other Motor Neuron Disord.* 3: 23-24.
444. Garcia-Ovejero, D., Arevalo-Martin, A., Petrosino, S., Docagne, F. and others. (2009). The endocannabinoid system is modulated in response to spinal cord injury in rats. *Neurobiol.Dis.* 33: 57-71.
445. Hama, A. and Sagen, J. (2007). Antinociceptive effect of cannabinoid agonist WIN 55,212-2 in rats with a spinal cord injury. *Exp.Neurol.* 204: 454-457.
446. Hama, A. and Sagen, J. (2009). Sustained antinociceptive effect of cannabinoid receptor agonist WIN 55,212-2 over time in rat model of neuropathic spinal cord injury pain. *J.Rehabil.Res.Dev.* 46: 135-143.
447. Malec, J., Harvey, R. F., and Cayner, J. J. (1982). Cannabis effect on spasticity in spinal cord injury. *Arch.Phys.Med.Rehabil.* 63: 116-118.
448. Maurer, M., Henn, V., Dittrich, A., and Hofmann, A. (1990). Delta-9-tetrahydrocannabinol shows antispastic and analgesic effects in a single case double-blind trial. *Eur.Arch.Psychiatry Clin.Neurosci.* 240: 1-4.
449. Wade, D. T., Robson, P., House, H., Makela, P. and others. (2003). A preliminary controlled study to determine whether whole-plant cannabis extracts can improve intractable neurogenic symptoms. *Clin.Rehabil.* 17: 21-29.
450. Hagenbach, U., Luz, S., Ghafoor, N., Berger, J. M. and others. (2007). The treatment of spasticity with Delta9-tetrahydrocannabinol in persons with spinal cord injury. *Spinal Cord.* 45: 551-562.
451. Pooyania, S., Ethans, K., Szturm, T., Casey, A. and others. (2010). A randomized, double-blinded, crossover pilot study assessing the effect of nabilone on spasticity in persons with spinal cord injury. *Arch.Phys.Med.Rehabil.* 91: 703-707.
452. Falenski, K. W., Blair, R. E., Sim-Selley, L. J., Martin, B. R. and others. (2007). Status epilepticus causes a long-lasting redistribution of hippocampal cannabinoid type 1 receptor expression and function in the rat pilocarpine model of acquired epilepsy. *Neuroscience.* 146: 1232-1244.
453. Ludanyi, A., Eross, L., Czirjak, S., Vajda, J. and others. (2008). Downregulation of the CB1 cannabinoid receptor and related molecular elements of the endocannabinoid system in epileptic human hippocampus. *J.Neurosci.* 28: 2976-2990.
454. Falenski, K. W., Carter, D. S., Harrison, A. J., Martin, B. R. and others. (2009). Temporal characterization of changes in hippocampal cannabinoid CB(1) receptor expression following pilocarpine-induced status epilepticus. *Brain Res.* 1262: 64-72.
455. Romigi, A., Bari, M., Placidi, F., Marciani, M. G. and others. (2010). Cerebrospinal fluid levels of the endocannabinoid anandamide are reduced in patients with untreated newly diagnosed temporal lobe epilepsy. *Epilepsia.* 51: 768-772.
456. Wallace, M. J., Martin, B. R., and DeLorenzo, R. J. (2002). Evidence for a physiological role of endocannabinoids in the modulation of seizure threshold and severity. *Eur.J.Pharmacol.* 452: 295-301.
457. Mechoulam, R. and Lichtman, A. H. (2003). Neuroscience. Stout guards of the central nervous system. *Science.* 302: 65-67.
458. Wallace, M. J., Blair, R. E., Falenski, K. W., Martin, B. R. and others. (2003). The endogenous cannabinoid system regulates seizure frequency and duration in a model of temporal lobe epilepsy. *J.Pharmacol.Exp.Ther.* 307: 129-137.

459. Shafaroodi, H., Samini, M., Moezi, L., Homayoun, H. and others. (2004). The interaction of cannabinoids and opioids on pentylenetetrazole-induced seizure threshold in mice. *Neuropharmacology*. 47: 390-400.
460. Clement, A. B., Hawkins, E. G., Lichtman, A. H., and Cravatt, B. F. (2003). Increased seizure susceptibility and proconvulsant activity of anandamide in mice lacking fatty acid amide hydrolase. *J.Neurosci*. 23: 3916-3923.
461. Alger, B. E. (2002). Retrograde signaling in the regulation of synaptic transmission: focus on endocannabinoids. *Prog.Neurobiol*. 68: 247-286.
462. Smith, P. F. (2005). Cannabinoids as potential anti-epileptic drugs. *Curr.Opin.Investig.Drugs*. 6: 680-685.
463. Hoffman, A. F. and Lupica, C. R. (2000). Mechanisms of cannabinoid inhibition of GABA(A) synaptic transmission in the hippocampus. *J.Neurosci*. 20: 2470-2479.
464. Nakatsuka, T., Chen, H. X., Roper, S. N., and Gu, J. G. (2003). Cannabinoid receptor-1 activation suppresses inhibitory synaptic activity in human dentate gyrus. *Neuropharmacology*. 45: 116-121.
465. Gordon, E. and Devinsky, O. (2001). Alcohol and marijuana: effects on epilepsy and use by patients with epilepsy. *Epilepsia*. 42: 1266-1272.
466. Lutz, B. (2004). On-demand activation of the endocannabinoid system in the control of neuronal excitability and epileptiform seizures. *Biochem.Pharmacol*. 68: 1691-1698.
467. Gloss, D. and Vickrey, B. (2012). Cannabinoids for epilepsy. *Cochrane.Database.Syst.Rev*. 6: CD009270-
468. Hama, A. T. and Sagen, J. (2010). Cannabinoid receptor-mediated antinociception with acetaminophen drug combinations in rats with neuropathic spinal cord injury pain. *Neuropharmacology*. 58: 758-766.
469. Manzanares, J., Julian, M., and Carrascosa, A. (2006). Role of the cannabinoid system in pain control and therapeutic implications for the management of acute and chronic pain episodes. *Curr.Neuropharmacol*. 4: 239-257.
470. Christie, M. J. and Mallet, C. (2009). Endocannabinoids can open the pain gate. *Sci.Signal*. 2: pe57.-
471. Ostenfeld, T., Price, J., Albanese, M., Bullman, J. and others. (2011). A Randomized, Controlled Study to Investigate the Analgesic Efficacy of Single Doses of the Cannabinoid Receptor-2 Agonist GW842166, Ibuprofen or Placebo in Patients With Acute Pain Following Third Molar Tooth Extraction. *Clin.J.Pain*. 27: 668-676.
472. Karst, M., Wippermann, S., and Ahrens, J. (2010). Role of cannabinoids in the treatment of pain and (painful) spasticity. *Drugs*. 70: 2409-2438.
473. Mirchandani, A., Saleeb, M., and Sinatra, R. *Acute and Chronic Mechanisms of Pain. Essentials of Pain Management*. Vadivelu, N., Urman, R. D., and Hines, R. L. New York: Springer, 2011.
474. Seifert, F. and Maihofner, C. (2011). Functional and structural imaging of pain-induced neuroplasticity. *Curr.Opin.Anaesthesiol*. 24: 515-523.
475. Zubieta, J. K. and Stohler, C. S. (2009). Neurobiological mechanisms of placebo responses. *Ann.N.Y.Acad.Sci*. 1156: 198-210.
476. Finniss, D. G., Kaptchuk, T. J., Miller, F., and Benedetti, F. (2010). Biological, clinical, and ethical advances of placebo effects. *Lancet*. 375: 686-695.
477. Martin, B. R., Compton, D. R., Semus, S. F., Lin, S. and others. (1993). Pharmacological evaluation of iodo and nitro analogs of delta 8-THC and delta 9-THC. *Pharmacol.Biochem.Behav*. 46: 295-301.

478. Meng, I. D., Manning, B. H., Martin, W. J., and Fields, H. L. (1998). An analgesia circuit activated by cannabinoids. *Nature*. 395: 381-383.
479. Finn, D. P., Jhaveri, M. D., Beckett, S. R., Roe, C. H. and others. (2003). Effects of direct periaqueductal grey administration of a cannabinoid receptor agonist on nociceptive and aversive responses in rats. *Neuropharmacology*. 45: 594-604.
480. Azad, S. C., Monory, K., Marsicano, G., Cravatt, B. F. and others. (2004). Circuitry for associative plasticity in the amygdala involves endocannabinoid signaling. *J.Neurosci*. 24: 9953-9961.
481. Hill, S. Y., Schwin, R., Goodwin, D. W., and Powell, B. J. (1974). Marijuana and pain. *J.Pharmacol.Exp.Ther*. 188: 415-418.
482. Milstein, S. L., MacCannell, K., Karr, G., and Clark, S. (1975). Marijuana-produced changes in pain tolerance. Experienced and non-experienced subjects. *Int.Pharmacopsychiatry*. 10: 177-182.
483. Naef, M., Curatolo, M., Petersen-Felix, S., Arendt-Nielsen, L. and others. (2003). The analgesic effect of oral delta-9-tetrahydrocannabinol (THC), morphine, and a THC-morphine combination in healthy subjects under experimental pain conditions. *Pain*. 105: 79-88.
484. Kraft, B., Frickey, N. A., Kaufmann, R. M., Reif, M. and others. (2008). Lack of analgesia by oral standardized cannabis extract on acute inflammatory pain and hyperalgesia in volunteers. *Anesthesiology*. 109: 101-110.
485. Redmond, W. J., Goffaux, P., Potvin, S., and Marchand, S. (2008). Analgesic and antihyperalgesic effects of nabilone on experimental heat pain. *Curr.Med.Res.Opin*. 24: 1017-1024.
486. Holdcroft, A., Maze, M., Dore, C., Tebbs, S. and others. (2006). A multicenter dose-escalation study of the analgesic and adverse effects of an oral cannabis extract (Cannador) for postoperative pain management. *Anesthesiology*. 104: 1040-1046.
487. Wu, C. L. and Raja, S. N. (2011). Treatment of acute postoperative pain. *Lancet*. 377: 2215-2225.
488. Jain, A. K., Ryan, J. R., McMahon, F. G., and Smith, G. (1981). Evaluation of intramuscular levonantradol and placebo in acute postoperative pain. *J.Clin.Pharmacol*. 21: 320S-326S.
489. Buggy, D. J., Toogood, L., Maric, S., Sharpe, P. and others. (2003). Lack of analgesic efficacy of oral delta-9-tetrahydrocannabinol in postoperative pain. *Pain*. 106: 169-172.
490. Beaulieu, P. (2006). Effects of nabilone, a synthetic cannabinoid, on postoperative pain. *Can.J.Anaesth*. 53: 769-775.
491. Beaulieu, P. (2007). Cannabinoids for postoperative pain. *Anesthesiology*. 106: 397-398.
492. Voscopoulos, C. and Lema, M. (2010). When does acute pain become chronic? *Br.J.Anaesth*. 105 Suppl 1: i69-i85.
493. Fine, P. G., Burton, A. W., and Passik, S. D. (2012). Transformation of acute cancer pain to chronic cancer pain syndromes. *J.Support.Oncol*. 10: 89-95.
494. Berlach, D. M., Shir, Y., and Ware, M. A. (2006). Experience with the synthetic cannabinoid nabilone in chronic noncancer pain. *Pain Med*. 7: 25-29.
495. Walker, J. M. and Huang, S. M. (2002). Cannabinoid analgesia. *Pharmacol.Ther*. 95: 127-135.
496. Rahn, E. J. and Hohmann, A. G. (2009). Cannabinoids as pharmacotherapies for neuropathic pain: from the bench to the bedside. *Neurotherapeutics*. 6: 713-737.

497. Costa, B., Trovato, A. E., Comelli, F., Giagnoni, G. and others. (2007). The non-psychoactive cannabis constituent cannabidiol is an orally effective therapeutic agent in rat chronic inflammatory and neuropathic pain. *Eur.J.Pharmacol.* 556: 75-83.
498. Toth, C. C., Jedrzejewski, N. M., Ellis, C. L., and Frey, W. H. (2010). Cannabinoid-mediated modulation of neuropathic pain and microglial accumulation in a model of murine type I diabetic peripheral neuropathic pain. *Mol.Pain.* 6: 16-
499. Ashton, J. C. and Milligan, E. D. (2008). Cannabinoids for the treatment of neuropathic pain: clinical evidence. *Curr.Opin.Investig.Drugs.* 9: 65-75.
500. Grotenhermen, F. (2007). The toxicology of cannabis and cannabis prohibition. *Chem.Biodivers.* 4: 1744-1769.
501. Wilsey, B., Marcotte, T., Deutsch, R., Gouaux, B. and others. (2012). Low-Dose Vaporized Cannabis Significantly Improves Neuropathic Pain. *J.Pain.* 14: 136-148.
502. Narang, S., Gibson, D., Wasan, A. D., Ross, E. L. and others. (2008). Efficacy of dronabinol as an adjuvant treatment for chronic pain patients on opioid therapy. *J.Pain.* 9: 254-264.
503. Berman, J. S., Symonds, C., and Birch, R. (2004). Efficacy of two cannabis based medicinal extracts for relief of central neuropathic pain from brachial plexus avulsion: results of a randomised controlled trial. *Pain.* 112: 299-306.
504. Rog, D. J., Nurmikko, T. J., Friede, T., and Young, C. A. (2005). Randomized, controlled trial of cannabis-based medicine in central pain in multiple sclerosis. *Neurology.* 65: 812-819.
505. Moulin, D. E., Clark, A. J., Gilron, I., Ware, M. A. and others. (2007). Pharmacological management of chronic neuropathic pain - consensus statement and guidelines from the Canadian Pain Society. *Pain Res.Manag.* 12: 13-21.
506. Fitzcharles, M. A., Ste-Marie, P. A., Goldenberg, D. L., Pereira, J. X., Abbey, S., Choinier, M., Ko, G., Moulin, D., Panopalis, D., Proulx, J., and Shir, Y. 2012 Canadian Guidelines for the Diagnosis and Management of Fibromyalgia Syndrome. 2012.
507. Noyes, R., Jr., Brunk, S. F., Baram, D. A., and Canter, A. (1975). Analgesic effect of delta-9-tetrahydrocannabinol. *J.Clin.Pharmacol.* 15: 139-143.
508. Noyes, R., Jr., Brunk, S. F., Avery, D. A., and Canter, A. C. (1975). The analgesic properties of delta-9-tetrahydrocannabinol and codeine. *Clin.Pharmacol.Ther.* 18: 84-89.
509. Johnson, J. R., Lossignol, D., Burnell-Nugent, M., and Fallon, M. T. (2012). An Open-Label Extension Study to Investigate the Long-Term Safety and Tolerability of THC/CBD Oromucosal Spray and Oromucosal THC Spray in Patients With Terminal Cancer-Related Pain Refractory to Strong Opioid Analgesics. *J.Pain Symptom.Manage.*
510. Bushlin, J., Rozenfeld, R., and Devi, L. A. (2010). Cannabinoid-opioid interactions during neuropathic pain and analgesia. *Curr.Opin.Pharmacol.* 10: 80-86.
511. Desroches, J. and Beaulieu, P. (2010). Opioids and cannabinoids interactions: involvement in pain management. *Curr.Drug Targets.* 11: 462-473.
512. Parolaro, D., Rubino, T., Vigano, D., Massi, P. and others. (2010). Cellular mechanisms underlying the interaction between cannabinoid and opioid system. *Curr.Drug Targets.* 11: 393-405.
513. Rios, C., Gomes, I., and Devi, L. A. (2006). mu opioid and CBI cannabinoid receptor interactions: reciprocal inhibition of receptor signaling and neurogenesis. *Br.J.Pharmacol.* 148: 387-395.

514. Rozenfeld, R., Bushlin, I., Gomes, I., Tzavaras, N. and others. (2012). Receptor heteromerization expands the repertoire of cannabinoid signaling in rodent neurons. *PLoS.One.* 7: e29239-
515. Welch, S. P. and Stevens, D. L. (1992). Antinociceptive activity of intrathecally administered cannabinoids alone, and in combination with morphine, in mice. *J.Pharmacol.Exp.Ther.* 262: 10-18.
516. Pugh, G., Jr., Smith, P. B., Dombrowski, D. S., and Welch, S. P. (1996). The role of endogenous opioids in enhancing the antinociception produced by the combination of delta 9-tetrahydrocannabinol and morphine in the spinal cord. *J.Pharmacol.Exp.Ther.* 279: 608-616.
517. Smith, F. L., Cichewicz, D., Martin, Z. L., and Welch, S. P. (1998). The enhancement of morphine antinociception in mice by delta9-tetrahydrocannabinol. *Pharmacol.Biochem.Behav.* 60: 559-566.
518. Cichewicz, D. L. and McCarthy, E. A. (2003). Antinociceptive synergy between delta(9)-tetrahydrocannabinol and opioids after oral administration. *J.Pharmacol.Exp.Ther.* 304: 1010-1015.
519. Cichewicz, D. L. (2004). Synergistic interactions between cannabinoid and opioid analgesics. *Life Sci.* 74: 1317-1324.
520. Smith, P. A., Selley, D. E., Sim-Selley, L. J., and Welch, S. P. (2007). Low dose combination of morphine and delta9-tetrahydrocannabinol circumvents antinociceptive tolerance and apparent desensitization of receptors. *Eur.J.Pharmacol.* 571: 129-137.
521. Roberts, J. D., Gennings, C., and Shih, M. (2006). Synergistic affective analgesic interaction between delta-9-tetrahydrocannabinol and morphine. *Eur.J.Pharmacol.* 530: 54-58.
522. Greco, R., Gasperi, V., Maccarrone, M., and Tassorelli, C. (2010). The endocannabinoid system and migraine. *Exp.Neurol.* 224: 85-91.
523. Napchan, U., Buse, D. C., and Loder, E. W. (2011). The use of marijuana or synthetic cannabinoids for the treatment of headache. *Headache.* 51: 502-505.
524. McGeeney, B. E. (2012). Hallucinogens and cannabinoids for headache. *Headache.* 52 Suppl 2: 94-97.
525. Russo, E. B. (2004). Clinical endocannabinoid deficiency (CECD): can this concept explain therapeutic benefits of cannabis in migraine, fibromyalgia, irritable bowel syndrome and other treatment-resistant conditions? *Neuro.Endocrinol.Lett.* 25: 31-39.
526. Sarchielli, P., Pini, L. A., Coppola, F., Rossi, C. and others. (2007). Endocannabinoids in chronic migraine: CSF findings suggest a system failure. *Neuropsychopharmacology.* 32: 1384-1390.
527. Villalon, C. M. and Olesen, J. (2009). The role of CGRP in the pathophysiology of migraine and efficacy of CGRP receptor antagonists as acute antimigraine drugs. *Pharmacol.Ther.* 124: 309-323.
528. Cupini, L. M., Costa, C., Sarchielli, P., Bari, M. and others. (2008). Degradation of endocannabinoids in chronic migraine and medication overuse headache. *Neurobiol.Dis.* 30: 186-189.
529. Evans, R. W. and Ramadan, N. M. (2004). Are cannabis-based chemicals helpful in headache? *Headache.* 44: 726-727.
530. Robbins, M. S., Tarshish, S., Solomon, S., and Grosberg, B. M. (2009). Cluster attacks responsive to recreational cannabis and dronabinol. *Headache.* 49: 914-916.
531. Leroux, E., Taifas, I., Valade, D., Donnet, A. and others. (2012). Use of cannabis among 139 cluster headache sufferers. *Cephalalgia.* 33: 208-213.

690. Estrada, G., Fatjo-Vilas, M., Munoz, M. J., Pulido, G. and others. (2011). Cannabis use and age at onset of psychosis: further evidence of interaction with COMT Val158Met polymorphism. *Acta Psychiatr.Scand.* 123: 485-492.
691. van Winkel, R., van Beveren, N. J., and Simons, C. (2011). AKT1 moderation of cannabis-induced cognitive alterations in psychotic disorder. *Neuropsychopharmacology.* 36: 2529-2537.
692. van Winkel, R. (2011). Family-based analysis of genetic variation underlying psychosis-inducing effects of cannabis: sibling analysis and proband follow-up. *Arch.Gen.Psychiatry.* 68: 148-157.
693. Di Forti, M., Iyegbe, C., Sallis, H., Kolliakou, A. and others. (2012). Confirmation that the AKT1 (rs2494732) Genotype Influences the Risk of Psychosis in Cannabis Users. *Biol.Psychiatry.* 72: 811-816.
694. Leweke, F. M., Schneider, U., Radwan, M., Schmidt, E. and others. (2000). Different effects of nabilone and cannabidiol on binocular depth inversion in Man. *Pharmacol.Biochem.Behav.* 66: 175-181.
695. Musty, R. Cannabinoids and anxiety. *Cannabinoids as Therapeutics.* Mechoulam, R. Basel: Birkhäuser, 2005.
696. Zuardi, A. W., Rodrigues, J. A., and Cunha, J. M. (1991). Effects of cannabidiol in animal models predictive of antipsychotic activity. *Psychopharmacology (Berl).* 104: 260-264.
697. Moreira, F. A. and Guimaraes, F. S. (2005). Cannabidiol inhibits the hyperlocomotion induced by psychotomimetic drugs in mice. *Eur.J.Pharmacol.* 512: 199-205.
698. Leweke, F. M., Piomelli, D., Pahlisch, F., Muhl, D. and others. (2012). Cannabidiol enhances anandamide signaling and alleviates psychotic symptoms of schizophrenia. *Transl.Psychiatry.* 2: e94-
699. Zuardi, A. W. (2008). Cannabidiol: from an inactive cannabinoid to a drug with wide spectrum of action. *Rev.Bras.Psiquiatr.* 30: 271-280.
700. Benito, C., Nunez, E., Pazos, M. R., Tolon, R. M. and others. (2007). The endocannabinoid system and Alzheimer's disease. *Mol.Neurobiol.* 36: 75-81.
701. Koppel, J. and Davies, P. (2008). Targeting the endocannabinoid system in Alzheimer's disease. *J.Alzheimers.Dis.* 15: 495-504.
702. van der Stelt, M., Mazzola, C., Esposito, G., Matias, I. and others. (2006). Endocannabinoids and beta-amyloid-induced neurotoxicity in vivo: effect of pharmacological elevation of endocannabinoid levels. *Cell Mol.Life Sci.* 63: 1410-1424.
703. Jung, K. M., Astarita, G., Yasar, S., Vasilevko, V. and others. (2011). An amyloid beta(42)-dependent deficit in anandamide mobilization is associated with cognitive dysfunction in Alzheimer's disease. *Neurobiol.Aging.* 33: 1522-1532.
704. Noonan, J., Tanveer, R., Klompas, A., Gowran, A. and others. (2010). Endocannabinoids prevent beta-amyloid-mediated lysosomal destabilization in cultured neurons. *J.Biol.Chem.* 285: 38543-38554.
705. Eubanks, L. M., Rogers, C. J., Beuscher, A. E., Koob, G. F. and others. (2006). A molecular link between the active component of marijuana and Alzheimer's disease pathology. *Mol.Pharm.* 3: 773-777.
706. Iuvone, T., Esposito, G., Esposito, R., Santamaria, R. and others. (2004). Neuroprotective effect of cannabidiol, a non-psychoactive component from *Cannabis sativa*, on beta-amyloid-induced toxicity in PC12 cells. *J.Neurochem.* 89: 134-141.

532. Ducros, A., Boukobza, M., Porcher, R., Sarov, M. and others. (2007). The clinical and radiological spectrum of reversible cerebral vasoconstriction syndrome. A prospective series of 67 patients. *Brain*. 130: 3091-3101.
533. Levin, K. H., Copersino, M. L., Heishman, S. J., Liu, F. and others. (2010). Cannabis withdrawal symptoms in non-treatment-seeking adult cannabis smokers. *Drug Alcohol Depend.* 111: 120-127.
534. Dunkley, L and Tattersall, R. (2012). Osteoarthritis and the inflammatory arthritides. *Surgery*. 30: 67-71.
535. Sagar, D. R., Staniaszek, L. E., Okine, B. N., Woodhams, S. and others. (2010). Tonic modulation of spinal hyperexcitability by the endocannabinoid receptor system in a rat model of osteoarthritis pain. *Arthritis Rheum.* 62: 3666-3676.
536. Schuelert, N. and McDougall, J. J. (2008). Cannabinoid-mediated antinociception is enhanced in rat osteoarthritic knees. *Arthritis Rheum.* 58: 145-153.
537. Schuelert, N., Johnson, M. P., Oskins, J. L., Jassal, K. and others. (2011). Local application of the endocannabinoid hydrolysis inhibitor URB597 reduces nociception in spontaneous and chemically induced models of osteoarthritis. *Pain*. 152: 975-981.
538. Richards, B. L., Whittle, S. L., and Buchbinder, R. (2012). Neuromodulators for pain management in rheumatoid arthritis. *Cochrane.Database.Syst.Rev.* 1: CD008921-
539. McInnes, I. B. and Schett, G. (2011). The pathogenesis of rheumatoid arthritis. *N.Engl.J.Med.* 365: 2205-2219.
540. Smith, H. S., Bracken, D., and Smith, J. M. (2011). Pharmacotherapy for fibromyalgia. *Front Pharmacol.* 2: 17-
541. Julien, N., Goffaux, P., Arsenault, P., and Marchand, S. (2005). Widespread pain in fibromyalgia is related to a deficit of endogenous pain inhibition. *Pain*. 114: 295-302.
542. Clauw, D. J., Arnold, L. M., and McCarberg, B. H. (2011). The science of fibromyalgia. *Mayo Clin.Proc.* 86: 907-911.
543. Normand, E., Potvin, S., Gaumont, I., Cloutier, G. and others. (2011). Pain inhibition is deficient in chronic widespread pain but normal in major depressive disorder. *J.Clin.Psychiatry.* 72: 219-224.
544. Becker, S. and Schweinhardt, P. (2012). Dysfunctional neurotransmitter systems in fibromyalgia, their role in central stress circuitry and pharmacological actions on these systems. *Pain Res.Treat.* 2012: 741746-
545. de Souza, J. B., Potvin, S., Goffaux, P., Charest, J. and others. (2009). The deficit of pain inhibition in fibromyalgia is more pronounced in patients with comorbid depressive symptoms. *Clin.J.Pain.* 25: 123-127.
546. Wolfe, F., Clauw, D. J., Fitzcharles, M. A., Goldenberg, D. L. and others. (2010). The American College of Rheumatology preliminary diagnostic criteria for fibromyalgia and measurement of symptom severity. *Arthritis Care Res.(Hoboken.)*. 62: 600-610.
547. Idris, A. I., Sophocleous, A., Landao-Bassonga, E., Canals, M. and others. (2009). Cannabinoid receptor type 1 protects against age-related osteoporosis by regulating osteoblast and adipocyte differentiation in marrow stromal cells. *Cell Metab.* 10: 139-147.
548. Bab, I., Smoum, R., Bradshaw, H., and Mechoulam, R. (2011). Skeletal lipidomics: regulation of bone metabolism by fatty acid amide family. *Br.J.Pharmacol.* 163: 1441-1446.
549. Idris, A. I., van 't Hof, R. J., Greig, I. R., Ridge, S. A. and others. (2005). Regulation of bone mass, bone loss and osteoclast activity by cannabinoid receptors. *Nat.Med.* 11: 774-779.

550. Whyte, L. S., Ford, L., Ridge, S. A., Cameron, G. A. and others. (2012). Cannabinoids and bone: endocannabinoids modulate human osteoclast function in vitro. *Br.J.Pharmacol.* 165: 2584-2597.
551. Tam, J., Ofek, O., Fride, E., Ledent, C. and others. (2006). Involvement of neuronal cannabinoid receptor CB1 in regulation of bone mass and bone remodeling. *Mol.Pharmacol.* 70: 786-792.
552. Tam, J., Trembovier, V., Di, Marzo, V., Petrosino, S. and others. (2008). The cannabinoid CB1 receptor regulates bone formation by modulating adrenergic signaling. *FASEB J.* 22: 285-294.
553. Rossi, F., Siniscalco, D., Luongo, L., De, Petrocellis L. and others. (2009). The endovanilloid/endocannabinoid system in human osteoclasts: possible involvement in bone formation and resorption. *Bone.* 44: 476-484.
554. Ofek, O., Karsak, M., Leclerc, N., Fogel, M. and others. (2006). Peripheral cannabinoid receptor, CB2, regulates bone mass. *Proc.Natl.Acad.Sci.U.S.A.* 103: 696-701.
555. Sophocleous, A., Landao-Bassonga, E., van't Hof, R. J., Idris, A. I. and others. (2011). The type 2 cannabinoid receptor regulates bone mass and ovariectomy-induced bone loss by affecting osteoblast differentiation and bone formation. *Endocrinology.* 152: 2141-2149.
556. Idris, A. I., Sophocleous, A., Landao-Bassonga, E., van't Hof, R. J. and others. (2008). Regulation of bone mass, osteoclast function, and ovariectomy-induced bone loss by the type 2 cannabinoid receptor. *Endocrinology.* 149: 5619-5626.
557. Karsak, M., Cohen-Sotai, M., Freudenberg, J., Ostertag, A. and others. (2005). Cannabinoid receptor type 2 gene is associated with human osteoporosis. *Hum.Mol.Genet.* 14: 3389-3396.
558. Karsak, M., Malkin, I., Toliat, M. R., Kubisch, C. and others. (2009). The cannabinoid receptor type 2 (CNR2) gene is associated with hand bone strength phenotypes in an ethnically homogeneous family sample. *Hum.Genet.* 126: 629-636.
559. Huang, Q. Y., Li, G. H., and Kung, A. W. (2009). Multiple osteoporosis susceptibility genes on chromosome 1p36 in Chinese. *Bone.* 44: 984-988.
560. Fernandez-Ruiz, J. (2009). The endocannabinoid system as a target for the treatment of motor dysfunction. *Br.J.Pharmacol.* 156: 1029-1040.
561. Glass, M., Dragunow, M., and Faull, R. L. (2000). The pattern of neurodegeneration in Huntington's disease: a comparative study of cannabinoid, dopamine, adenosine and GABA(A) receptor alterations in the human basal ganglia in Huntington's disease. *Neuroscience.* 97: 505-519.
562. Romero, J., Berrendero, F., Perez-Rosado, A., Manzanares, J. and others. (2000). Unilateral 6-hydroxydopamine lesions of nigrostriatal dopaminergic neurons increased CB1 receptor mRNA levels in the caudate-putamen. *Life Sci.* 66: 485-494.
563. Lastres-Becker, I., Fezza, F., Cebeira, M., Bisogno, T. and others. (2001). Changes in endocannabinoid transmission in the basal ganglia in a rat model of Huntington's disease. *Neuroreport.* 12: 2125-2129.
564. Garcia-Arencibia, M., Garcia, C., and Fernandez-Ruiz, J. (2009). Cannabinoids and Parkinson's disease. *CNS.Neurol.Disord.Drug Targets.* 8: 432-439.
565. Richter, A. and Loscher, W. (1994). (+)-WIN 55,212-2, a novel cannabinoid receptor agonist, exerts antidystonic effects in mutant dystonic hamsters. *Eur.J.Pharmacol.* 264: 371-377.
566. Richter, A. and Loscher, W. (2002). Effects of pharmacological manipulations of cannabinoid receptors on severity of dystonia in a genetic model of paroxysmal dyskinesia. *Eur.J.Pharmacol.* 454: 145-151.

567. Madsen, M. V., Peacock, L. P., Werge, T., Andersen, M. B. and others. (2011). Effects of cannabinoid CB(1) receptor agonism and antagonism on SKF81297-induced dyskinesia and haloperidol-induced dystonia in *Cebus apella* monkeys. *Neuropharmacology*. 60: 418-422.
568. Uribe Roca, M. C., Micheli, F., and Viotti, R. (2005). Cannabis sativa and dystonia secondary to Wilson's disease. *Mov Disord*. 20: 113-115.
569. Jabusch, H. C., Schneider, U., and Altenmuller, E. (2004). Delta9-tetrahydrocannabinol improves motor control in a patient with musician's dystonia. *Mov Disord*. 19: 990-991.
570. Consroe, P., Sandyk, R., and Snider, S. R. (1986). Open label evaluation of cannabidiol in dystonic movement disorders. *Int.J.Neurosci*. 30: 277-282.
571. Fox, S. H., Kellett, M., Moore, A. P., Crossman, A. R. and others. (2002). Randomised, double-blind, placebo-controlled trial to assess the potential of cannabinoid receptor stimulation in the treatment of dystonia. *Mov Disord*. 17: 145-149.
572. Denovan-Wright, E. M. and Robertson, H. A. (2000). Cannabinoid receptor messenger RNA levels decrease in a subset of neurons of the lateral striatum, cortex and hippocampus of transgenic Huntington's disease mice. *Neuroscience*. 98: 705-713.
573. Lastres-Becker, I., Gomez, M., de Miguel R., Ramos, J. A. and others. (2002). Loss of cannabinoid CB(1) receptors in the basal ganglia in the late akinetic phase of rats with experimental Huntington's disease. *Neurotox.Res*. 4: 601-608.
574. Naver, B., Stub, C., Molier, M., Fenger, K. and others. (2003). Molecular and behavioral analysis of the R6/1 Huntington's disease transgenic mouse. *Neuroscience*. 122: 1049-1057.
575. McCaw, E. A., Hu, H., Gomez, G. T., Hebb, A. L. and others. (2004). Structure, expression and regulation of the cannabinoid receptor gene (CB1) in Huntington's disease transgenic mice. *Eur.J.Biochem*. 271: 4909-4920.
576. Centonze, D., Rossi, S., Prosperetti, C., Tschertner, A. and others. (2005). Abnormal sensitivity to cannabinoid receptor stimulation might contribute to altered gamma-aminobutyric acid transmission in the striatum of R6/2 Huntington's disease mice. *Biol.Psychiatry*. 57: 1583-1589.
577. Pazos, M. R., Sagredo, O., and Fernandez-Ruiz, J. (2008). The endocannabinoid system in Huntington's disease. *Curr.Pharm.Des*. 14: 2317-2325.
578. Dowie, M. J., Bradshaw, H. B., Howard, M. L., Nicholson, L. F. and others. (2009). Altered CB1 receptor and endocannabinoid levels precede motor symptom onset in a transgenic mouse model of Huntington's disease. *Neuroscience*. 163: 456-465.
579. Blazquez, C., Chiarlone, A., Sagredo, O., Aguado, T. and others. (2011). Loss of striatal type 1 cannabinoid receptors is a key pathogenic factor in Huntington's disease. *Brain*. 134: 119-136.
580. Casteels, C., Vandeputte, C., Rangarajan, J. R., Dresselaers, T. and others. (2011). Metabolic and type 1 cannabinoid receptor imaging of a transgenic rat model in the early phase of Huntington disease. *Exp.Neurol*. 229: 440-449.
581. Mievis, S., Blum, D., and Ledent, C. (2011). Worsening of Huntington disease phenotype in CB1 receptor knockout mice. *Neurobiol.Dis*. 42: 524-529.
582. Van Laere K., Casteels, C., Dhallander, I., Goffin, K. and others. (2010). Widespread decrease of type 1 cannabinoid receptor availability in Huntington disease in vivo. *J.Nucl.Med*. 51: 1413-1417.

583. Palazuelos, J., Aguado, T., Pazos, M. R., Julien, B. and others. (2009). Microglial CB2 cannabinoid receptors are neuroprotective in Huntington's disease excitotoxicity. *Brain*. 132: 3152-3164.
584. Dowie, M. J., Howard, M. L., Nicholson, L. F., Faull, R. L. and others. (2010). Behavioural and molecular consequences of chronic cannabinoid treatment in Huntington's disease transgenic mice. *Neuroscience*. 170: 324-336.
585. Consroe, P., Laguna, J., Allender, J., Snider, S. and others. (1991). Controlled clinical trial of cannabidiol in Huntington's disease. *Pharmacol.Biochem.Behav.* 40: 701-708.
586. Curtis, A., Mitchell, I., Patel, S., Ives, N. and others. (2009). A pilot study using nabilone for symptomatic treatment in Huntington's disease. *Mov Disord*. 24: 2254-2259.
587. Muller-Vahl, K. R., Schneider, U., and Emrich, H. M. (1999). Nabilone increases choreatic movements in Huntington's disease. *Mov Disord*. 14: 1038-1040.
588. Curtis, A. and Rickards, H. (2006). Nabilone could treat chorea and irritability in Huntington's disease. *J.Neuropsychiatry Clin.Neurosci.* 18: 553-554.
589. Pisani, V., Moschella, V., Bari, M., Fezza, F. and others. (2010). Dynamic changes of anandamide in the cerebrospinal fluid of Parkinson's disease patients. *Mov Disord*. 25: 920-924.
590. Pisani, A., Fezza, F., Galati, S., Battista, N. and others. (2005). High endogenous cannabinoid levels in the cerebrospinal fluid of untreated Parkinson's disease patients. *Ann.Neurol.* 57: 777-779.
591. Garcia-Arencibia, M., Garcia, C., Kurz, A., Rodriguez-Navarro, J. A. and others. (2009). Cannabinoid CB1 receptors are early downregulated followed by a further upregulation in the basal ganglia of mice with deletion of specific park genes. *J.Neural Transm.Suppl.* 269-275.
592. Papa, S. M. (2008). The cannabinoid system in Parkinson's disease: multiple targets to motor effects. *Exp.Neurol.* 211: 334-338.
593. Garcia, C., Palomo, C., Garcia-Arencibia, M., Ramos, J. A. and others. (2011). Symptom-relieving and neuroprotective effects of the phytocannabinoid D(9) -THCV in animal models of Parkinson's disease. *Br.J.Pharmacol.* 163: 1495-1506.
594. Frankel, J. P., Hughes, A., Lees, A. J., and Stern, G. M. (1990). Marijuana for parkinsonian tremor. *J.Neurol.Neurosurg.Psychiatry.* 53: 436.-
595. Sieradzan, K. A., Fox, S. H., Hill, M., Dick, J. P. and others. (2001). Cannabinoids reduce levodopa-induced dyskinesia in Parkinson's disease: a pilot study. *Neurology.* 57: 2108-2111.
596. Carroll, C. B., Bain, P. G., Teare, L., Liu, X. and others. (2004). Cannabis for dyskinesia in Parkinson disease: a randomized double-blind crossover study. *Neurology.* 63: 1245-1250.
597. Sandyk, R. and Awerbuch, G. (1988). Marijuana and Tourette's syndrome. *J.Clin.Psychopharmacol.* 8: 444-445.
598. Hemming, M. and Yellowlees, P. M. (1993). Effective treatment of Tourette's syndrome with marijuana. *Journal of Psychopharmacology.* 7: 389-391.
599. Muller-Vahl, K. R., Koblenz, A., Jobges, M., Kolbe, H. and others. (2001). Influence of treatment of Tourette syndrome with delta9-tetrahydrocannabinol (delta9-THC) on neuropsychological performance. *Pharmacopsychiatry.* 34: 19-24.

600. Muller-Vahl, K. R., Schneider, U., Prevedel, H., Theloe, K. and others. (2003). Delta 9-tetrahydrocannabinol (THC) is effective in the treatment of tics in Tourette syndrome: a 6-week randomized trial. *J.Clin.Psychiatry*. 64: 459-465.
601. Curtis, A., Clarke, C. E., and Rickards, H. E. (2009). Cannabinoids for Tourette's Syndrome. *Cochrane.Database.Syst.Rev*. CD006565.-
602. Cheung, W., Guo, L., and Cordeiro, M. F. (2008). Neuroprotection in glaucoma: drug-based approaches. *Optom.Vis.Sci*. 85: 406-416.
603. Jarvinen, T., Pate, D. W., and Laine, K. (2002). Cannabinoids in the treatment of glaucoma. *Pharmacol.Ther*. 95: 203-220.
604. Jampel, H. (2010). American glaucoma society position statement: marijuana and the treatment of glaucoma. *J.Glaucoma*. 19: 75-76.
605. Chen, J., Matias, I., Dinh, T., Lu, T. and others. (2005). Finding of endocannabinoids in human eye tissues: implications for glaucoma. *Biochem.Biophys.Res.Commun*. 330: 1062-1067.
606. Porcella, A., Casellas, P., Gessa, G. L., and Pani, L. (1998). Cannabinoid receptor CB1 mRNA is highly expressed in the rat ciliary body: implications for the antiglaucoma properties of marihuana. *Brain Res.Mol.Brain Res*. 58: 240-245.
607. Straiker, A. J., Maguire, G., Mackie, K., and Lindsey, J. (1999). Localization of cannabinoid CB1 receptors in the human anterior eye and retina. *Invest Ophthalmol.Vis.Sci*. 40: 2442-2448.
608. Porcella, A., Maxia, C., Gessa, G. L., and Pani, L. (2000). The human eye expresses high levels of CB1 cannabinoid receptor mRNA and protein. *Eur.J.Neurosci*. 12: 1123-1127.
609. Song, Z. H. and Slowey, C. A. (2000). Involvement of cannabinoid receptors in the intraocular pressure-lowering effects of WIN55212-2. *J.Pharmacol.Exp.Ther*. 292: 136-139.
610. Porcella, A., Maxia, C., Gessa, G. L., and Pani, L. (2001). The synthetic cannabinoid WIN55212-2 decreases the intraocular pressure in human glaucoma resistant to conventional therapies. *Eur.J.Neurosci*. 13: 409-412.
611. Flach, A. J. (2002). Delta-9-tetrahydrocannabinol (THC) in the treatment of end-stage open-angle glaucoma. *Trans.Am.Ophthalmol.Soc*. 100: 215-222.
612. Yoles, E., Belkin, M., and Schwartz, M. (1996). HU-211, a nonpsychotropic cannabinoid, produces short- and long-term neuroprotection after optic nerve axotomy. *J.Neurotrauma*. 13: 49-57.
613. Shen, M. and Thayer, S. A. (1998). Cannabinoid receptor agonists protect cultured rat hippocampal neurons from excitotoxicity. *Mol.Pharmacol*. 54: 459-462.
614. Levin, L. A. (1999). Direct and indirect approaches to neuroprotective therapy of glaucomatous optic neuropathy. *Surv.Ophthalmol*. 43 Suppl 1: S98-101.
615. Jin, K. L., Mao, X. O., Goldsmith, P. C., and Greenberg, D. A. (2000). CB1 cannabinoid receptor induction in experimental stroke. *Ann.Neuro*. 48: 257-261.
616. Panikashvili, D., Simeonidou, C., Ben-Shabat, S., Hanus, L. and others. (2001). An endogenous cannabinoid (2-AG) is neuroprotective after brain injury. *Nature*. 413: 527-531.
617. Marsicano, G., Moosmann, B., Hermann, H., Lutz, B. and others. (2002). Neuroprotective properties of cannabinoids against oxidative stress: role of the cannabinoid receptor CB1. *J.Neurochem*. 80: 448-456.

618. Mechoulam, R., Panikashvili, D., and Shohami, E. (2002). Cannabinoids and brain injury: therapeutic implications. *Trends Mol.Med.* 8: 58-61.
619. Braida, D., Pegorini, S., Arcidiacono, M. V., Consalez, G. G. and others. (2003). Post-ischemic treatment with cannabidiol prevents electroencephalographic flattening, hyperlocomotion and neuronal injury in gerbils. *Neurosci.Lett.* 346: 61-64.
620. El-Remessy, A. B., Al-Shabrawey, M., Khalifa, Y., Tsai, N. T. and others. (2006). Neuroprotective and blood-retinal barrier-preserving effects of cannabidiol in experimental diabetes. *Am.J.Pathol.* 168: 235-244.
621. Gilbert, G. L., Kim, H. J., Waataja, J. J., and Thayer, S. A. (2007). Delta9-tetrahydrocannabinol protects hippocampal neurons from excitotoxicity. *Brain Res.* 1128: 61-69.
622. Wan, M. J., Daniel, S., Kassam, F., Mutti, G. and others. (2012). Survey of complementary and alternative medicine use in glaucoma patients. *J.Glaucoma.* 21: 79-82.
623. Hepner, R. S. and Frank, I. R. (1971). Marijuana smoking and intraocular pressure. *JAMA.* 217: 1392.-
624. Merritt, J. C., Crawford, W. J., Alexander, P. C., Anduze, A. L. and others. (1980). Effect of marijuana on intraocular and blood pressure in glaucoma. *Ophthalmology.* 87: 222-228.
625. Zhan, G. L., Camras, C. B., Palmberg, P. F., and Toris, C. B. (2005). Effects of marijuana on aqueous humor dynamics in a glaucoma patient. *J.Glaucoma.* 14: 175-177.
626. Abboud, R. T. and Sanders, H. D. (1976). Effect of oral administration of delta-tetrahydrocannabinol on airway mechanics in normal and asthmatic subjects. *Chest.* 70: 480-485.
627. Calignano, A., Katona, I., Desarnaud, F., Giuffrida, A. and others. (2000). Bidirectional control of airway responsiveness by endogenous cannabinoids. *Nature.* 408: 96-101.
628. Jan, T. R., Farraj, A. K., Harkema, J. R., and Kaminski, N. E. (2003). Attenuation of the ovalbumin-induced allergic airway response by cannabinoid treatment in A/J mice. *Toxicol.Appl.Pharmacol.* 188: 24-35.
629. Giannini, L., Nistri, S., Mastroianni, R., Cinci, L. and others. (2008). Activation of cannabinoid receptors prevents antigen-induced asthma-like reaction in guinea pigs. *J.Cell Mol.Med.* 12: 2381-2394.
630. Fukuda, H., Abe, T., and Yoshihara, S. (2010). The cannabinoid receptor agonist WIN 55,212-2 inhibits antigen-induced plasma extravasation in guinea pig airways. *Int.Arch.Allergy Immunol.* 152: 295-300.
631. Vachon, L., FitzGerald, M. X., Solliday, N. H., Gould, I. A. and others. (1973). Single-dose effects of marijuana smoke. Bronchial dynamics and respiratory-center sensitivity in normal subjects. *N.Engl.J.Med.* 288: 985-989.
632. Tashkin, D. P., Shapiro, B. J., and Frank, I. M. (1973). Acute pulmonary physiologic effects of smoked marijuana and oral 9-tetrahydrocannabinol in healthy young men. *N.Engl.J.Med.* 289: 336-341.
633. Tashkin, D. P. (2001). Airway effects of marijuana, cocaine, and other inhaled illicit agents. *Curr.Opin.Pulm.Med.* 7: 43-61.
634. Tashkin, D. P., Shapiro, B. J., and Frank, I. M. (1974). Acute effects of smoked marijuana and oral delta9-tetrahydrocannabinol on specific airway conductance in asthmatic subjects. *Am.Rev.Respir.Dis.* 109: 420-428.
635. Tashkin, D. P., Shapiro, B. J., Lee, Y. E., and Harper, C. E. (1975). Effects of smoked marijuana in experimentally induced asthma. *Am.Rev.Respir.Dis.* 112: 377-386.

636. Gong, H., Jr., Tashkin, D. P., Simmons, M. S., Calvarese, B. and others. (1984). Acute and subacute bronchial effects of oral cannabinoids. *Clin.Pharmacol.Ther.* 35: 26-32.
637. Williams, S. J., Hartley, J. P., and Graham, J. D. (1976). Bronchodilator effect of delta1-tetrahydrocannabinol administered by aerosol of asthmatic patients. *Thorax.* 31: 720-723.
638. Hartley, J. P., Nogrady, S. G., and Seaton, A. (1978). Bronchodilator effect of delta1-tetrahydrocannabinol. *Br.J.Clin.Pharmacol.* 5: 523-525.
639. Tashkin, D. P., Shapiro, B. J., Lee, Y. E., and Harper, C. E. (1976). Subacute effects of heavy marihuana smoking on pulmonary function in healthy men. *N.Engl.J.Med.* 294: 125-129.
640. Davies, B. H., Radcliffe, S., Seaton, A., and Graham, J. D. (1975). A trial of oral delta-1-(trans)-tetrahydrocannabinol in reversible airways obstruction. *Thorax.* 30: 80-85.
641. Gong, H., Jr., Tashkin, D. P., and Calvarese, B. (1983). Comparison of bronchial effects of nabilone and terbutaline in healthy and asthmatic subjects. *J.Clin.Pharmacol.* 23: 127-133.
642. Pacher, P., Batkai, S., and Kunos, G. (2005). Cardiovascular pharmacology of cannabinoids. *Handb.Exp.Pharmacol.* 599-625.
643. Crawford, W. J. and Merritt, J. C. (1979). Effects of tetrahydrocannabinol on arterial and intraocular hypertension. *Int.J.Clin.Pharmacol.Biopharm.* 17: 191-196.
644. Denson, T. F. and Earleywine, M. (2006). Decreased depression in marijuana users. *Addict.Behav.* 31: 738-742.
645. Witkin, J. M., Tzavara, E. T., and Nomikos, G. G. (2005). A role for cannabinoid CB1 receptors in mood and anxiety disorders. *Behav.Pharmacol.* 16: 315-331.
646. Moreira, F. A. and Wotjak, C. T. (2010). Cannabinoids and anxiety. *Curr.Top.Behav.Neurosci.* 2: 429-450.
647. Bambico, F. R., Katz, N., Debonnel, G., and Gobbi, G. (2007). Cannabinoids elicit antidepressant-like behavior and activate serotonergic neurons through the medial prefrontal cortex. *J.Neurosci.* 27: 11700-11711.
648. Bambico, F. R. and Gobbi, G. (2008). The cannabinoid CB1 receptor and the endocannabinoid anandamide: possible antidepressant targets. *Expert.Opin.Ther.Targets.* 12: 1347-1366.
649. Hill, M. N. and Gorzalka, B. B. (2005). Pharmacological enhancement of cannabinoid CB1 receptor activity elicits an antidepressant-like response in the rat forced swim test. *Eur.Neuropsychopharmacol.* 15: 593-599.
650. Christensen, R., Kristensen, P. K., Bartels, E. M., Bliddal, H. and others. (2007). Efficacy and safety of the weight-loss drug rimonabant: a meta-analysis of randomised trials. *Lancet.* 370: 1706-1713.
651. Gorzalka, B. B. and Hill, M. N. (2010). Putative role of endocannabinoid signaling in the etiology of depression and actions of antidepressants. *Prog.Neuropsychopharmacol.Biol.Psychiatry.* 35: 1575-1585.
652. Hill, M. N., Miller, G. E., Carrier, E. J., Gorzalka, B. B. and others. (2009). Circulating endocannabinoids and N-acyl ethanolamines are differentially regulated in major depression and following exposure to social stress. *Psychoneuroendocrinology.* 34: 1257-1262.
653. Woolridge, E., Barton, S., Samuel, J., Osorio, J. and others. (2005). Cannabis use in HIV for pain and other medical symptoms. *J.Pain Symptom.Manage.* 29: 358-367.
654. Page, S. A. and Verhoef, M. J. (2006). Medicinal marijuana use: experiences of people with multiple sclerosis. *Can.Fam.Physician.* 52: 64-65.

655. Mitchell, P. B. and Morris, M. J. (2007). Depression and anxiety with rimonabant. *Lancet*. 370: 1671-1672.
656. Morgan, C. J., Schafer, G., Freeman, T. P., and Curran, H. V. (2010). Impact of cannabidiol on the acute memory and psychotomimetic effects of smoked cannabis: naturalistic study: naturalistic study [corrected]. *Br.J.Psychiatry*. 197: 285-290.
657. Guimaraes, F. S., de Aguiar, J. C., Mechoulam, R., and Breuer, A. (1994). Anxiolytic effect of cannabidiol derivatives in the elevated plus-maze. *Gen.Pharmacol*. 25: 161-164.
658. Crippa, J. A., Zuardi, A. W., Garrido, G. E., Wichert-Ana, L. and others. (2004). Effects of cannabidiol (CBD) on regional cerebral blood flow. *Neuropsychopharmacology*. 29: 417-426.
659. Bergamaschi, M. M., Queiroz, R. H., Chagas, M. H., de Oliveira, D. C. and others. (2011). Cannabidiol reduces the anxiety induced by simulated public speaking in treatment-naive social phobia patients. *Neuropsychopharmacology*. 36: 1219-1226.
660. Resstel, L. B., Tavares, R. F., Lisboa, S. F., Joca, S. R. and others. (2009). 5-HT1A receptors are involved in the cannabidiol-induced attenuation of behavioural and cardiovascular responses to acute restraint stress in rats. *Br.J.Pharmacol*. 156: 181-188.
661. Gomes, F. V., Resstel, L. B., and Guimaraes, F. S. (2011). The anxiolytic-like effects of cannabidiol injected into the bed nucleus of the stria terminalis are mediated by 5-HT1A receptors. *Psychopharmacology (Berl)*. 213: 465-473.
662. Koethe, D., Schreiber, D., Giuffrida, A., Mauss, C. and others. (2009). Sleep deprivation increases oleylethanolamide in human cerebrospinal fluid. *J.Neural Transm*. 116: 301-305.
663. Herrera-Solis, A., Vasquez, K. G., and Prospero-Garcia, O. (2010). Acute and subchronic administration of anandamide or oleamide increases REM sleep in rats. *Pharmacol.Biochem.Behav*. 95: 106-112.
664. Bolla, K. I., Lesage, S. R., Gamaldo, C. E., Neubauer, D. N. and others. (2008). Sleep disturbance in heavy marijuana users. *Sleep*. 31: 901-908.
665. Bolla, K. I., Lesage, S. R., Gamaldo, C. E., Neubauer, D. N. and others. (2010). Polysomnogram changes in marijuana users who report sleep disturbances during prior abstinence. *Sleep Med*. 11: 882-889.
666. Lutz, B. (2007). The endocannabinoid system and extinction learning. *Mol.Neurobiol*. 36: 92-101.
667. Pamplona, F. A., Prediger, R. D., Pandolfo, P., and Takahashi, R. N. (2006). The cannabinoid receptor agonist WIN 55,212-2 facilitates the extinction of contextual fear memory and spatial memory in rats. *Psychopharmacology (Berl)*. 188: 641-649.
668. Marsicano, G., Wotjak, C. T., Azad, S. C., Bisogno, T. and others. (2002). The endogenous cannabinoid system controls extinction of aversive memories. *Nature*. 418: 530-534.
669. Varvel, S. A. and Lichtman, A. H. (2002). Evaluation of CB1 receptor knockout mice in the Morris water maze. *J.Pharmacol.Exp.Ther*. 301: 915-924.
670. Chhatwal, J. P., Davis, M., Maguschak, K. A., and Ressler, K. J. (2005). Enhancing cannabinoid neurotransmission augments the extinction of conditioned fear. *Neuropsychopharmacology*. 30: 516-524.
671. Pava, M. J. and Woodward, J. J. (2012). A review of the interactions between alcohol and the endocannabinoid system: implications for alcohol dependence and future directions for research. *Alcohol*. 46: 185-204.
672. Deikel, S. M. and Carder, B. (1976). Attenuation of precipitated abstinence in methadone-dependent rats by delta9-THC. *Psychopharmacol.Commun*. 2: 61-65.

673. Vela, G., Fuentes, J. A., Bonnin, A., Fernandez-Ruiz, J. and others. (1995). Perinatal exposure to delta 9-tetrahydrocannabinol (delta 9-THC) leads to changes in opioid-related behavioral patterns in rats. *Brain Res.* 680: 142-147.
674. Yamaguchi, T., Hagiwara, Y., Tanaka, H., Sugiura, T. and others. (2001). Endogenous cannabinoid, 2-arachidonoylglycerol, attenuates naloxone-precipitated withdrawal signs in morphine-dependent mice. *Brain Res.* 909: 121-126.
675. Reiman, A. (2009). Cannabis as a substitute for alcohol and other drugs. *Harm.Reduct.J.* 6: 35-39.
676. Fernandez-Espejo, E., Viveros, M. P., Nunez, L., Ellenbroek, B. A. and others. (2009). Role of cannabis and endocannabinoids in the genesis of schizophrenia. *Psychopharmacology (Berl)*. 206: 531-549.
677. Koethe, D., Giuffrida, A., Schreiber, D., Hellmich, M. and others. (2009). Anandamide elevation in cerebrospinal fluid in initial prodromal states of psychosis. *Br.J.Psychiatry.* 194: 371-372.
678. Leweke, F. M., Giuffrida, A., Wurster, U., Emrich, H. M. and others. (1999). Elevated endogenous cannabinoids in schizophrenia. *Neuroreport.* 10: 1665-1669.
679. De Marchi, N., De Petrocellis, L., Orlando, P., Daniele, F. and others. (2003). Endocannabinoid signalling in the blood of patients with schizophrenia. *Lipids Health Dis.* 2: 5.-
680. Rodriguez de Fonseca, F., Gorriti, M. A., Bilbao, A., Escuredo, L. and others. (2001). Role of the endogenous cannabinoid system as a modulator of dopamine transmission: implications for Parkinson's disease and schizophrenia. *Neurotox.Res.* 3: 23-35.
681. McCreadie, R. G. (2002). Use of drugs, alcohol and tobacco by people with schizophrenia: case-control study. *Br.J.Psychiatry.* 181: 321-325.
682. Gregg, L., Barrowclough, C., and Haddock, G. (2007). Reasons for increased substance use in psychosis. *Clin.Psychol.Rev.* 27: 494-510.
683. Khantzian, E. J. (1985). The self-medication hypothesis of addictive disorders: focus on heroin and cocaine dependence. *Am.J.Psychiatry.* 142: 1259-1264.
684. Lembke, A. (2012). Time to abandon the self-medication hypothesis in patients with psychiatric disorders. *Am.J Drug Alcohol Abuse.* 38: 524-529.
685. Chambers, R. A., Krystal, J. H., and Self, D. W. (2001). A neurobiological basis for substance abuse comorbidity in schizophrenia. *Biol.Psychiatry.* 50: 71-83.
686. Caspi, A., Moffitt, T. E., Cannon, M., McClay, J. and others. (2005). Moderation of the effect of adolescent-onset cannabis use on adult psychosis by a functional polymorphism in the catechol-O-methyltransferase gene: longitudinal evidence of a gene X environment interaction. *Biol.Psychiatry.* 57: 1117-1127.
687. Henquet, C., Rosa, A., Krabbendam, L., Papiol, S. and others. (2006). An experimental study of catechol-o-methyltransferase Val158Met moderation of delta-9-tetrahydrocannabinol-induced effects on psychosis and cognition. *Neuropsychopharmacology.* 31: 2748-2757.
688. Henquet, C., Rosa, A., Delespaul, P., Papiol, S. and others. (2009). COMT ValMet moderation of cannabis-induced psychosis: a momentary assessment study of 'switching on' hallucinations in the flow of daily life. *Acta Psychiatr.Scand.* 119: 156-160.
689. Pelayo-Teran, J. M., Perez-Iglesias, R., Mata, I., Carrasco-Marin, E. and others. (2010). Catechol-O-Methyltransferase (COMT) Val158Met variations and cannabis use in first-episode non-affective psychosis: clinical-onset implications. *Psychiatry Res.* 179: 291-296.

707. Esposito, G., De Filippis D., Carnuccio, R., Izzo, A. A. and others. (2006). The marijuana component cannabidiol inhibits beta-amyloid-induced tau protein hyperphosphorylation through Wnt/beta-catenin pathway rescue in PC12 cells. *J.Mol.Med.(Berl)*. 84: 253-258.
708. Booz, G. W. (2011). Cannabidiol as an emergent therapeutic strategy for lessening the impact of inflammation on oxidative stress. *Free Radic.Biol.Med*. 51: 1054-1061.
709. Esposito, G., Scuderi, C., Savani, C., Steardo, L., Jr. and others. (2007). Cannabidiol in vivo blunts beta-amyloid induced neuroinflammation by suppressing IL-1beta and iNOS expression. *Br.J.Pharmacol*. 151: 1272-1279.
710. Esposito, G., Iuvone, T., Savani, C., Scuderi, C. and others. (2007). Opposing control of cannabinoid receptor stimulation on amyloid-beta-induced reactive gliosis: in vitro and in vivo evidence. *J.Pharmacol.Exp.Ther*. 322: 1144-1152.
711. Walther, S., Mahlberg, R., Eichmann, U., and Kunz, D. (2006). Delta-9-tetrahydrocannabinol for nighttime agitation in severe dementia. *Psychopharmacology (Berl)*. 185: 524-528.
712. Passmore, M. J. (2008). The cannabinoid receptor agonist nabilone for the treatment of dementia-related agitation. *Int.J.Geriatr.Psychiatry*. 23: 116-117.
713. Krishnan, S., Cairns, R., and Howard, R. (2009). Cannabinoids for the treatment of dementia. *Cochrane.Database.Syst.Rev*. CD007204.-
714. Klein, T. W. (2005). Cannabinoid-based drugs as anti-inflammatory therapeutics. *Nat.Rev.Immunol*. 5: 400-411.
715. Nagarkatti, P., Pandey, R., Rieder, S. A., Hegde, V. L. and others. (2009). Cannabinoids as novel anti-inflammatory drugs. *Future.Med.Chem*. 1: 1333-1349.
716. Stander, S., Schmelz, M., Metze, D., Luger, T. and others. (2005). Distribution of cannabinoid receptor 1 (CB1) and 2 (CB2) on sensory nerve fibers and adnexal structures in human skin. *J.Dermatol.Sci*. 38: 177-188.
717. Toth, B. J., Dobrosi, N., Dajnoki, A., Czifra, G. and others. (2011). Endocannabinoids modulate human epidermal keratinocyte proliferation and survival via the sequential engagement of cannabinoid receptor-1 and transient receptor potential vanilloid-1. *J.Invest Dermatol*. 131: 1095-1104.
718. Maccarrone, M., Di Rienzo M., Battista, N., Gasperi, V. and others. (2003). The endocannabinoid system in human keratinocytes. Evidence that anandamide inhibits epidermal differentiation through CB1 receptor-dependent inhibition of protein kinase C, activation protein-1, and transglutaminase. *J.Biol.Chem*. 278: 33896-33903.
719. Wilkinson, J. D. and Williamson, E. M. (2007). Cannabinoids inhibit human keratinocyte proliferation through a non-CB1/CB2 mechanism and have a potential therapeutic value in the treatment of psoriasis. *J.Dermatol.Sci*. 45: 87-92.
720. Dvorak, M., Watkinson, A., McGlone, F., and Rukwied, R. (2003). Histamine induced responses are attenuated by a cannabinoid receptor agonist in human skin. *Inflamm.Res*. 52: 238-245.
721. Rukwied, R., Watkinson, A., McGlone, F., and Dvorak, M. (2003). Cannabinoid agonists attenuate capsaicin-induced responses in human skin. *Pain*. 102: 283-288.
722. Watson, E. S., Murphy, J. C., and Turner, C. E. (1983). Allergenic properties of naturally occurring cannabinoids. *J.Pharm.Sci*. 72: 954-955.
723. Williams, C., Thompstone, J., and Wilkinson, M. (2008). Work-related contact urticaria to Cannabis sativa. *Contact Dermatitis*. 58: 62-63.

724. Mikuriya, T. H. (1969). Marijuana in medicine: past, present and future. *Calif.Med.* 110: 34-40.
725. Kalant, H. (2001). Medicinal use of cannabis: history and current status. *Pain Res.Manag.* 6: 80-91.
726. Zuardi, A. W. (2006). History of cannabis as a medicine: a review. *Rev.Bras.Psiquiatr.* 28: 153-157.
727. Duncan, M., Thomas, A. D., Cluny, N. L., Patel, A. and others. (2008). Distribution and function of monoacylglycerol lipase in the gastrointestinal tract. *Am.J.Physiol Gastrointest.Liver Physiol.* 295: G1255-G1265.
728. Kennedy, P. J., Clarke, G., Quigley, E. M., Groeger, J. A. and others. (2012). Gut memories: towards a cognitive neurobiology of irritable bowel syndrome. *Neurosci.Biobehav.Rev.* 36: 310-340.
729. Storr, M. A., Yuce, B., Andrews, C. N., and Sharkey, K. A. (2008). The role of the endocannabinoid system in the pathophysiology and treatment of irritable bowel syndrome. *Neurogastroenterol.Motil.* 20: 857-868.
730. Yao, X., Yang, Y. S., Cui, L. H., Zhao, K. B. and others. (2012). Subtypes of irritable bowel syndrome on Rome III criteria: a multicenter study. *J.Gastroenterol.Hepatol.* 27: 760-765.
731. Camilleri, M., Carlson, P., McKinzie, S., Grudell, A. and others. (2008). Genetic variation in endocannabinoid metabolism, gastrointestinal motility, and sensation. *Am.J.Physiol Gastrointest.Liver Physiol.* 294: G13-G19.
732. Park, J. M., Choi, M. G., Cho, Y. K., Lee, I. S. and others. (2011). Cannabinoid receptor 1 gene polymorphism and irritable bowel syndrome in the Korean population: a hypothesis-generating study. *J.Clin.Gastroenterol.* 45: 45-49.
733. Camilleri, M. and Katzka, D. A. (2012). Irritable bowel syndrome: methods, mechanisms, and pathophysiology. Genetic epidemiology and pharmacogenetics in irritable bowel syndrome. *Am.J.Physiol Gastrointest.Liver Physiol.* 302: G1075-G1084.
734. Sanson, M., Bueno, L., and Fioramonti, J. (2006). Involvement of cannabinoid receptors in inflammatory hypersensitivity to colonic distension in rats. *Neurogastroenterol.Motil.* 18: 949-956.
735. Brusberg, M., Arvidsson, S., Kang, D., Larsson, H. and others. (2009). CB1 receptors mediate the analgesic effects of cannabinoids on colorectal distension-induced visceral pain in rodents. *J.Neurosci.* 29: 1554-1564.
736. Kimball, E. S., Wallace, N. H., Schneider, C. R., D'Andrea, M. R. and others. (2010). Small intestinal cannabinoid receptor changes following a single colonic insult with oil of mustard in mice. *Front Pharmacol.* 1: 132-
737. Esfandyari, T., Camilleri, M., Ferber, I., Burton, D. and others. (2006). Effect of a cannabinoid agonist on gastrointestinal transit and postprandial satiation in healthy human subjects: a randomized, placebo-controlled study. *Neurogastroenterol.Motil.* 18: 831-838.
738. Esfandyari, T., Camilleri, M., Busciglio, J., Burton, D. and others. (2007). Effects of a cannabinoid receptor agonist on colonic motor and sensory functions in humans: a randomized, placebo-controlled study. *Am.J.Physiol Gastrointest.Liver Physiol.* 293: G137-G145.
739. Rao, S. S. and Singh, S. (2010). Clinical utility of colonic and anorectal manometry in chronic constipation. *J.Clin.Gastroenterol.* 44: 597-609.
740. Coulie, B., Camilleri, M., Bharucha, A. E., Sandborn, W. J. and others. (2001). Colonic motility in chronic ulcerative proctosigmoiditis and the effects of nicotine on colonic motility in patients and healthy subjects. *Aliment.Pharmacol.Ther.* 15: 653-663.

741. Lembo, T., Wright, R. A., Bagby, B., Decker, C. and others. (2001). Alosetron controls bowel urgency and provides global symptom improvement in women with diarrhea-predominant irritable bowel syndrome. *Am.J.Gastroenterol.* 96: 2662-2670.
742. Wong, B. S., Camilleri, M., Busciglio, I., Carlson, P. and others. (2011). Pharmacogenetic trial of a cannabinoid agonist shows reduced fasting colonic motility in patients with nonconstipated irritable bowel syndrome. *Gastroenterology.* 141: 1638-1647.
743. Wong, B. S., Camilleri, M., Eckert, D., Carlson, P. and others. (2012). Randomized pharmacodynamic and pharmacogenetic trial of dronabinol effects on colon transit in irritable bowel syndrome-diarrhea. *Neurogastroenterol.Motil.* 24: 358-e169.
744. Di Sabatino, A., Battista, N., Biancheri, P., Rapino, C. and others. (2011). The endogenous cannabinoid system in the gut of patients with inflammatory bowel disease. *Mucosal.Immunol.* 4: 574-583.
745. Mowat, C., Cole, A., Windsor, A., Ahmad, T. and others. (2011). Guidelines for the management of inflammatory bowel disease in adults. *Gut.* 60: 571-607.
746. Ligresti, A., Bisogno, T., Matias, I., De, Petrocellis L. and others. (2003). Possible endocannabinoid control of colorectal cancer growth. *Gastroenterology.* 125: 677-687.
747. Guagnini, F., Valenti, M., Mukenge, S., Matias, I. and others. (2006). Neural contractions in colonic strips from patients with diverticular disease: role of endocannabinoids and substance P. *Gut.* 55: 946-953.
748. D'Argenio, G., Petrosino, S., Gianfrani, C., Valenti, M. and others. (2007). Overactivity of the intestinal endocannabinoid system in celiac disease and in methotrexate-treated rats. *J.Mol.Med.* 85: 523-530.
749. Massa, F., Marsicano, G., Hermann, H., Cannich, A. and others. (2004). The endogenous cannabinoid system protects against colonic inflammation. *J.Clin.Invest.* 113: 1202-1209.
750. D'Argenio, G., Valenti, M., Scaglione, G., Cosenza, V. and others. (2006). Up-regulation of anandamide levels as an endogenous mechanism and a pharmacological strategy to limit colon inflammation. *FASEB J.* 20: 568-570.
751. Kimball, E. S., Schneider, C. R., Wallace, N. H., and Hornby, P. J. (2006). Agonists of cannabinoid receptor 1 and 2 inhibit experimental colitis induced by oil of mustard and by dextran sulfate sodium. *Am.J.Physiol Gastrointest.Liver Physiol.* 291: G364-G371.
752. Storr, M., Emmerdinger, D., Diegelmann, J., Yuce, B. and others. (2009). The role of fatty acid hydrolase gene variants in inflammatory bowel disease. *Aliment.Pharmacol.Ther.* 29: 542-551.
753. Engel, M. A., Kellermann, C. A., Burnat, G., Hahn, E. G. and others. (2010). Mice lacking cannabinoid CB1-, CB2-receptors or both receptors show increased susceptibility to trinitrobenzene sulfonic acid (TNBS)-induced colitis. *J.Physiol Pharmacol.* 61: 89-97.
754. Jamontt, J. M., Molleman, A., Pertwee, R. G., and Parsons, M. E. (2010). The effects of Delta-tetrahydrocannabinol and cannabidiol alone and in combination on damage, inflammation and in vitro motility disturbances in rat colitis. *Br.J.Pharmacol.* 160: 712-723.
755. Di Paola, R., Mazzon, E., Patel, N. S., Genovese, T. and others. (2005). Beneficial effects of GW274150 treatment on the development of experimental colitis induced by dinitrobenzene sulfonic acid. *Eur.J.Pharmacol.* 507: 281-289.
756. Storr, M. A., Keenan, C. M., Zhang, H., Patel, K. D. and others. (2009). Activation of the cannabinoid 2 receptor (CB2) protects against experimental colitis. *Inflamm.Bowel.Dis.* 15: 1678-1685.

757. Borrelli, F., Aviello, G., Romano, B., Orlando, P. and others. (2009). Cannabidiol, a safe and non-psychotropic ingredient of the marijuana plant *Cannabis sativa*, is protective in a murine model of colitis. *J.Mol.Med.(Berl)*. 87: 1111-1121.
758. Schicho, R. and Storr, M. (2012). Topical and systemic cannabidiol improves trinitrobenzene sulfonic acid colitis in mice. *Pharmacology*. 89: 149-155.
759. Alhouayek, M., Lambert, D. M., Delzenne, N. M., Cani, P. D. and others. (2011). Increasing endogenous 2-arachidonoylglycerol levels counteracts colitis and related systemic inflammation. *FASEB J*. 25: 2711-2721.
760. Singh, U. P., Singh, N. P., Singh, B., Price, R. L. and others. (2012). Cannabinoid receptor-2 (CB2) agonist ameliorates colitis in IL-10(-/-) mice by attenuating the activation of T cells and promoting their apoptosis. *Toxicol.Appl.Pharmacol*. 258: 256-267.
761. Izzo, A. A., Capasso, R., Aviello, G., Borrelli, F. and others. (2012). Inhibitory effect of cannabichromene, a major non-psychotropic cannabinoid extracted from *Cannabis sativa*, on inflammation-induced hypermotility in mice. *Br.J.Pharmacol*. 166: 1444-1460.
762. Klooker, T. K., Leliefeld, K. E., van den Wijngaard, R. M., and Boeckxstaens, G. E. (2011). The cannabinoid receptor agonist delta-9-tetrahydrocannabinol does not affect visceral sensitivity to rectal distension in healthy volunteers and IBS patients. *Neurogastroenterol.Motil*. 23: 30-5, e2.
763. Tam, J., Liu, J., Mukhopadhyay, B., Cinar, R. and others. (2011). Endocannabinoids in liver disease. *Hepatology*. 53: 346-355.
764. Teixeira-Clerc, F., Belot, M. P., Manin, S., Deveaux, V. and others. (2010). Beneficial paracrine effects of cannabinoid receptor 2 on liver injury and regeneration. *Hepatology*. 52: 1046-1059.
765. Giannone, F. A., Baldassarre, M., Domenicali, M., Zaccherini, G. and others. (2012). Reversal of liver fibrosis by the antagonism of endocannabinoid CB1 receptor in a rat model of CCl(4)-induced advanced cirrhosis. *Lab Invest*. 92: 384-395.
766. Lim, M. P., Devi, L. A., and Rozenfeld, R. (2011). Cannabidiol causes activated hepatic stellate cell death through a mechanism of endoplasmic reticulum stress-induced apoptosis. *Cell Death.Dis*. 2: e170-
767. Mukhopadhyay, P., Rajesh, M., Horvath, B., Batkai, S. and others. (2011). Cannabidiol protects against hepatic ischemia/reperfusion injury by attenuating inflammatory signaling and response, oxidative/nitrative stress, and cell death. *Free Radic.Biol.Med*. 50: 1368-1381.
768. Fouad, A. A. and Jresat, I. (2011). Therapeutic potential of cannabidiol against ischemia/reperfusion liver injury in rats. *Eur.J.Pharmacol*. 670: 216-223.
769. Avraham, Y., Grigoriadis, N., Poutahidis, T., Vorobiev, L. and others. (2011). Cannabidiol improves brain and liver function in a fulminant hepatic failure-induced model of hepatic encephalopathy in mice. *Br.J.Pharmacol*. 162: 1650-1658.
770. Batkai, S., Mukhopadhyay, P., Horvath, B., Rajesh, M. and others. (2012). Delta8-Tetrahydrocannabivarin prevents hepatic ischaemia/reperfusion injury by decreasing oxidative stress and inflammatory responses through cannabinoid CB2 receptors. *Br.J.Pharmacol*. 165: 2450-2461.
771. Silvestri, C., Ligresti, A., and Di, Marzo, V. (2011). Peripheral effects of the endocannabinoid system in energy homeostasis: adipose tissue, liver and skeletal muscle. *Rev.Endocr.Metab Disord*. 12: 153-162.
772. O'Hare, J. D., Zielinski, E., Cheng, B., Scherer, T. and others. (2011). Central endocannabinoid signaling regulates hepatic glucose production and systemic lipolysis. *Diabetes*. 60: 1055-1062.

773. Engeli, S. (2012). Central and peripheral cannabinoid receptors as therapeutic targets in the control of food intake and body weight. *Handb.Exp.Pharmacol.* 357-381.
774. Li, C., Jones, P. M., and Persaud, S. J. (2011). Role of the endocannabinoid system in food intake, energy homeostasis and regulation of the endocrine pancreas. *Pharmacol.Ther.* 129: 307-320.
775. Osei-Hyiaman, D., Depetrillo, M., Pacher, P., Liu, J. and others. (2005). Endocannabinoid activation at hepatic CB1 receptors stimulates fatty acid synthesis and contributes to diet-induced obesity. *J.Clin.Invest.* 115: 1298-1305.
776. Pagano, C., Pilon, C., Calcagno, A., Urbanet, R. and others. (2007). The endogenous cannabinoid system stimulates glucose uptake in human fat cells via phosphatidylinositol 3-kinase and calcium-dependent mechanisms. *J.Clin.Endocrinol.Metab.* 92: 4810-4819.
777. Cardinal, P., Bellocchio, L., Clark, S., Cannich, A. and others. (2012). Hypothalamic CB1 cannabinoid receptors regulate energy balance in mice. *Endocrinology.* 153: 4136-4143.
778. Cota, D., Marsicano, G., Tschop, M., Grubler, Y. and others. (2003). The endogenous cannabinoid system affects energy balance via central orexigenic drive and peripheral lipogenesis. *J.Clin.Invest.* 112: 423-431.
779. Quarta, C., Bellocchio, L., Mancini, G., Mazza, R. and others. (2010). CB(1) signaling in forebrain and sympathetic neurons is a key determinant of endocannabinoid actions on energy balance. *Cell Metab.* 11: 273-285.
780. Osei-Hyiaman, D., Liu, J., Zhou, L., Godlewski, G. and others. (2008). Hepatic CB1 receptor is required for development of diet-induced steatosis, dyslipidemia, and insulin and leptin resistance in mice. *J.Clin.Invest.* 118: 3160-3169.
781. Liu, J., Zhou, L., Xiong, K., Godlewski, G. and others. (2012). Hepatic cannabinoid receptor-1 mediates diet-induced insulin resistance via inhibition of insulin signaling and clearance in mice. *Gastroenterology.* 142: 1218-1228.
782. Ravinet, Trillou C., Arnone, M., Delgorge, C., Gonalons, N. and others. (2003). Anti-obesity effect of SR141716, a CB1 receptor antagonist, in diet-induced obese mice. *Am.J.Physiol Regul.Integr.Comp Physiol.* 284: R345-R353.
783. Poirier, B., Bidouard, J. P., Cadrouvele, C., Marniquet, X. and others. (2005). The anti-obesity effect of rimonabant is associated with an improved serum lipid profile. *Diabetes Obes.Metab.* 7: 65-72.
784. Jbilo, O., Ravinet-Trillou, C., Arnone, M., Buisson, I. and others. (2005). The CB1 receptor antagonist rimonabant reverses the diet-induced obesity phenotype through the regulation of lipolysis and energy balance. *FASEB J.* 19: 1567-1569.
785. Watanabe, T., Kubota, N., Ohsugi, M., Kubota, T. and others. (2009). Rimonabant ameliorates insulin resistance via both adiponectin-dependent and adiponectin-independent pathways. *J.Biol.Chem.* 284: 1803-1812.
786. Jourdan, T., Djaouti, L., Demizieux, L., Gresti, J. and others. (2010). CB1 antagonism exerts specific molecular effects on visceral and subcutaneous fat and reverses liver steatosis in diet-induced obese mice. *Diabetes.* 59: 926-934.
787. Crespillo, A., Suarez, J., Bermudez-Silva, F. J., Rivera, P. and others. (2011). Expression of the cannabinoid system in muscle: effects of a high-fat diet and CB1 receptor blockade. *Biochem.J.* 433: 175-185.
788. Bell-Anderson, K. S., Aouad, L., Williams, H., Sanz, F. R. and others. (2011). Coordinated improvement in glucose tolerance, liver steatosis and obesity-associated inflammation by cannabinoid I receptor antagonism in fat Aussie mice. *Int.J.Obes.(Lond).* 35: 1539-1548.

789. Van Gaal, L. F., Rissanen, A. M., Scheen, A. J., Ziegler, O. and others. (2005). Effects of the cannabinoid-1 receptor blocker rimonabant on weight reduction and cardiovascular risk factors in overweight patients: 1-year experience from the RIO-Europe study. *Lancet*. 365: 1389-1397.
790. Despres, J. P., Golay, A., and Sjostrom, L. (2005). Effects of rimonabant on metabolic risk factors in overweight patients with dyslipidemia. *N.Engl.J.Med.* 353: 2121-2134.
791. Pi-Sunyer, F. X., Aronne, L. J., Heshmati, H. M., Devin, J. and others. (2006). Effect of rimonabant, a cannabinoid-1 receptor blocker, on weight and cardiometabolic risk factors in overweight or obese patients: RIO-North America: a randomized controlled trial. *JAMA*. 295: 761-775.
792. Scheen, A. J., Finer, N., Hollander, P., Jensen, M. D. and others. (2006). Efficacy and tolerability of rimonabant in overweight or obese patients with type 2 diabetes: a randomised controlled study. *Lancet*. 368: 1660-1672.
793. Van Gaal, L. F., Scheen, A. J., Rissanen, A. M., Rossner, S. and others. (2008). Long-term effect of CB1 blockade with rimonabant on cardiometabolic risk factors: two year results from the RIO-Europe Study. *Eur.Heart J.* 29: 1761-1771.
794. Van Gaal, L., Pi-Sunyer, X., Despres, J. P., McCarthy, C. and others. (2008). Efficacy and safety of rimonabant for improvement of multiple cardiometabolic risk factors in overweight/obese patients: pooled 1-year data from the Rimonabant in Obesity (RIO) program. *Diabetes Care*. 31 Suppl 2: S229-S240.
795. Despres, J. P., Ross, R., Boka, G., Almeras, N. and others. (2009). Effect of rimonabant on the high-triglyceride/low-HDL-cholesterol dyslipidemia, intraabdominal adiposity, and liver fat: the ADAGIO-Lipids trial. *Arterioscler.Thromb.Vasc.Biol.* 29: 416-423.
796. Hayatbakhsh, M. R., O'Callaghan, M. J., Mamun, A. A., Williams, G. M. and others. (2010). Cannabis use and obesity and young adults. *Am.J Drug Alcohol Abuse*. 36: 350-356.
797. Le Strat, Y. and Le Foll, B. (2011). Obesity and cannabis use: results from 2 representative national surveys. *Am.J.Epidemiol.* 174: 929-933.
798. Deveaux, V., Cadoudal, T., Ichigotani, Y., Teixeira-Clerc, F. and others. (2009). Cannabinoid CB2 receptor potentiates obesity-associated inflammation, insulin resistance and hepatic steatosis. *PLoS.One.* 4: e5844-
799. Agudo, J., Martin, M., Roca, C., Molas, M. and others. (2010). Deficiency of CB2 cannabinoid receptor in mice improves insulin sensitivity but increases food intake and obesity with age. *Diabetologia*. 53: 2629-2640.
800. Levendal, R. A., Schumann, D., Donath, M., and Frost, C. L. (2012). Cannabis exposure associated with weight reduction and beta-cell protection in an obese rat model. *Phytomedicine*. 19: 575-582.
801. Li, C., Bowe, J. E., Huang, G. C., Amiel, S. A. and others. (2011). Cannabinoid receptor agonists and antagonists stimulate insulin secretion from isolated human islets of Langerhans. *Diabetes Obes.Metab.* 13: 903-910.
802. Li, C., Vilches-Flores, A., Zhao, M., Amiel, S. A. and others. (2012). Expression and function of monoacylglycerol lipase in mouse beta-cells and human islets of Langerhans. *Cell Physiol Biochem*. 30: 347-358.
803. Bermudez-Silva, F. J., Suarez, J., Baixeras, E., Cobo, N. and others. (2008). Presence of functional cannabinoid receptors in human endocrine pancreas. *Diabetologia*. 51: 476-487.
804. De Pasquale, A., Costa, G., and Trovato, A. (1978). The influence of cannabis on glucoregulation. *Bull.Narc.* 30: 33-41.
805. Bermudez-Siva, F. J., Serrano, A., Diaz-Molina, F. J., Sanchez, Vera, I and others. (2006). Activation of cannabinoid CB1 receptors induces glucose intolerance in rats. *Eur.J.Pharmacol.* 531: 282-284.

806. Hollister, L. E. and Reaven, G. M. (1974). Delta-9-tetrahydrocannabinol and glucose tolerance. *Clin.Pharmacol.Ther.* 16: 297-302.
807. Rajavashisth, T. B., Shaheen, M., Norris, K. C., Pan, D. and others. (2012). Decreased prevalence of diabetes in marijuana users: cross-sectional data from the National Health and Nutrition Examination Survey (NHANES) III. *BMJ Open.* 2: e000494-
808. Michalski, C. W., Laukert, T., Sauliunaite, D., Pacher, P. and others. (2007). Cannabinoids ameliorate pain and reduce disease pathology in cerulein-induced acute pancreatitis. *Gastroenterology.* 132: 1968-1978.
809. Michalski, C. W., Maier, M., Erkan, M., Sauliunaite, D. and others. (2008). Cannabinoids reduce markers of inflammation and fibrosis in pancreatic stellate cells. *PLoS.One.* 3: e1701-
810. Matsuda, K., Mikami, Y., Takeda, K., Fukuyama, S. and others. (2005). The cannabinoid 1 receptor antagonist, AM251, prolongs the survival of rats with severe acute pancreatitis. *Tohoku J.Exp.Med.* 207: 99-107.
811. Dembinski, A., Warzecha, Z., Ceranowicz, P., Dembinski, M. and others. (2006). Cannabinoids in acute gastric damage and pancreatitis. *J.Physiol Pharmacol.* 57 Suppl 5: 137-154.
812. Zyromski, N. J., Mathur, A., Pitt, H. A., Wade, T. E. and others. (2009). Cannabinoid receptor-1 blockade attenuates acute pancreatitis in obesity by an adiponectin mediated mechanism. *J.Gastrointest.Surg.* 13: 831-838.
813. Petrella, C., Agostini, S., Alema, G. S., Casolini, P. and others. (2010). Cannabinoid agonist WIN55,212 in vitro inhibits interleukin-6 (IL-6) and monocyte chemo-attractant protein-1 (MCP-1) release by rat pancreatic acini and in vivo induces dual effects on the course of acute pancreatitis. *Neurogastroenterol.Motil.* 22: 1248-56, e323.
814. Malfitano, A. M., Ciaglia, E., Gangemi, G., Gazzerro, P. and others. (2011). Update on the endocannabinoid system as an anticancer target. *Expert.Opin.Ther.Targets.* 15: 297-308.
815. Pagotto, U., Marsicano, G., Fezza, F., Theodoropoulou, M. and others. (2001). Normal human pituitary gland and pituitary adenomas express cannabinoid receptor type 1 and synthesize endogenous cannabinoids: first evidence for a direct role of cannabinoids on hormone modulation at the human pituitary level. *J.Clin.Endocrinol.Metab.* 86: 2687-2696.
816. Schmid, P. C., Wold, L. E., Krebsbach, R. J., Berdyshev, E. V. and others. (2002). Anandamide and other N-acyl ethanolamines in human tumors. *Lipids.* 37: 907-912.
817. Nithipatikom, K., Endsley, M. P., Isbell, M. A., Falck, J. R. and others. (2004). 2-arachidonoylglycerol: a novel inhibitor of androgen-independent prostate cancer cell invasion. *Cancer Res.* 64: 8826-8830.
818. Petersen, G., Moesgaard, B., Schmid, P. C., Schmid, H. H. and others. (2005). Endocannabinoid metabolism in human glioblastomas and meningiomas compared to human non-tumour brain tissue. *J.Neurochem.* 93: 299-309.
819. Cianchi, F., Papucci, L., Schiavone, N., Lulli, M. and others. (2008). Cannabinoid receptor activation induces apoptosis through tumor necrosis factor alpha-mediated ceramide de novo synthesis in colon cancer cells. *Clin.Cancer Res.* 14: 7691-7700.
820. Grimaldi, C. and Capasso, A. (2011). The endocannabinoid system in the cancer therapy: an overview. *Curr.Med.Chem.* 18: 1575-1583.
821. Pisanti, S. and Bifulco, M. (2009). Endocannabinoid system modulation in cancer biology and therapy. *Pharmacol.Res.* 60: 107-116.

822. Stella, N. (2010). Cannabinoid and cannabinoid-like receptors in microglia, astrocytes, and astrocytomas. *Glia*. 58: 1017-1030.
823. McAllister, S. D., Chan, C., Taft, R. J., Luu, T. and others. (2005). Cannabinoids selectively inhibit proliferation and induce death of cultured human glioblastoma multiforme cells. *J.Neurooncol.* 74: 31-40.
824. Cudaback, E., Marrs, W., Moeller, T., and Stella, N. (2010). The expression level of CB1 and CB2 receptors determines their efficacy at inducing apoptosis in astrocytomas. *PLoS.One.* 5: e8702.-
825. Caffarel, M. M., Sarrio, D., Palacios, J., Guzman, M. and others. (2006). Delta9-tetrahydrocannabinol inhibits cell cycle progression in human breast cancer cells through Cdc2 regulation. *Cancer Res.* 66: 6615-6621.
826. McKallip, R. J., Nagarkatti, M., and Nagarkatti, P. S. (2005). Delta-9-tetrahydrocannabinol enhances breast cancer growth and metastasis by suppression of the antitumor immune response. *J.Immunol.* 174: 3281-3289.
827. Takeda, S., Yamaori, S., Motoya, E., Matsunaga, T. and others. (2008). Delta(9)-Tetrahydrocannabinol enhances MCF-7 cell proliferation via cannabinoid receptor-independent signaling. *Toxicology.* 245: 141-146.
828. Ligresti, A., Moriello, A. S., Starowicz, K., Matias, I. and others. (2006). Antitumor activity of plant cannabinoids with emphasis on the effect of cannabidiol on human breast carcinoma. *J.Pharmacol.Exp.Ther.* 318: 1375-1387.
829. Preet, A., Ganju, R. K., and Groopman, J. E. (2008). Delta9-Tetrahydrocannabinol inhibits epithelial growth factor-induced lung cancer cell migration in vitro as well as its growth and metastasis in vivo. *Oncogene.* 27: 339-346.
830. Greenhough, A., Patsos, H. A., Williams, A. C., and Paraskeva, C. (2007). The cannabinoid delta(9)-tetrahydrocannabinol inhibits RAS-MAPK and PI3K-AKT survival signalling and induces BAD-mediated apoptosis in colorectal cancer cells. *Int.J.Cancer.* 121: 2172-2180.
831. Galve-Roperh, I., Sánchez, C., Cortes, M. L., Gomez, del Pulgar and others. (2000). Anti-tumoral action of cannabinoids: involvement of sustained ceramide accumulation and extracellular signal-regulated kinase activation. *Nat.Med.* 6: 313-319.
832. Caffarel, M. M., Andradas, C., Mira, E., Perez-Gomez, E. and others. (2010). Cannabinoids reduce ErbB2-driven breast cancer progression through Akt inhibition. *Mol.Cancer.* 9: 196-206.
833. McAllister, S. D., Murase, R., Christian, R. T., Lau, D. and others. (2010). Pathways mediating the effects of cannabidiol on the reduction of breast cancer cell proliferation, invasion, and metastasis. *Breast Cancer Res.Treat.* 129: 37-47.
834. Carracedo, A., Lorente, M., Egia, A., Blazquez, C. and others. (2006). The stress-regulated protein p8 mediates cannabinoid-induced apoptosis of tumor cells. *Cancer Cell.* 9: 301-312.
835. Torres, S., Lorente, M., Rodriguez-Fornes, F., Hernandez-Tiedra, S. and others. (2011). A combined preclinical therapy of cannabinoids and temozolomide against glioma. *Mol.Cancer Ther.* 10: 90-103.
836. Guzman, M., Duarte, M. J., Blazquez, C., Ravina, J. and others. (2006). A pilot clinical study of Delta9-tetrahydrocannabinol in patients with recurrent glioblastoma multiforme. *Br.J.Cancer.* 95: 197-203.
837. Dinnes, J., Cave, C., Huang, S., and Milne, R. (2002). A rapid and systematic review of the effectiveness of temozolomide for the treatment of recurrent malignant glioma. *Br.J.Cancer.* 86: 501-505.
838. Brem, H., Piantadosi, S., Burger, P. C., Walker, M. and others. (1995). Placebo-controlled trial of safety and efficacy of intraoperative controlled delivery by biodegradable polymers of chemotherapy for recurrent gliomas. The Polymer-brain Tumor Treatment Group. *Lancet.* 345: 1008-1012.

839. Marcu, J. P., Christian, R. T., Lau, D., Zielinski, A. J. and others. (2010). Cannabidiol enhances the inhibitory effects of delta9-tetrahydrocannabinol on human glioblastoma cell proliferation and survival. *Mol.Cancer Ther.* 9: 180-189.
840. Velasco, G., Carracedo, A., Blazquez, C., Lorente, M. and others. (2007). Cannabinoids and gliomas. *Mol.Neurobiol.* 36: 60-67.
841. Parolaro, D. and Massi, P. (2008). Cannabinoids as potential new therapy for the treatment of gliomas. *Expert.Rev.Neurother.* 8: 37-49.
842. Alexander, A., Smith, P. F., and Rosengren, R. J. (2009). Cannabinoids in the treatment of cancer. *Cancer Lett.* 285: 6-12.
843. Steffens, S., Veillard, N. R., Arnaud, C., Pelli, G. and others. (2005). Low dose oral cannabinoid therapy reduces progression of atherosclerosis in mice. *Nature.* 434: 782-786.
844. Sugamura, K., Sugiyama, S., Nozaki, T., Matsuzawa, Y. and others. (2009). Activated endocannabinoid system in coronary artery disease and antiinflammatory effects of cannabinoid 1 receptor blockade on macrophages. *Circulation.* 119: 28-36.
845. Zhao, Y., Yuan, Z., Liu, Y., Xue, J. and others. (2010). Activation of cannabinoid CB2 receptor ameliorates atherosclerosis associated with suppression of adhesion molecules. *J.Cardiovasc.Pharmacol.* 55: 292-298.
846. Hoyer, F. F., Steinmetz, M., Zimmer, S., Becker, A. and others. (2011). Atheroprotection via cannabinoid receptor-2 is mediated by circulating and vascular cells in vivo. *J.Mol.Cell Cardiol.* 51: 1007-1014.
847. Willecke, F., Zeschky, K., Ortiz, Rodriguez A., Colberg, C. and others. (2011). Cannabinoid receptor 2 signaling does not modulate atherogenesis in mice. *PLoS.One.* 6: e19405-
848. Netherland, C. D., Pickle, T. G., Bales, A., and Thewke, D. P. (2010). Cannabinoid receptor type 2 (CB2) deficiency alters atherosclerotic lesion formation in hyperlipidemic Ldlr-null mice. *Atherosclerosis.* 213: 102-108.
849. Rajesh, M., Mukhopadhyay, P., Hasko, G., Liaudet, L. and others. (2010). Cannabinoid-1 receptor activation induces reactive oxygen species-dependent and -independent mitogen-activated protein kinase activation and cell death in human coronary artery endothelial cells. *Br.J.Pharmacol.* 160: 688-700.
850. Singla, S., Sachdeva, R., and Mehta, J. L. (2012). Cannabinoids and atherosclerotic coronary heart disease. *Clin.Cardiol.* 35: 329-335.
851. Montecucco, F. and Di Marzo, V. (2012). At the heart of the matter: the endocannabinoid system in cardiovascular function and dysfunction. *Trends Pharmacol.Sci.* 33: 331-340.
852. Steffens, S. and Pacher, P. (2012). Targeting cannabinoid receptor CB(2) in cardiovascular disorders: promises and controversies. *Br.J.Pharmacol.* 167: 313-323.
853. Takeda, S., Usami, N., Yamamoto, I., and Watanabe, K. (2009). Cannabidiol-2',6'-dimethyl ether, a cannabidiol derivative, is a highly potent and selective 15-lipoxygenase inhibitor. *Drug Metab Dispos.* 37: 1733-1737.
854. Hezode, C., Zafrani, E. S., Roudot-Thoraval, F., Costentin, C. and others. (2008). Daily cannabis use: a novel risk factor of steatosis severity in patients with chronic hepatitis C. *Gastroenterology.* 134: 432-439.
855. Barrett, S. P., Darredeau, C., and Pihl, R. O. (2006). Patterns of simultaneous polysubstance use in drug using university students. *Hum.Psychopharmacol.* 21: 255-263.

856. Agrawal, A. and Lynskey, M. T. (2007). Does gender contribute to heterogeneity in criteria for cannabis abuse and dependence? Results from the national epidemiological survey on alcohol and related conditions. *Drug Alcohol Depend.* 88: 300-307.
857. Kuepper, R., van Os J., Lieb, R., Wittchen, H. U. and others. (2011). Continued cannabis use and risk of incidence and persistence of psychotic symptoms: 10 year follow-up cohort study. *BMJ.* 342: d738.-
858. Budney, A. J. and Hughes, J. R. (2006). The cannabis withdrawal syndrome. *Curr.Opin.Psychiatry.* 19: 233-238.
859. National Institute on Drug Abuse. NIDA InfoFacts: Marijuana. 2009.
860. Mathew, R. J., Wilson, W. H., Coleman, R. E., Turkington, T. G. and others. (1997). Marijuana intoxication and brain activation in marijuana smokers. *Life Sci.* 60: 2075-2089.
861. Ramaekers, J. G., Kauert, G., van, Ruitenbeek P., Theunissen, E. L. and others. (2006). High-potency marijuana impairs executive function and inhibitory motor control. *Neuropsychopharmacology.* 31: 2296-2303.
862. Kurzthaler, I., Hummer, M., Miller, C., Sperner-Unterweger, B. and others. (1999). Effect of cannabis use on cognitive functions and driving ability. *J.Clin.Psychiatry.* 60: 395-399.
863. Reisfield, G. M., Wasan, A. D., and Jamison, R. N. (2009). The prevalence and significance of cannabis use in patients prescribed chronic opioid therapy: a review of the extant literature. *Pain Med.* 10: 1434-1441.
864. Ashton, C. H. (1999). Adverse effects of cannabis and cannabinoids. *Br.J.Anaesth.* 83: 637-649.
865. de Jong, F. A., Engels, F. K., Mathijssen, R. H., van, Zuylen L. and others. (2005). Medicinal cannabis in oncology practice: still a bridge too far? *J.Clin.Oncol.* 23: 2886-2891.
866. Ballard, M. E. and de, Wit H. (2011). Combined effects of acute, very-low-dose ethanol and delta(9)-tetrahydrocannabinol in healthy human volunteers. *Pharmacol.Biochem.Behav.* 97: 627-631.
867. Battista, N., Pasquariello, N., Di, Tommaso M., and Maccarrone, M. (2008). Interplay between endocannabinoids, steroids and cytokines in the control of human reproduction. *J.Neuroendocrinol.* 20 Suppl 1: 82-89.
868. Habayeb, O. M., Taylor, A. H., Finney, M., Evans, M. D. and others. (2008). Plasma anandamide concentration and pregnancy outcome in women with threatened miscarriage. *JAMA.* 299: 1135-1136.
869. Fried, P. A. (2002). Conceptual issues in behavioral teratology and their application in determining long-term sequelae of prenatal marijuana exposure. *J.Child Psychol.Psychiatry.* 43: 81-102.
870. Richardson, G. A., Ryan, C., Willford, J., Day, N. L. and others. (2002). Prenatal alcohol and marijuana exposure: effects on neuropsychological outcomes at 10 years. *Neurotoxicol.Teratol.* 24: 309-320.
871. Perez-Reyes, M. and Wall, M. E. (1982). Presence of delta9-tetrahydrocannabinol in human milk. *N.Engl.J.Med.* 307: 819-820.
872. Garry, A., Rigourd, V., Amirouche, A., Fauroux, V. and others. (2009). Cannabis and breastfeeding. *J.Toxicol.* 2009: 596149.-
873. Chait, L. D. and Perry, J. L. (1994). Acute and residual effects of alcohol and marijuana, alone and in combination, on mood and performance. *Psychopharmacology (Berl).* 115: 340-349.
874. Lukas, S. E. and Orozco, S. (2001). Ethanol increases plasma Delta(9)-tetrahydrocannabinol (THC) levels and subjective effects after marijuana smoking in human volunteers. *Drug Alcohol Depend.* 64: 143-149.

875. Li, X. Q., Andersson, T. B., Ahlstrom, M., and Weidolf, L. (2004). Comparison of inhibitory effects of the proton pump-inhibiting drugs omeprazole, esomeprazole, lansoprazole, pantoprazole, and rabeprazole on human cytochrome P450 activities. *Drug Metab Dispos.* 32: 821-827.
876. Spina, E., Santoro, V., and D'Arrigo, C. (2008). Clinically relevant pharmacokinetic drug interactions with second-generation antidepressants: an update. *Clin. Ther.* 30: 1206-1227.
877. Davison, S. N. and Davison, J. S. (2011). Is there a legitimate role for the therapeutic use of cannabinoids for symptom management in chronic kidney disease? *J. Pain Symptom. Manage.* 41: 768-778.
878. Ellis, G. M., Jr., Mann, M. A., Judson, B. A., Schramm, N. T. and others. (1985). Excretion patterns of cannabinoid metabolites after last use in a group of chronic users. *Clin. Pharmacol. Ther.* 38: 572-578.
879. Lowe, R. H., Abraham, T. T., Darwin, W. D., Herning, R. and others. (2009). Extended urinary Delta9-tetrahydrocannabinol excretion in chronic cannabis users precludes use as a biomarker of new drug exposure. *Drug Alcohol Depend.* 105: 24-32.
880. Wang, T., Collet, J. P., Shapiro, S., and Ware, M. A. (2008). Adverse effects of medical cannabinoids: a systematic review. *CMAJ.* 178: 1669-1678.
881. Amos, A., Wiltshire, S., Bostock, Y., Haw, S. and others. (2004). 'You can't go without a fag...you need it for your hash'--a qualitative exploration of smoking, cannabis and young people. *Addiction.* 99: 77-81.
882. Hezode, C., Roudot-Thoraval, F., Nguyen, S., Grenard, P. and others. (2005). Daily cannabis smoking as a risk factor for progression of fibrosis in chronic hepatitis C. *Hepatology.* 42: 63-71.
883. Vandrey, R. G., Budney, A. J., Hughes, J. R., and Liguori, A. (2008). A within-subject comparison of withdrawal symptoms during abstinence from cannabis, tobacco, and both substances. *Drug Alcohol Depend.* 92: 48-54.
884. Agrawal, A., Scherrer, J. F., Lynskey, M. T., Sartor, C. E. and others. (2011). Patterns of use, sequence of onsets and correlates of tobacco and cannabis. *Addict. Behav.* 36: 1141-1147.
885. Wu, T. C., Tashkin, D. P., Djahed, B., and Rose, J. E. (1988). Pulmonary hazards of smoking marijuana as compared with tobacco. *N. Engl. J. Med.* 318: 347-351.
886. Taylor, D. R., Fergusson, D. M., Milne, B. J., Horwood, L. J. and others. (2002). A longitudinal study of the effects of tobacco and cannabis exposure on lung function in young adults. *Addiction.* 97: 1055-1061.
887. Tashkin, D. P. Marijuana and the lung. *Marijuana and medicine.* Nahas, C. G., Sutin, K. M., Harvey, D. J., and Agurell, S. Totowa, New Jersey: Humana Press, 1999.
888. Taylor, D. R., Poulton, R., Moffitt, T. E., Ramankutty, P. and others. (2000). The respiratory effects of cannabis dependence in young adults. *Addiction.* 95: 1669-1677.
889. Denning, D. W., Follansbee, S. E., Scolaro, M., Norris, S. and others. (1991). Pulmonary aspergillosis in the acquired immunodeficiency syndrome. *N. Engl. J. Med.* 324: 654-662.
890. Moore, B. A., Augustson, E. M., Moser, R. P., and Budney, A. J. (2005). Respiratory effects of marijuana and tobacco use in a U.S. sample. *J. Gen. Intern. Med.* 20: 33-37.
891. Roth, M. D., Whittaker, K., Salehi, K., Tashkin, D. P. and others. (2004). Mechanisms for impaired effector function in alveolar macrophages from marijuana and cocaine smokers. *J. Neuroimmunol.* 147: 82-86.

892. LIU, D. Z., HU, C. M., HUANG, C. H., WEY, S. P. and others. (2010). Cannabidiol attenuates delayed-type hypersensitivity reactions via suppressing T-cell and macrophage reactivity. *Acta Pharmacol.Sin.* 31: 1611-1617.
893. Kozela, E., Pietr, M., Juknat, A., Rimmerman, N. and others. (2010). Cannabinoids Delta(9)-tetrahydrocannabinol and cannabidiol differentially inhibit the lipopolysaccharide-activated NF-kappaB and interferon-beta/STAT proinflammatory pathways in BV-2 microglial cells. *J.Biol.Chem.* 285: 1616-1626.
894. Reiss, C. S. (2010). Cannabinoids and viral infections. *Pharmaceuticals (Basel)*. 3: 1873-1886.
895. Mishkin, E. M. and Cabral, G. A. (1985). delta-9-Tetrahydrocannabinol decreases host resistance to herpes simplex virus type 2 vaginal infection in the B6C3F1 mouse. *J.Gen.Virol.* 66 (Pt 12): 2539-2549.
896. Cabral, G. A., McNerney, P. J., and Mishkin, E. M. (1986). Delta-9-tetrahydrocannabinol enhances release of herpes simplex virus type 2. *J.Gen.Virol.* 67 (Pt 9): 2017-2022.
897. Roth, M. D., Baldwin, G. C., and Tashkin, D. P. (2002). Effects of delta-9-tetrahydrocannabinol on human immune function and host defense. *Chem.Phys.Lipids.* 121: 229-239.
898. Buchweitz, J. P., Karmaus, P. W., Harkema, J. R., Williams, K. J. and others. (2007). Modulation of airway responses to influenza A/PR/8/34 by Delta9-tetrahydrocannabinol in C57BL/6 mice. *J.Pharmacol.Exp.Ther.* 323: 675-683.
899. Zhang, X., Wang, J. F., Kunos, G., and Groopman, J. E. (2007). Cannabinoid modulation of Kaposi's sarcoma-associated herpesvirus infection and transformation. *Cancer Res.* 67: 7230-7237.
900. Herrera, R. A., Oved, J. H., and Reiss, C. S. (2008). Disruption of IFN-gamma-mediated antiviral activity in neurons: the role of cannabinoids. *Viral Immunol.* 21: 141-152.
901. Bredt, B. M., Higuera-Alhino, D., Shade, S. B., Hebert, S. J. and others. (2002). Short-term effects of cannabinoids on immune phenotype and function in HIV-1-infected patients. *J.Clin.Pharmacol.* 42: 82S-89S.
902. Chao, C., Jacobson, L. P., Tashkin, D., Martinez-Maza, O. and others. (2008). Recreational drug use and T lymphocyte subpopulations in HIV-uninfected and HIV-infected men. *Drug Alcohol Depend.* 94: 165-171.
903. Di Franco, M. J., Sheppard, H. W., Hunter, D. J., Tosteson, T. D. and others. (1996). The lack of association of marijuana and other recreational drugs with progression to AIDS in the San Francisco Men's Health Study. *Ann.Epidemiol.* 6: 283-289.
904. Ishida, J. H., Peters, M. G., Jin, C., Louie, K. and others. (2008). Influence of cannabis use on severity of hepatitis C disease. *Clin.Gastroenterol.Hepatol.* 6: 69-75.
905. Bonn-Miller, M. O., Oser, M. L., Bucossi, M. M., and Trafton, J. A. (2012). Cannabis use and HIV antiretroviral therapy adherence and HIV-related symptoms (in press). *J.Behav.Med.*
906. Rossato, M., Pagano, C., and Vettor, R. (2008). The cannabinoid system and male reproductive functions. *J.Neuroendocrinol.* 20 Suppl 1: 90-93.
907. Hembree, W. C., Nahas, G. G., Zeidenberg, P., and Huang, H. F. S. Changes in human spermatozoa associated with high-dose marijuana smoking. *Marihuana and medicine*. Nahas, G. G., Sutin, K. M., Harvey, D. J., and Agurell, S. Totowa: Humana Press, 1999.
908. Hong, C. Y., Chaput de Saintonge, D. M., Turner, P., and Fairbairn, J. W. (1982). Comparison of the inhibitory action of delta-9-tetrahydrocannabinol and petroleum spirit extract of herbal cannabis on human sperm motility. *Hum.Toxicol.* 1: 151-154.

909. Whan, L. B., West, M. C., McClure, N., and Lewis, S. E. (2006). Effects of delta-9-tetrahydrocannabinol, the primary psychoactive cannabinoid in marijuana, on human sperm function in vitro. *Fertil.Steril.* 85: 653-660.
910. Lacson, J. C., Carroll, J. D., Tuazon, E., Castela, E. J. and others. (2012). Population-based case-control study of recreational drug use and testis cancer risk confirms an association between marijuana use and nonseminoma risk. *Cancer.* 118: 5374-5383.
911. Zuckerman, B., Frank, D. A., Hingson, R., Amaro, H. and others. (1989). Effects of maternal marijuana and cocaine use on fetal growth. *N.Engl.J.Med.* 320: 762-768.
912. Hurd, Y. L., Wang, X., Anderson, V., Beck, O. and others. (2005). Marijuana impairs growth in mid-gestation fetuses. *Neurotoxicol.Teratol.* 27: 221-229.
913. El-Marroun, H., Tiemeier, H., Steegers, E. A. Jaddoe, V. W and others. (2009). Intrauterine Cannabis Exposure Affects Fetal Growth Trajectories: The Generation R Study. *J.Am.Acad.Child Adolesc.Psychiatry.* 48: 1173-1181.
914. Gray, T. R., Eiden, R. D., Leonard, K. E., Connors, G. J. and others. (2010). Identifying prenatal cannabis exposure and effects of concurrent tobacco exposure on neonatal growth. *Clin.Chem.* 56: 1442-1450.
915. Scragg, R. K., Mitchell, E. A., Ford, R. P., Thompson, J. M. and others. (2001). Maternal cannabis use in the sudden death syndrome. *Acta Paediatr.* 90: 57-60.
916. Shiono, P. H., Klebanoff, M. A., Nugent, R. P., Cotch, M. F. and others. (1995). The impact of cocaine and marijuana use on low birth weight and preterm birth: a multicenter study. *Am.J.Obstet.Gynecol.* 172: 19-27.
917. Fried, P. A., Watkinson, B., and Gray, R. (1999). Growth from birth to early adolescence in offspring prenatally exposed to cigarettes and marijuana. *Neurotoxicol.Teratol.* 21: 513-525.
918. van Gelder, M. M., Reefhuis, J., Caton, A. R., Werler, M. M. and others. (2010). Characteristics of pregnant illicit drug users and associations between cannabis use and perinatal outcome in a population-based study. *Drug Alcohol Depend.* 109: 243-247.
919. Ahmad, G. R. and Ahmad, N. (1990). Passive consumption of marijuana through milk: a low level chronic exposure to delta-9-tetrahydrocannabinol(THC). *J.Toxicol.Clin.Toxicol.* 28: 255-260.
920. Astley, S. J. and Little, R. E. (1990). Maternal marijuana use during lactation and infant development at one year. *Neurotoxicol.Teratol.* 12: 161-168.
921. Mittleman, M. A. and Mostofsky, E. (2011). Physical, psychological and chemical triggers of acute cardiovascular events: preventive strategies. *Circulation.* 124: 346-354.
922. Aronow, W. S. and Cassidy, J. (1974). Effect of marihuana and placebo-marihuana smoking on angina pectoris. *N.Engl.J.Med.* 291: 65-67.
923. Sidney, S. (2002). Cardiovascular consequences of marijuana use. *J.Clin.Pharmacol.* 42: 64S-70S.
924. Fisher, B. A., Ghuran, A., Vadamalai, V., and Antonios, T. F. (2005). Cardiovascular complications induced by cannabis smoking: a case report and review of the literature. *Emerg.Med.J.* 22: 679-680.
925. Chesher, G. and Hall, W. Effects of cannabis on the cardiovascular and gastrointestinal systems. The health effects of cannabis. Kalant, H., Corrigall, W., Hall, W., and Smart, R. Toronto: Centre of Addiction and Mental Health, 1999.

926. Merritt, J. C., Cook, C. E., and Davis, K. H. (1982). Orthostatic hypotension after delta 9-tetrahydrocannabinol marijuana inhalation. *Ophthalmic Res.* 14: 124-128.
927. Purnell, J. Q., Zambon, A., Knopp, R. H., Pizzuti, D. J. and others. (2000). Effect of ritonavir on lipids and post-heparin lipase activities in normal subjects. *AIDS.* 14: 51-57.
928. Cottencin, O., Karila, L., Lambert, M., Arveiller, C. and others. (2010). Cannabis arteritis: review of the literature. *J.Addict.Med.* 4: 191-196.
929. Noel, B., Ruf, J., and Panizzon, R. G. (2008). Cannabis arteritis. *J.Am.Acad.Dermatol.* 58: S65-S67.
930. Combemale, P., Consort, T., Denis-Thelis, L., Estival, J. L. and others. (2005). Cannabis arteritis. *Br.J.Dermatol.* 152: 166-169.
931. Disdier, P., Granel, B., Serratrice, J., Constans, J. and others. (2001). Cannabis arteritis revisited--ten new case reports. *Angiology.* 52: 1-5.
932. Wolff, V., Lauer, V., Rouyer, O., Sellal, F. and others. (2011). Cannabis use, ischemic stroke, and multifocal intracranial vasoconstriction: a prospective study in 48 consecutive young patients. *Stroke.* 42: 1778-1780.
933. Cox, B., Chhabra, A., Adler, M., Simmons, J. and others. (2012). Cannabinoid hyperemesis syndrome: case report of a paradoxical reaction with heavy marijuana use. *Case.Report.Med.* 2012: 757696-
934. Batkai, S., Jarai, Z., Wagner, J. A., Goparaju, S. K. and others. (2001). Endocannabinoids acting at vascular CB1 receptors mediate the vasodilated state in advanced liver cirrhosis. *Nat.Med.* 7: 827-832.
935. Fernandez-Rodriguez, C. M., Romero, J., Petros, T. J., Bradshaw, H. and others. (2004). Circulating endogenous cannabinoid anandamide and portal, systemic and renal hemodynamics in cirrhosis. *Liver Int.* 24: 477-483.
936. Julien, B., Grenard, P., Teixeira-Clerc, F., Van Nhieu, J. T. and others. (2005). Antifibrogenic role of the cannabinoid receptor CB2 in the liver. *Gastroenterology.* 128: 742-755.
937. Teixeira-Clerc, F., Julien, B., Grenard, P., Tran Van, Nhieu J. and others. (2006). CB1 cannabinoid receptor antagonism: a new strategy for the treatment of liver fibrosis. *Nat.Med.* 12: 671-676.
938. Siegmund, S. V. and Schwabe, R. F. (2008). Endocannabinoids and liver disease. II. Endocannabinoids in the pathogenesis and treatment of liver fibrosis. *Am.J.Physiol Gastrointest.Liver Physiol.* 294: G357-G362.
939. Mallat, A., Hezode, C., and Lotersztajn, S. (2008). Environmental factors as disease accelerators during chronic hepatitis C. *J.Hepatol.* 48: 657-665.
940. Guy, G. W. and Stott, C. G. The development of Sativex®-- a natural cannabis-based medicine. *Cannabinoids as Therapeutics.* Mechoulam, R. Basel: Birkhäuser Verlag, 2005.
941. Bolla, K. I., Brown, K., Eldreth, D., Tate, K. and others. (2002). Dose-related neurocognitive effects of marijuana use. *Neurology.* 59: 1337-1343.
942. Rubino, T. and Parolaro, D. (2008). Long lasting consequences of cannabis exposure in adolescence. *Mol.Cell Endocrinol.* 286: S108-S113.
943. Heishman, S. J., Stitzer, M. L., and Yingling, J. E. (1989). Effects of tetrahydrocannabinol content on marijuana smoking behavior, subjective reports, and performance. *Pharmacol.Biochem.Behav.* 34: 173-179.
944. Wetzell, C. D., Janowsky, D. S., and Clopton, P. L. (1982). Remote memory during marijuana intoxication. *Psychopharmacology (Berl).* 76: 278-281.

945. Fletcher, J. M., Page, J. B., Francis, D. J., Copeland, K. and others. (1996). Cognitive correlates of long-term cannabis use in Costa Rican men. *Arch.Gen Psychiatry*. 53: 1051-1057.
946. Pope, H. G., Jr., Gruber, A. J., Hudson, J. I., Cohane, G. and others. (2003). Early-onset cannabis use and cognitive deficits: what is the nature of the association? *Drug Alcohol Depend*. 69: 303-310.
947. Messinis, L., Kyprianidou, A., Malefaki, S., and Papathanasopoulos, P. (2006). Neuropsychological deficits in long-term frequent cannabis users. *Neurology*. 66: 737-739.
948. Lyketsos, C. G., Garrett, E., Liang, K. Y., and Anthony, J. C. (1999). Cannabis use and cognitive decline in persons under 65 years of age. *Am.J.Epidemiol*. 149: 794-800.
949. Pope, H. G., Jr., Gruber, A. J., Hudson, J. I., Huestis, M. A. and others. (2001). Neuropsychological performance in long-term cannabis users. *Arch.Gen Psychiatry*. 58: 909-915.
950. Pope, H. G., Jr., Gruber, A. J., and Yurgelun-Todd, D. (1995). The residual neuropsychological effects of cannabis: the current status of research. *Drug Alcohol Depend*. 38: 25-34.
951. Gonzalez, R., Carey, C., and Grant, I. (2002). Nonacute (residual) neuropsychological effects of cannabis use: a qualitative analysis and systematic review. *J.Clin.Pharmacol*. 42: 48S-57S.
952. Meier, M. H., Caspi, A., Ambler, A., Harrington, H. and others. (2012). Persistent cannabis users show neuropsychological decline from childhood to midlife. *Proc.Natl.Acad.Sci.U.S.A*. 109: E2657-E2664.
953. Menetrey, A., Augsburger, M., Favrat, B., Pin, M. A. and others. (2005). Assessment of driving capability through the use of clinical and psychomotor tests in relation to blood cannabinoids levels following oral administration of 20 mg dronabinol or of a cannabis decoction made with 20 or 60 mg Delta9-THC. *J.Anal.Toxicol*. 29: 327-338.
954. Asbridge, M., Poulin, C., and Donato, A. (2005). Motor vehicle collision risk and driving under the influence of cannabis: evidence from adolescents in Atlantic Canada. *Accid.Anal.Prev*. 37: 1025-1034.
955. Gieringer, D. H. (1988). Marijuana, driving, and accident safety. *J.Psychoactive Drugs*. 20: 93-101.
956. Sexton, B. F., Tunbridge, R. J., Brook-Carter, N., Jackson, P. G. and others. The influence of cannabis on driving. (2007). 477. Berkshire, UK: TRL Limited.
957. Moskowitz, H. (1985). Marijuana and driving. *Accid.Anal.Prev*. 17: 323-345.
958. Bosker, W. M., Kuypers, K. P., Theunissen, E. L., Surinx, A. and others. (2012). Medicinal Delta(9) - tetrahydrocannabinol (dronabinol) impairs on-the-road driving performance of occasional and heavy cannabis users but is not detected in Standard Field Sobriety Tests. *Addiction*. 107: 1837-1844.
959. Kuypers, K. P., Legrand, S. A., Ramaekers, J. G., and Verstraete, A. G. (2012). A case-control study estimating accident risk for alcohol, medicines and illegal drugs. *PLoS.One*. 7: e43496-
960. Laumon, B., Gadegbeku, B., Martin, J. L., and Biecheler, M. B. (2005). Cannabis intoxication and fatal road crashes in France: population based case-control study. *BMJ*. 331: 1371-
961. Khiabani, H. Z., Bramness, J. G., Bjerneboe, A., and Morland, J. (2006). Relationship between THC concentration in blood and impairment in apprehended drivers. *Traffic.Inj.Prev*. 7: 111-116.
962. Thornicroft, G. (1990). Cannabis and psychosis. Is there epidemiological evidence for an association? *Br.J.Psychiatry*. 157: 25-33.

963. Morrison, P. D., Zois, V., McKeown, D. A., Lee, T. D. and others. (2009). The acute effects of synthetic intravenous Delta9-tetrahydrocannabinol on psychosis, mood and cognitive functioning. *Psychol.Med.* 39: 1607-1616.
964. Kotin, J., Post, R. M., and Goodwin, F. K. (1973). 9-Tetrahydrocannabinol in depressed patients. *Arch.Gen.Psychiatry.* 28: 345-348.
965. Ablon, S. L. and Goodwin, F. K. (1974). High frequency of dysphoric reactions to tetrahydrocannabinol among depressed patients. *Am.J.Psychiatry.* 131: 448-453.
966. Glass, R. M., Uhlenhuth, E. H., Hartel, F. W., Schuster, C. R. and others. (1980). A single dose study of nabilone, a synthetic cannabinoid. *Psychopharmacology (Berl).* 71: 137-142.
967. Glass, R. M., Uhlenhuth, E. H., Hartel, F. W., Schuster, C. R. and others. (1981). Single-dose study of nabilone in anxious volunteers. *J.Clin.Pharmacol.* 21: 383S-396S.
968. Degenhardt, L., Hall, W., and Lynskey, M. (2003). Exploring the association between cannabis use and depression. *Addiction.* 98: 1493-1504.
969. Harder, V. S., Morral, A. R., and Arkes, J. (2006). Marijuana use and depression among adults: Testing for causal associations. *Addiction.* 101: 1463-1472.
970. Harder, V. S., Stuart, E. A., and Anthony, J. C. (2008). Adolescent cannabis problems and young adult depression: male-female stratified propensity score analyses. *Am.J.Epidemiol.* 168: 592-601.
971. Stinson, F. S., Ruan, W. J., Pickering, R., and Grant, B. F. (2006). Cannabis use disorders in the USA: prevalence, correlates and co-morbidity. *Psychol.Med.* 36: 1447-1460.
972. van Laar, M., van Dorsselaer, S., Monshouwer, K., and de Graaf, R. (2007). Does cannabis use predict the first incidence of mood and anxiety disorders in the adult population? *Addiction.* 102: 1251-1260.
973. Chabrol, H., Chauchard, E., and Girabet, J. (2008). Cannabis use and suicidal behaviours in high-school students. *Addict.Behav.* 33: 152-155.
974. Pedersen, W. (2008). Does cannabis use lead to depression and suicidal behaviours? A population-based longitudinal study. *Acta Psychiatr.Scand.* 118: 395-403.
975. Henquet, C., Krabbendam, L., de Graaf R., ten Have M. and others. (2006). Cannabis use and expression of mania in the general population. *J.Affect.Disord.* 95: 103-110.
976. Strakowski, S. M., DelBello, M. P., Fleck, D. E., Adler, C. M. and others. (2007). Effects of co-occurring cannabis use disorders on the course of bipolar disorder after a first hospitalization for mania. *Arch.Gen.Psychiatry.* 64: 57-64.
977. Agrawal, A., Nurnberger, J. I., Jr., and Lynskey, M. T. (2011). Cannabis involvement in individuals with bipolar disorder. *Psychiatry Res.* 185: 459-461.
978. De Pradier, M., Gorwood, P., Beaufils, B., Ades, J. and others. (2010). Influence of the serotonin transporter gene polymorphism, cannabis and childhood sexual abuse on phenotype of bipolar disorder: a preliminary study. *Eur.Psychiatry.* 25: 323-327.
979. Baethge, C., Hennen, J., Khalsa, H. M., Salvatore, P. and others. (2008). Sequencing of substance use and affective morbidity in 166 first-episode bipolar I disorder patients. *Bipolar.Disord.* 10: 738-741.
980. Martinez-Gras, I., Hoenicka, J., Ponce, G., Rodriguez-Jimenez, R. and others. (2006). (AAT)n repeat in the cannabinoid receptor gene, CNR1: association with schizophrenia in a Spanish population. *Eur.Arch.Psychiatry Clin.Neurosci.* 256: 437-441.

981. Monteleone, P., Bifulco, M., Maina, G., Tortorella, A. and others. (2010). Investigation of CNRI and FAAH endocannabinoid gene polymorphisms in bipolar disorder and major depression. *Pharmacol.Res.* 61: 400-404.
982. Lagerberg, T. V., Sundet, K., Aminoff, S. R., Berg, A. O. and others. (2011). Excessive cannabis use is associated with earlier age at onset in bipolar disorder. *Eur.Arch.Psychiatry Clin.Neurosci.* 261: 397-405.
983. Braga, R. J., Burdick, K. E., Derosse, P., and Malhotra, A. K. (2012). Cognitive and clinical outcomes associated with cannabis use in patients with bipolar I disorder. *Psychiatry Res.* 200: 242-245.
984. Allebeck, P. Cannabis and psychiatric syndrome. *Marihuana and medicine.* Nahas, C. G., Sutin, K. M., Harvey, D. J., and Agurell, S. Totowa, New Jersey: Humana Press, 1999.
985. Caspari, D. (1999). Cannabis and schizophrenia: results of a follow-up study. *Eur.Arch.Psychiatry Clin.Neurosci.* 249: 45-49.
986. Zammit, S., Allebeck, P., Andreasson, S., Lundberg, I. and others. (2002). Self reported cannabis use as a risk factor for schizophrenia in Swedish conscripts of 1969: historical cohort study. *BMJ.* 325: 1199-1203.
987. Arseneault, L., Cannon, M., Poulton, R., Murray, R. and others. (2002). Cannabis use in adolescence and risk for adult psychosis: longitudinal prospective study. *BMJ.* 325: 1212-1213.
988. Arseneault, L., Cannon, M., Witton, J., and Murray, R. M. (2004). Causal association between cannabis and psychosis: examination of the evidence. *Br.J.Psychiatry.* 184: 110-117.
989. Decoster, J., van, Os J., Kenis, G., Henquet, C. and others. (2011). Age at onset of psychotic disorder: cannabis, BDNF Val66Met, and sex-specific models of gene-environment interaction. *Am.J.Med.Genet.B Neuropsychiatr.Genet.* 156B: 363-369.
990. Tunbridge, E. M., Harrison, P. J., and Weinberger, D. R. (2006). Catechol-o-methyltransferase, cognition, and psychosis: Val158Met and beyond. *Biol.Psychiatry.* 60: 141-151.
991. McIntosh, A. M., Baig, B. J., Hail, J., Job, D. and others. (2007). Relationship of catechol-O-methyltransferase variants to brain structure and function in a population at high risk of psychosis. *Biol.Psychiatry.* 61: 1127-1134.
992. Miyake, N., Thompson, J., Skinbjerg, M., and Abi-Dargham, A. (2011). Presynaptic dopamine in schizophrenia. *CNS.Neurosci.Ther.* 17: 104-109.
993. Fan, J. B., Zhang, C. S., Gu, N. F., Li, X. W. and others. (2005). Catechol-O-methyltransferase gene Val/Met functional polymorphism and risk of schizophrenia: a large-scale association study plus meta-analysis. *Biol.Psychiatry.* 57: 139-144.
994. Ayalew, M., Le-Niculescu, H., Levey, D. F., Jain, N. and others. (2012). Convergent functional genomics of schizophrenia: from comprehensive understanding to genetic risk prediction. *Mol.Psychiatry.* 17: 887-905.
995. D'Souza, D. C., Sewell, R. A., and Ranganathan, M. (2009). Cannabis and psychosis/schizophrenia: human studies. *Eur.Arch.Psychiatry Clin.Neurosci.* 259: 413-431.
996. Castle, D. J. and Solowij, N. Acute and subacute psychomimetic effects of cannabis in humans. *Marijuana and madness.* Castle, D. and Murray, R. Cambridge: Cambridge University Press, 2004.
997. Fridberg, D. J., Vollmer, J. M., O'Donnell, B. F., and Skosnik, P. D. (2011). Cannabis users differ from non-users on measures of personality and schizotypy. *Psychiatry Res.* 186: 46-52.

998. Thompson, G. R., Rosenkrantz, H., Schaeppi, U. H., and Braude, M. C. (1973). Comparison of acute oral toxicity of cannabinoids in rats, dogs and monkeys. *Toxicol.Appl.Pharmacol.* 25: 363-372.
999. Robson, P. (2001). Therapeutic aspects of cannabis and cannabinoids. *Br.J.Psychiatry.* 178: 107-115.
1000. Weinstein, A. M. and Gorelick, D. A. (2011). Pharmacological treatment of cannabis dependence. *Curr.Pharm.Des.* 17: 1351-1358.